Effects of some disinfectants on *Toxocara* spp. eggs viability of dogs and cats

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Received: 28.05.2019 • Accepted/Published Online: 03.03.2020 • Final Version: 02.06.2020

Abstract: The main cause of *larva migrans syndrome* (LMS) in humans is represented by *Toxocara* spp. found in dogs and cats. In the present study, the effects of three disinfectants based on quaternary ammonium compounds (Kohrsolin® FF, Trioton® rapid AF and Terralin® protect) and one based on propanol and ethanol (Bacillol®), against *Toxocara canis* and *Toxocara cati* eggs, were tested. These disinfectants have a strong antimicrobial and virucidal action. Two different exposure times and different concentrations for solutions were used. All four disinfectants presented high percentages of destructive effects against *Toxocara canis* eggs, after 21 days of exposure. The vast majority of eggs with pronounced degenerative changes in the eggshell and embryonic development were observed using Bacillol® for both *Toxocara* species. Terralin® protect was ineffective against *Toxocara cati* eggs. The ovicidal effect of these compounds has been noticed thus allowing their use as disinfectants for surfaces contaminated with *Toxocara* spp. eggs.

Keywords: *Toxocara* spp., disinfectants, exposure, carnivores, efficiency

1. Introduction

Human toxocariasis is a zoonosis caused by the larval stage of *Toxocara canis* and *Toxocara cati* species, which are gastrointestinal nematodes infecting dogs and cats, found especially in young carnivores under one year of age [1]. People get infected by ingesting the larvated eggs presenting second stage larvae or third stage larvae, found in contaminated soil [2,3], vegetables [4–7], water or animal hair [8,9]. Poorly cooked meat from paratenic hosts (cows, sheep, chickens) is another major source of infection [10]. Children with pica are most commonly diagnosed. They infest themselves with the larvated eggs from sandpits or various playgrounds, where dog or cat faeces are found. The specific human syndromes are *larva migrans visceralis* and *larva migrans ocularis*.

The morphological aspects of *T. canis* and *T. cati* eggs are very similar. A female ascarid can lay up to 200,000 eggs per day, which are highly resistant in the external environment due to the five-layer eggshell (uterine, vitelline, chitinous, granular, and fibrous layers) [11]. To reach the infective stage, the eggs require approximately 2 to 5 weeks until the larvae forms inside at temperatures between 15 °C and 35 °C and over 85% humidity [12]. The complete development of the eggs can also occur at temperatures of −2 °C or 1 °C and they can last about 6 weeks [13].

In temperate areas, eggs can survive from 6 to 12 months and the most commonly used disinfectants are not effective against them (2% glutaraldehyde, 10% benzalkonium chloride, 1% potassium permanganate, 70% ethyl alcohol, 10% potassium hydroxide and 3% phenol) [11]. Instead, products based on iodine and ethanol can be successfully used against *T. canis* eggs [11,12]. Contradictory results have been observed regarding the action of sodium hypochlorite. Some authors argue its efficacy against *T. canis* eggs [12], while the others sustain the opposite [11].

In hospitals and veterinary clinics, and even in animal shelters, disinfection is a crucial step. The aim of this study was to test the effects of four disinfectants (Kohrsolin® FF, Trioton® rapid AF, Terralin® protect, Bacillol®) on the viability of *T. canis* and *T. cati* eggs. The bactericidal, fungicidal, and virucidal actions of these substances are well-known, but the ovicidal effect is not specified. These products are used intensively in human hospitals for disinfecting surfaces and medical instruments, therefore we considered studying their ovicidal action in hopes of discovering a viable disinfectant for usage in veterinary clinics and hospitals and in other related institutions that come in contact with a possible source of contamination.
2. Materials and methods

2.1. Gathering the eggs
The female ascarids were longitudinally sectioned at tegumentary level in order to highlight the uterine horns. These were separately collected and longitudinally sectioned, after which they were milled with distilled water and filtered through a 0.5 mm mesh sieve, obtaining the eggs of *T. canis* and *T. Cati* [14].

2.2. Exposure of unembryonated eggs to disinfectants
The concentrations used for each disinfectant and their chemical composition are shown schematically in Table 1. These were established according to the manufacturers of each product, choosing those concentrations most commonly used in human hospitals for short exposure times (15 and 30 min, respectively).

Approximately 10⁴ *T. canis* and *T. cati* eggs were obtained and subsequently placed in 48 glass tubes with 5 mL from each disinfectant.

For each disinfectant, 3 samples were prepared. Thereby, from the total of 48 samples, 24 were with *T. canis* eggs and 24 with *T. cati* eggs.

The eggs were then exposed for 15 min and 30 min, at room temperature (22–24 °C), as follows: from a total of 24 samples of *T. canis* eggs, 12 were exposed to the action of disinfectants for a period of 15 min and the other 12, for a period of 30 min. The same procedure was also performed for *T. cati* eggs.

After the exposure, the samples were centrifuged for 7 min, at 2500 rpm, a greater centrifugal force than the one used by Gautam et al. [15], for a more efficient and correct sedimentation. The centrifugation process and the removal of the supernatant followed by a replacement with distilled water was repeated 4 times. The eggs immersed in distilled water were subsequently kept at room temperature (22–24 °C) for 21 days, being sputtered daily in order to ensure the necessary oxygen. Periodical examinations were performed [15]. The light/darkness cycle provided throughout the experiment was 10/14 h, daily.

For each disinfectant, the standard deviation and the average of the results expressed in percentage obtained from 3 samples were calculated.

2.3. Egg classification / disinfectant efficiency
A total of 6 control samples (3 with *T. canis* eggs and 3 with *T. cati* eggs) were maintained in 5 mL of physiological serum for the same period of time.

2.4. Statistical analysis
Statistical analysis of the results was made using GraphPad Prism program, QuickCalcs, with two-tailed Fischer’s exact test to obtain the P-value. The results obtained in the samples exposed to disinfectants (the number of nonviable eggs) were compared with those from the control group.

The percentages of *T. canis* nonviable eggs were higher in samples exposed to disinfectants for 15 and 30 min.

### Table 1. Chemical composition and concentration of disinfectants.

<table>
<thead>
<tr>
<th>Disinfectant</th>
<th>Chemical composition</th>
<th>Solution</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kohrsolin FF (BODE Chemie GmbH, Hamburg, Germany)</td>
<td>Synergistic combination of glutaral and quaternary ammonium salts</td>
<td>1%</td>
</tr>
<tr>
<td>Terralin protect (Schülke &amp; Mayr GmbH, Norderstedt, Germany)</td>
<td>A combination of quaternary ammonium compounds, aromatic alcohols, glycerin amphiphilic derivatives and nonionic surfactants</td>
<td>2%</td>
</tr>
<tr>
<td>Trioton rapid AF (LC Pliwa Ljubomir Cugurovic e.Kfm, Malsfeld, Germany)</td>
<td>Quaternary ammonium compounds and amines</td>
<td>0.25%</td>
</tr>
<tr>
<td>Bacillol (BODE Chemie GmbH, Hamburg, Germany)</td>
<td>Propan-1-ol, propan-2-ol, ethanol</td>
<td>1%</td>
</tr>
</tbody>
</table>
compared to the control group with extremely significant differences ($P < 0.0001$) after 21 days of exposure. Only Terralin® protect was ineffective against *T. cati* eggs, the percentages of nonviable eggs being similar to those found in control samples ($P > 0.005$), after 21 days.

The most effective disinfectant against *T. canis* eggs was Bacillol® 1%, regardless of the exposure time, followed by Terralin® protect 2%, Kohrsolin® FF 1% and Trioton® rapid AF 0.25% solutions. No significant differences were identified between the individual actions of disinfectants based on quaternary ammonium compounds ($P > 0.05$). Instead, extremely significant differences were noticed regarding the individual action of Bacillol® compared to the other products ($P < 0.001$).

After 21 days, the percentages of *T. cati* nonviable eggs were also higher in samples exposed to Kohrsolin® FF, Trioton® rapid AF and Bacillol® compared to the control group regardless of the exposure time, with extremely significant differences ($P < 0.001$). No significant differences were identified regarding the action of Terralin® protect solution compared to control values ($P > 0.05$).

Regarding the action of disinfectants against *T. cati* eggs, after a 15-min exposure, the percentages of nonviable eggs observed in samples exposed to Bacillol® compared to those identified in samples exposed to Kohrsolin® were similar ($P > 0.05$). Higher percentages of nonviable eggs have been noted in samples exposed to Bacillol® compared to those observed in samples exposed to Trioton® rapid AF,
the differences being very significant ($P < 0.01$). Significant differences ($P < 0.05$) were observed by comparing the individual action of Kohrsolin® to Trioton®’s rapid AF. After a 30-min exposure, Kohrsolin® FF, Trioton® rapid AF and Bacillol® determined similar percentages of nonviable eggs, the differences between their action being insignificant ($P > 0.05$).

Considering the action of the same disinfectant both on *T. canis* eggs and on *T. cati* eggs, extremely significant differences ($P < 0.0001$) regarding the effect of Kohrsolin® FF, Terralin® protect and Trioton® rapid AF, exposed for 15 or 30 min, were observed. Regarding Bacillol®’s action, significant differences ($P < 0.05$), after 15-min exposure and insignificant differences ($P > 0.05$), after 30-min exposure were identified.

### 4. Discussion

In a similar study, Laciak et al.[16] tested a disinfectant based on quaternary ammonium compounds against *T. canis* eggs. Saniten® product was used in a 10% concentration, a higher concentration compared to 0.25% (Trioton® rapid AF), 1% (Kohrsolin® FF) and 2% (Terralin® protect) used in the current study. *T. canis* eggs had been exposed for a longer time to chemicals (60, 90, and 180 min, respectively). The percentages of nonviable eggs observed at 21 days in the samples exposed to disinfectant were 56.32% (60 min), 63.07% (90 min), and 56.01% (180 min). In the current study, similar results were obtained for *T. canis* nonviable eggs examined after the same period of time. Higher percentages of *T. cati* nonviable eggs in samples exposed to Kohrsolin®FF and Trioton® rapid AF were obtained. This suggests a greater sensitivity of *T. cati* eggs to these disinfectants, except Terralin® protect. The most effective disinfectant, identified in the study conducted by Laciak et al. [16], was H1 100% that contains

hydrogen peroxide, tenside and sodium hydroxide, with 94.18% (60 min), 97.96% (90 min), and 98.04% (180 min) nonviable eggs.

Short exposure times were used in this current study in order to mimic the real-time conditions from veterinary clinics and hospitals, where a surface disinfection is performed after each patient and the period between two consultations is 30 min maximum. Perhaps a longer exposure of *T. canis* and *T. cati* eggs to disinfectants would have led to an increase in percentage of nonviable eggs.

Since Terralin® protect contains the same basic ingredients (quaternary ammonium salts) as the other two disinfectants and the conditions of egg exposure and storage were identical, further studies are required to identify the mechanism of action of these solutions, especially on *T. cati* eggs.

Other researchers revealed that the hydrogen peroxide 3% solution has an inhibitory activity on 99.73% of *T. canis* unembryonated eggs after a 24-h exposure [17].

Von Dohlen et al. [18] looked at the effect of sodium hypochlorite 5.25% solution on *T. canis* eggs. The results obtained have shown that this disinfectant is ineffective against *T. canis* eggs and that they can later develop into the infestation stage. Higher concentrations, such as 7% sodium hypochlorite solution were ineffective against *T. canis* eggs [11]. Instead, Verocai et al. [19] showed that 2%–2.5% sodium hypochlorite and 70% ethanol solutions negatively influenced the embryogenesis of *T. canis* eggs, which also caused changes in their integrity. Similar results were obtained by Morrondo et al., indicating the possibility of using sodium hypochlorite solution against *T. canis* eggs [12].

El-Sayed observed that *Zingiber officinale* ethanol extract, with 100 mg/mL concentration has ovicidal activity on *T. canis* eggs with an efficiency of 98.2% after

### Table 2. Effect of disinfectants on *Toxocara spp.* after 21 days.

<table>
<thead>
<tr>
<th>Disinfectant</th>
<th><em>Toxocaracanis</em></th>
<th><em>Toxocaracati</em></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Exposure time</td>
<td>Exposure time</td>
</tr>
<tr>
<td></td>
<td>15 min</td>
<td>30 min</td>
</tr>
<tr>
<td>Mean± SD [1]</td>
<td>Mean ± SD</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Kohrsolin FF</td>
<td>65.33 ± 0.57</td>
<td>66.66 ± 0.57</td>
</tr>
<tr>
<td>Terralin protect</td>
<td>65.66 ± 1.15</td>
<td>69.66 ± 1.52</td>
</tr>
<tr>
<td>Trioton rapid AF</td>
<td>57.66 ± 1.52</td>
<td>68.66 ± 1.52</td>
</tr>
<tr>
<td>Bacillol</td>
<td>87.66 ± 1.52</td>
<td>93.66 ± 1.15</td>
</tr>
<tr>
<td>Control</td>
<td>27.33 ± 2.51</td>
<td>23.33 ± 1.15</td>
</tr>
</tbody>
</table>

[1] = the mean value was obtained by summing up the nonviable eggs in the three samples and dividing them by three, [2] = standard deviation.
24 hours. The efficacy was 59.22% and 82.5%, respectively, after 24 h, in case of 25, 50 mg/mL concentrations usage [20].

In a recent study, El-Dakhly et al. [21] tested the effects of 6 commercial disinfectants on Toxascarisleonina embryo genesis and larval development. Only 4 of them caused a significant reduction of larval development (P ≤ 0.05): 58.8% (TH4 - glutaraldehyde), 85.8% (70% ethanol) and 100% (Dettol® - chloroxylenol and Virkon® - potassium peroxymonosulfate, sodium chloride). Sodium hypochlorite and phenol have led to an insignificant reduction of larval development (2.8% and 21.0%, respectively).

In conclusion, these solutions based on quaternary ammonium compounds, propanol and ethanol have ovicidal action and can be used for disinfection of surfaces contaminated with Toxocarapp. eggs, except Terralin® protect which was ineffective against Toxocaracati eggs. Further studies are needed to identify the chemical and pathophysiological mechanism of action of these disinfectants on ascarid egg (especially T.cati eggs) and the morphological changes that occur.

**Conflict of interest**
The authors have no conflicts of interest to disclose.

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