

Emine ALP¹
Mehmet YERER¹
Duygu ESEL²
Gökhan METAN¹
Mehmet DOĞANAY¹

Risk factors for acquisition of methicillin-resistant *Staphylococcus aureus* and clonal spread of the isolates in a medical intensive care unit*

Aim: Methicillin-resistant *Staphylococcus aureus* (MRSA) is still the commonest pathogen in hospital-acquired infections with high morbidity and mortality. MRSA colonization usually precedes infection and dissemination of the microorganism. The aim of this study was to determine risk factors for the colonization and infection with MRSA in a medical intensive care unit (MICU) and to show the genetic relation of strains.

Materials and methods: This study was conducted prospectively between 1 December 2004 and 31 January 2006 in MICU. Patients (>16 years) admitted to the MICU were screened for MRSA on admission (in the first 48 hours), at the end of the first week, and at the ICU discharge using anterior nares, axilla, and groin swabs. Risk factors for colonization and infection of MRSA were evaluated. Strains isolated from colonized patients were evaluated for genetic relation.

Results: During the study period, 259 patients were evaluated for the risk factors of MRSA acquisition. The colonization rate was 18.5%, and 64.6% of the patients were colonized in the first week. In multiple logistic regression analysis, only the length of stay in MICU and mechanical ventilation were significant risk factors for colonization. MRSA infection occurred in 15 (31%) of 48 colonized patients during MICU stay. MRSA colonization and tracheostomy were significant risk factors for MRSA infection. A genetic relation was found in 48 isolates from colonized patients in MICU and 23 isolates from colonized patients on admission. Overall, 4 clones (clone A, B, C, and D) were determined from colonized patients on admission; clone A (37%), clone B (32%), clone C (21%), and clone D (10%). Moreover, patients colonized during MICU stay had similar clones (clone A-30%, clone B-57%, and clone C-13%) with these patients.

Conclusion: This study shows a high colonization rate and dissemination of MRSA in a developing country with inadequate infrastructure (lack of nurse, lack of isolation rooms, and heavy workload).

Key words: *Staphylococcus aureus*, intensive care unit, colonization, infection control

Bir dahiliye yoğun bakım ünitesinde metisiline dirençli *Staphylococcus aureus* kazanımı için risk faktörleri ve izolatların klonal yayılımı

Amaç: Metisiline dirençli *Staphylococcus aureus* (MDSA) hastane kaynaklı enfeksiyonlarda en sık görülen ve yüksek morbidite ve mortaliteye sahip patojenlerden biridir. MDSA kolonizasyonu, genellikle enfeksiyona ve mikroorganizmanın yayılımına öncülük eder. Bu çalışmanın amacı bir dahiliye yoğun bakım ünitesi (DYBÜ)'nde MDSA kolonizasyonu ve enfeksiyonu için risk faktörlerini tespit etmek ve suşların genetik yakınlığını göstermekti.

Yöntem ve gereç: Bu çalışma prospektif olarak 1 Aralık 2004 ve 31 Ocak 2006 tarihleri arasında yapıldı. DYBÜ'ye yatan hastalar (>16 yaş) ön burun kanatları, aksilla ve kasık sürüntü örnekleri ile MDSA varlığı açısından yatışta (ilk 48 saat), birinci hafta ve YBÜ'den çıkarken araştırıldı. MDSA kolonizasyonu ve enfeksiyonu için risk faktörleri analiz edildi. Kolonize hastalardan izole edilen suşların genetik yakınlığı araştırıldı.

* This study was supported by Erciyes University Research Foundation and The Scientific and Technical Research Council of Turkey (project no: 105S482).

¹ Department of Infectious Diseases,
Faculty of Medicine,
Erciyes University,
Kayseri - TURKEY

² Department of Microbiology,
Faculty of Medicine,
Erciyes University,
Kayseri - TURKEY

Received: January 25, 2008
Accepted: July 09, 2009

Correspondence

Emine ALP
Department of Infectious Disease,
Faculty of Medicine,
Erciyes University,
38039 Kayseri - TURKEY
ealp@erciyes.edu.tr

Bulgular: Çalışma döneminde, 259 hasta MDSA kazanımı için risk faktörleri açısından değerlendirildi. Kolonizasyon oranı % 18.5 idi ve hastaların % 64.6'sı ilk haftada kolonize oldu. Çok değişkenli lojistik regresyon analizinde, sadece DYBÜ'de yatış süresi ve mekanik ventilasyon kolonizasyon için önemli risk faktörleri olarak bulundu. DYBÜ'sinde yatış sırasında kolonize olan 48 hastanın 15 (% 31)'inde MDSA enfeksiyonu gelişti. MDSA kolonizasyonu ve trakeostomi MDSA enfeksiyonu için önemli risk faktörleri olarak bulundu. DYBÜ'de kolonize olan hastalardan izole edilen 48 izolat ve yatışta kolonize hastalardan izole edilen 23 izolat arasında genetik ilişki bulundu. Yatışta kolonize olan hastalardan toplam olarak dört klon (klon A, B, C, D) tespit edildi; klon A (% 37), klon B (% 32), klon C (% 21) ve klon D (% 10). Ayrıca DYBÜ'sinde kolonize hastalardan izole edilen suşlarda benzer klonlara sahipti (klon A-% 30, klon B-% 57, klon C-% 13).

Sonuç: Bu çalışma yetersiz alt yapıya sahip (kısıtlı hemşire, izolasyon odası yokluğu, ağır iş yükü) gelişmekte olan bir ülkede, MDSA ile yüksek kolonizasyon oranını ve yayılımını göstermektedir.

Anahtar sözcükler: *Staphylococcus aureus*, yoğun bakım ünitesi, kolonizasyon, enfeksiyon kontrolü

Introduction

Nosocomial infections constitute an important health-care problem in ICU patients with high mortality, morbidity, and high cost. Methicillin-resistant *Staphylococcus aureus* (MRSA) is the leading cause of nosocomial infections, and they are usually associated with poor outcomes in ICUs (1). Furthermore, ICU-acquired infections are a serious problem in Turkey and the incidence was reported as 56 in 1000-patient days (2). In a one-day point prevalence study, a total of 56 ICUs from 22 university and teaching hospitals participated and 49% of patients had one or more ICU-acquired infections (3). Pneumonia and bacteremia are the most frequent nosocomial infections in Turkish ICUs and MRSA is one of the important cause of these infections with high mortality (3,4). MRSA infection is generally preceded by colonization and patients usually acquire MRSA during their hospital stay. Furthermore, colonization plays a major role in the dissemination of this organism (5).

Erciyes University Hospital is a 1300-bed tertiary-referral teaching hospital with 9 ICUs and approximately 250-875 patients admitted to each ICU annually. In the ICUs, the bed-to-bed space is very close (<1 m) and there is insufficient number of healthcare-workers (HCWs). Despite the efforts for infection control, nosocomial infection rates are still high (20/1000 patient days) in ICUs (surveillance report of hospital) and these infections are usually due to multi-drug resistant pathogens. *S.aureus* infections constitute 13% of ICU-acquired infections with excess

morbidity and mortality and 90%-95% of *S. aureus* isolates are resistant to methicillin (6-8). Probably, patients get colonized during their stay in our ICUs, and cross-transmission occurs from patient to patient because of inadequate infrastructure. This study was planned to determine the risk factors for the colonization and infection with MRSA in our medical ICU (MICU) and examine the genetic relation of the strains to show the dissemination of isolates.

Patients and methods

MICU design and infection control measures

This study was conducted prospectively between 1 December 2004 and 31 January 2006 in our MICU. There are 9 beds in the MICU and approximately 750 patients are admitted per year. Patient-based surveillance is performed actively and daily by an infection control nurse. Infection control physician visits the MICU twice a week and there are reminder posters about infection control measures and hand hygiene on the walls. Also there is a routine infection control education program for all health-care personnel twice a year. Standard infection control measures are used for all patients. Due to MRSA endemicity, all patients in ICUs have a daily bath with chlorhexidine. However, there is no isolation room for colonized patients and HCWs try to apply other contact measures (hand hygiene, use of gowns and gloves, etc.) for MRSA colonized or infected patients. Patient/nurse ratio is 4/1 in a shift. There are 3 washbasins for 9 beds and alcohol-based solutions are placed at the bedsides of all patients.

Patients

Patients (>16 years) admitted to the MICU were screened for MRSA on admission (in the first 48 hours), at the end of first week, and at ICU discharge using anterior nares, axilla, and groin swabs. Patients who stayed shorter than 48 h and already had MRSA colonization or infection were not evaluated for risk factors. Patients colonized only with MRSA were included into the study. Furthermore, HCWs who were close contact with these patients were screened for nasal colonization at the beginning and end of their ward rotation.

Data collected for risk factors included age, sex, APACHE II score, SOFA score, underlying diseases (diabetes mellitus, hypertension, cardiac disease, chronic obstructive lung disease, chronic renal failure, cirrhosis, cerebrovascular disease, active neoplastic disease, trauma, neutropenia, and immunosuppressive therapy), length of stay (LOS) in MICU before colonization, previous antibiotic use, length of antibiotic therapy before colonization, catheterization (peripheral, central venous, arterial, Swann-Ganz, and urinary), administration of total parenteral nutrition (TPN), intubation, mechanical ventilation, application of tracheostomy, presence of nasogastric tube, presence of prosthesis, prior surgery, presence of an open wound, and decubitus ulcer.

This study was approved by the local ethical committee of Erciyes University (Approval date: 04 May 2004; Number: 132).

Microbiological studies

Microorganism identification

MRSA isolates were preliminarily identified at the Bacteriology Laboratory of Erciyes University Hospital. Swabs were processed using selective mannitol salt agar with 6 mg/L of oxacillin for 24 h at 37 °C. Methicillin-resistant colonies were subcultured on 5% sheep blood agar for 18 h at 37 °C and underwent a rapid slide agglutination test for coagulase and protein A (Staphaurex; Plus).

All catalase positive gram positive isolates from clinical samples were tested for methicillin resistance by oxacillin (1 µg, Oxoid, UK) disk diffusion test. The rapid slide agglutination test for coagulase and protein A (Staphaurex; Plus) were performed for species identification. The staphylococci identified as MRSA were stored at -80 °C in microbanks until analysis.

Molecular typing

Strains isolated from colonized patients (colonized on admission and colonized in MICU) were evaluated for genetic relations. Genotyping analysis was performed by pulsed-field gel electrophoresis (PFGE) at the Molecular Laboratory of Infectious Diseases Unit, Hacettepe University, Ankara, Turkey.

First, isolates were subcultured to blood agar to ensure viability and purity. MRSA DNA embedded in agarose blocks was prepared as described by Lencastre et al. (9). The DNA fixed in the agarose disk was incubated for 20 h in 45 µL of restriction buffer and Sma I (20 U). The reaction was stopped with 5 µL of loading buffer. Gels with 1.1% agarose (Genexis Spechbach, Germany) were prepared in 0.5 × TBE buffer (50 mM boric acid, 0.2mM EDTA). PFGE was carried out by General Navigator, Pharmacia (Uppsala, Sweden). The running conditions were performed for 22 h at 14 °C at 150 V. Pulse times were 20 s for 15 h, 35 s for 7 h, 50 s for 15 h, and 90 s for the last 3 h. PFGE patterns were assigned to the same clone and clonal variant. Strains that differed by up to (and including) 6 bands were considered to belong to different clonal variants; strains that differed by more than 6 bands were considered to belong to different clones. Clones and clonal variants were designated by letters and by numbers in suffix, respectively (10).

Definitions

MRSA colonization: If any of the following cultures, anterior nares, axilla, or groin, was positive for MRSA, the patient was accepted as colonized. If the admission culture was positive for MRSA, it was labeled as “colonized on admission”. On the other hand, if the admission culture was negative for MRSA and any of the following cultures, anterior nares, axilla or groin, was positive for MRSA during ICU stay, it was defined as “colonized during MICU stay”.

MRSA infection: The diagnosis of infections was based on the criteria of the Centers for Disease Control and Prevention (CDC) (11). If the patient had the infection signs and symptoms with the yield of MRSA in culture (blood, sputum, urine, operation site, etc.), MRSA infection diagnosis was placed. For the diagnosis of ventilator associated pneumonia (VAP), quantitative cultures (the cut off value was $>10^5$) of endotracheal aspirates were used.

Statistical analysis

Data were given as mean \pm SD or median (minimum-maximum) and a P value of <0.05 was accepted as significant. Continuous variables were compared using Student's t test or the Mann-Whitney U test, when appropriate. Chi-square test was used for categorical variables. Univariate and multiple logistic regression analysis (forward stepwise) was performed for significant risk factors of MRSA acquisition. Variable with $P < 0.10$ value in the univariate logistic regression analysis included in the multiple logistic regression analysis. All analyses were carried out using SPSS 13.0.

Results

During the study period, 408 patients were admitted to the MICU and admission swabs were taken from all patients in the first 48 h. A total of 126 patients stayed in the MICU less than 48 hours and 23 (5.6%) patients were already colonized on admission. All these colonized patients had been transferred from other wards of the hospital.

Overall 259 patients were evaluated for risk factors of MRSA acquisition. The median age was 57.2 years with a range of 16-95 years. One hundred thirty six (52.5%) of the patients were male, whereas 123 (47.5%) were female. The median length of stay in the MICU was 10.1 days (range 3 to 111 days) and the median length of stay in the hospital was 18.3 days (range 3 to 113 days). Forty seven percent of 259 patients were transferred from other parts of the hospital to MICU.

While 137 patients stayed in the MICU for less than 7 days, 122 patients stayed longer. MRSA isolated from 28 (27.5%) of 102 swabs taken at the first-week and 20 (16.3%) (3 of these patients colonized in the first week) of 123 discharge swabs. Overall, 48 (18.5%) patients got colonized with MRSA during MICU stay and 64.6% of the patients colonized at the first week. The anterior nares were the most frequent site of MRSA colonization; 42 (87.5%) of the patients had only nasal colonization, whereas 1 patient had axilla and groin colonization, 3 patients had only axilla and 2 had only groin colonization.

MRSA infection occurred in 15 (31%) of 48 colonized patients. In 4 patients, MRSA infection developed unless colonization was determined. The majority of MRSA infections were VAP (12 of 19 patients), the others were bacteremia (6 of 19 patients) and soft-tissue infection (1 of 19 patients). The median time for the development of MRSA infection was 15.3 days (range 4 to 53 days) after admission.

Nasal carriage of MRSA was not detected in HCWs during the study period.

Risk factors for MRSA colonization

In the univariate analysis of the study, underlying diseases were not found as significant risk factors for acquisition of MRSA ($P > 0.05$) (Table 1). The significant risk factors were LOS in the MICU before colonization, previous use of antibiotic, length of antibiotic therapy before colonization, administration of total parenteral nutrition, intubation, mechanical ventilation, and application of tracheostomy (Table 2).

Table 1. Evaluation of underlying diseases for MRSA colonization in the univariate analysis.

Risk factors	MRSA colonized patients (n = 48)	MRSA non-colonized patients (n = 211)	P value*
Hypertension	16 (33.3%)	82 (38.9%)	0.584
Diabetes mellitus	12 (25%)	56 (26.5%)	0.970
Cardiac disease	8 (16.7%)	36 (17.1%)	1.000
Chronic obstructive lung disease	8 (16.7%)	37 (17.5%)	1.000
Chronic renal failure	8 (16.7%)	27 (12.8%)	0.635
Cirrhosis	3 (6.3%)	40 (19%)	0.055
Cerebrovascular disease	14 (29.2%)	42 (19.9%)	0.225
Active neoplastic disease	5 (10.4%)	33 (15.6%)	0.486
Neutropenia	0 (0.0%)	8 (3.8%)	0.358

Table 2. Risk factors for MRSA colonization in the univariate analysis.[&]

Risk factors	MRSA colonized patients (n = 48)	MRSA non-colonized patients (n = 211)	P value*
Age med (min - max)	64 (16 – 86)	64 (16 – 95)	0.502
Male (n = 136)	30 (62.5%)	106 (50.2%)	0.169
Female (n = 123)	18 (37.5%)	105 (49.8 %)	0.169
APACHE-II (mean ± SD)	23.27 ± 8.09	22.94 ± 8.38	0.802
SOFA (mean ± SD)	7.13 ± 3.05	6.95 ± 3.81	0.730
LOS in the MICU before colonization med (min -max)	15 (4 – 111)	5 (3 – 51)	0.001
Previous antibiotic use	45 (93.8%)	149 (70.6 %)	0.002
Length of antibiotic therapy med (min - max)	7 (1 – 26)	5 (1 – 14)	0.023
Peripheral venous catheter	33 (68.8%)	124 (58.8 %)	0.265
Central venous catheter	37 (77.1 %)	129 (61.1%)	0.056
Arterial catheter	6 (12.5 %)	21 (10%)	0.795
Swan-Ganz catheter	0 (0%)	2 (0.9 %)	1.000
Urinary catheter	46 (95.8%)	186 (88.2%)	0.190
TPN	38 (79.2%)	124 (58.8%)	0.013
Intubation	44 (91.7%)	102 (48.3%)	0.001
Mechanical ventilation	43 (89.6 %)	100 (47.4%)	0.001
Tracheostomy	18 (37.5 %)	13 (6.2%)	0.001
Prosthesis	2 (4.2 %)	11 (5.2%)	1.000
Operation	6 (12.5 %)	34 (16.1%)	0.686
Trauma	6 (12.5 %)	12 (5.7%)	0.114
Open wound	4 (8.3 %)	20 (9.5%)	1.000
Decubitus ulcer	4 (8.3%)	13 (6.2%)	0.529
Immunosuppressive therapy	7 (14.6%)	24 (11.4%)	0.710

[&] Mann-Whitney U test was performed for median values and Student's t-test was performed for mean ± SD values

*P < 0.05 was accepted significant

LOS: Length of stay; TPN: Total parenteral nutrition

In the univariate logistic regression analysis, LOS in the MICU before colonization, previous use of antibiotic, length of antibiotic therapy before colonization, central venous catheterization, administration of total parenteral nutrition, intubation, mechanical ventilation, and application of tracheostomy were found to be significant risk factors. However, in the multiple logistic regression analysis, only LOS in MICU before colonization and

mechanical ventilation were significant risk factors for colonization (Table 3).

Risk factors for MRSA infection

In univariate analysis of the study, underlying diseases were not found as significant risk factors for MRSA infection (P > 0.05) (Table 4). Significant risk factors in univariate analysis were LOS in MICU before colonization, previous use of antibiotic,

Table 3. Risk factors for MRSA colonization in the multiple logistic regression analysis (forward stepwise).

Risk factors	Odds ratio	95% CI	P value*
LOS in the MICU before colonization	1.083	1.040 - 1.129	0.001
Mechanical ventilation	4.186	1.359 - 12.895	0.013

*P < 0.05 was accepted as significant

Table 4. Evaluation of underlying diseases for MRSA infection in the univariate analysis.

Risk factors	MRSA colonized patients (n = 19)	MRSA non-colonized patients (n = 240)	P value*
Hypertension	7 (36.8 %)	91 (37.9%)	1.000
Diabetes mellitus	2 (10.5%)	66 (27.5%)	0.173
Cardiac disease	3 (15.8%)	41 (17.1%)	1.000
Chronic obstructive lung disease	2 (10.5%)	43 (17.9%)	0.543
Chronic renal failure	3 (15.8%)	32 (13.3%)	0.729
Cirrhosis	2 (10.5%)	41 (17.1%)	0.748
Cerebrovascular disease	6 (31.6 %)	50 (20.8%)	0.260
Active neoplastic disease	3 (15.8%)	35 (14.6%)	0.747
Neutropenia	0 (0.0%)	8 (3.3%)	1.000

administration of total parenteral nutrition, intubation, mechanical ventilation, application of tracheostomy, and MRSA colonization (Table 5).

In the univariate logistic regression analysis, LOS in the MICU before colonization, MRSA colonization, and application of tracheostomy were found as significant risk factors. However, in the multiple logistic regression analysis only MRSA colonization and application of tracheostomy were significant risk factors for infection (Table 6).

Genetic relation

A genetic relation was found in 48 isolates from colonized patients in MICU and 23 isolates from colonized patients on admission. Overall, 4 clones (clone A-D) were determined from colonized patients on admission; clone A (37%), clone B (32%), clone C (21%), and clone D (10%). Patients colonized during MICU stay had the same clones with the patients colonized on admission (clone A-30%, clone B-57%, clone C-13%).

Discussion

MRSA infections have an important effect on patient morbidity, mortality, and incremental hospital costs. Moreover, the changing epidemiology of MRSA by its movement from hospitals to the community poses a public threat. The emergence of vancomycin resistant *S. aureus* and *Enterococcus* spp. is another major reason for the control of MRSA because vancomycin use and MRSA infection are important risk factors for vancomycin resistant *S. aureus* and *Enterococcus* spp. (12,13). Consequently, control of MRSA is an important issue in many hospitals.

Colonized and infected patients are the major reservoirs and are responsible for the colonization of their environment and the other patients. In previous reports, the prevalence of MRSA colonization in ICUs has been reported as 4%-52% (14-18). Furthermore, the colonization rate was high (18.5%) in our MICU and 64.6% of the patients colonized in the first week. The anterior nares were the most common

Table 5. Risk factors for MRSA infection in the univariate analysis.[&]

Risk factors	MRSA colonized patients (n = 19)	MRSA non-colonized patients (n = 240)	P value*
Age med (min - max)	71 (19-86)	63 (16-95)	0.806
Male (n = 136)	12 (63.2%)	124 (51.7%)	0.467
Female (n = 123)	7 (36.8%)	116 (48.3 %)	0.467
APACHE-II (mean ± SD)	22.63 ± 6.52	23.03±8.45	0.899
SOFA (mean ± SD)	7 ± 2.57	7 ± 3.75	0.737
LOS in the MICU before colonization (colonization or infection?) med (min -max)	21 (8 – 111)	6 (3 – 89)	0.001
Previous antibiotic use	19 (100%)	175 (72.9 %)	0.005
Length of antibiotic therapy med (min - max)	6 (2 – 14)	7 (1 – 26)	0.773
Peripheral venous catheter	14 (73.7%)	143 (59.6 %)	0.334
Central venous catheter	16 (84.2 %)	150 (62.5 %)	0.099
Arterial catheter	3 (15.8%)	24 (10 %)	0.430
Swan-Ganz catheter	0 (0 %)	2 (0.8 %)	1.000
Nasogastric tube	7 (36.8 %)	125 (52.1%)	0.298
Urinary catheter	18 (94.7%)	214 (89.2%)	0.703
TPN	17 (89.5%)	145 (60.4%)	0.023
Intubation	19 (100%)	127 (52.9%)	0.001
Mechanical ventilation	19 (100 %)	124 (51.7%)	0.001
Tracheostomy	12 (63.2 %)	19 (7.9%)	0.001
Prosthesis	0 (0%)	13 (5.4%)	0.607
Operation	1 (5.3%)	39 (16.3%)	0.324
Trauma	1 (5.3%)	17 (7.1%)	1.000
Open wound	1 (5.3 %)	23 (9.6%)	1.000
Decubitus ulcer	3 (15.8%)	14 (5.8%)	0.118
Immunosuppressive therapy	2 (10.5%)	29 (12.1%)	1.000
MRSA colonization	15 (78.9 %)	33 (13.8 %)	0.001

[&] Mann-Whitney U test was performed for median values and Student’s t-test was performed for mean ± SD values

*P < 0.05 was accepted as significant,

LOS: Length of stay; TPN: Total parenteral nutrition

Table 6. Risk factors for MRSA colonization in the multiple logistic regression analysis (forward stepwise).

Risk factors	“Odds ratio”	% 95 CI	P value*
MRSA colonization	14.080	3.896 – 50.884	0.001
Tracheostomy	8.245	2.454 – 27.703	0.001

*P < 0.05 was accepted as significant.

colonization site (87.5%) in our study, which is in line with the literature. The low concentrations of systemically administered antibiotics in the nares allow the persistence of the microorganism. The daily bath with chlorhexidine in our ICUs may explain the low colonization rate of axilla and groin. Nasal decolonization with mupirocin in ICUs is an unresolved question and there is poor evidence for its use. On the other hand, widespread and prolonged use of mupirocin in these settings has been associated with the development of low- and high-level resistance (19). HCWs can be carrier of MRSA. However, we did not detect any HCW nasal carriage in our study, which is parallel with previous studies reporting low rates (19,20). This may be explained by transient or short term carriage. There is no evidence to support routine staff screening in endemic settings. Also nasal decolonization of HCW in endemic settings is not recommended due to recolonization (19).

Identifying the risk factors for MRSA colonization and infection will help to focus on infection-control measures at high-risk groups for MRSA colonization and infection. Despite our efforts to include all risk factors for MRSA colonization or infection, we still could not include all the risk factors (e.g. steroid use, transfer from another unit) and this was a limitation of the study. The length of stay in the hospital and ICU, serious underlying diseases, previous antibiotic therapy, use of invasive devices, and trauma were cited among the major risk factors for the colonization and infection of MRSA in previous studies (14,18,20,21). However, none of the underlying diseases was found to be a risk factor for colonization and infection with MRSA in our study. The length of stay in MICU before colonization, previous use of antibiotic, length of antibiotic therapy before colonization, administration of total parenteral nutrition, intubation, mechanical ventilation, and application of tracheostomy were found as risk factors for colonization in univariate analysis; however, only the length of stay in MICU before colonization and mechanical ventilation were found to be the most significant risk factors in the multiple logistic regression analysis. Probably, the high contact rate of HCWs and performing more invasive procedures increase colonization rates in ventilated patients. Therefore, HCWs should pay more attention to

infection control measures in ventilated patients and in those who stay long in ICUs. On the other hand, only MRSA colonization and tracheostomy were found to be significant risk factors for the MRSA infection. In the literature, a 10-fold increase in the rate of infection in colonized patients has been reported (22), and in our study MRSA infection occurred in 31% of colonized patients with a 14-fold increase. The most common MRSA infection was VAP in this study. Also MRSA is an important cause of VAP with high mortality in many ICUs (23-25). Although we did not calculate the attributable mortality for MRSA infections, the prognosis of MRSA infections was previously found poorer than methicillin-susceptible *S. aureus* infections (25). Despite appropriate glycopeptide therapy, there is an increased attributable mortality for MRSA pneumonia (26). Also, MRSA bacteremia was associated with significantly higher mortality rate compared to MSSA bacteremia. In a study from Turkey, investigators showed that methicillin resistance was one of the independent risk factors for high mortality rate (27).

Previous antibiotic use and length of antibiotic therapy were not found as important risk factors for colonization and infection in multiple logistic regression analysis in this study. However, higher antibiotic consumption is usually related to higher rates of antibiotic resistant microorganisms as well as MRSA in previous studies (14,16). Reduction of antibiotic selection pressure may have a beneficial effect on MRSA colonization. In addition, rational antibiotic therapy is an important issue to reduce antibiotic costs in countries with limited-resources.

Colonized patients are the main reservoir for the spread of MRSA from patient to patient, and HCWs' hands are the most common way of the spread. In our study, 5.6% of patients were already colonized on admission. All these patients had been transferred from other parts of the hospital and there were only 4 clones (clones A, B, C, and D). Moreover, strains from patients colonized during MICU stay were identical to these strains (clones A,B, and C). These results show the spread of isolates from patient-to-patient in the hospital and ICU from colonized patients. However, in this study we did not provide an estimation of colonization pressure during our study

period; previous reports show the impact of colonization pressure and high workload on patient acquisition of MRSA (28,29). An effective infection control program is needed to prevent and control MRSA colonization and infection in the ICU. Active surveillance of newly admitted patients for MRSA and rapid and effective isolation and treatment ("search and destroy policy") may help control the spread of MRSA (30,31). Furthermore, active surveillance cultures and contact/droplet precautions for control of MRSA was found cost-effective (32). The lack of isolation facilities is a common problem in developing countries. However, the success of single-room isolation is controversial in endemic settings and there are detrimental effects on patient care in ICUs, especially in understaffed centers. The isolation rooms may improve the compliance of HCWs to contact precautions (19,33,34).

The major mode of transmission from patient-to-patient is through contaminated hands of HCWs, so the hand hygiene compliance of HCWs and standard precautions are important infection control measures to control MRSA in endemic settings. In many studies, compliance of hand hygiene is poor (<50%) and many factors influence the compliance. In developing countries understaffing, high workload, poorly accessible sinks, and expensive alcohol-based hand disinfectants are the major barriers for the hand hygiene compliance, and so adequate compliance is practically impossible (35). However, we did not observe the hand hygiene compliance of HCWs; the dissemination of MRSA in MICU showed the low compliance despite the alcohol-based products placed at the bedsides and 3 washbasins for 9 beds. Probably understaffing (patient:nurse ratio was 4:1) and heavy workload are important reasons for low compliance and healthcare associated infections in our MICU. Also in many studies from Turkey, compliance with hand washing was low (12%-34 %) and related to low

staff-to-patient ratio, excessive use of gloves, and deficiencies in the infrastructure of ICU (36-41). Hugonnet et al. (42) showed that 26.7% of all infections acquired in the ICU could be prevented providing that around the clock nurse-to-patient ratio is maintained more than 2.2:1. Moreover, in a recent study, investigators examined the infection control practices in the hospitals that had low and high MRSA rates in the Mediterranean, including Turkey. They found that there was no significant correlation between the infection control set-up, hand hygiene facilities, and antibiotic stewardship practices between the hospitals. However, hospitals reporting frequent episodes of overcrowding, particularly involving several departments, and having on-going difficulties in providing isolation beds had significantly higher MRSA proportions. It was reported that the mere presence of infection control teams, suitable hand hygiene facilities, as well as antibiotic stewardship initiatives does not necessarily guarantee the adequate control of multiresistant infections such as MRSA (43).

This study shows the clonal spread of MRSA in an ICU and supports the results of previously published reports. Despite the efforts to improve hand hygiene, such as reminder posters, bedside alcohol-based products, and staff education, the problem emerges because of understaffing and high workload in developing countries.

Acknowledgements

We thank Serhat Ünal from Hacettepe University, Department of Internal Medicine of Medical Faculty for giving us a chance to study in the Molecular Laboratory of Infectious Disease Unit. We also thank staff in the Molecular Laboratory for helping genotyping analyses of isolates and Ahmet Öztürk for statistical analyses.

References

1. Borg MA, Kraker M, Scicluna E, Sande-Bruinsma N, Tiemersma E, Monen J, et al. Prevalence of methicillin-resistant *Staphylococcus aureus* (MRSA) in invasive isolates from southern and eastern Mediterranean countries. *J Antimicrob Chemother* 2007;60:1310-5.
2. Erbay H, Yalçın AN, Serin S, Turgut H, Tomatir E, Cetin B, et al. Nosocomial infections in intensive care unit in a Turkish university hospital: a 2 year survey. *Intensive Care Med* 2003;29:1482-8.

3. Esen S, Leblebicioglu H. Prevalence of nosocomial infections at intensive care units in Turkey: a multicentre 1-day point prevalence study. *Scand J Infect Dis* 2004;36:144-8.
4. Çagatay AA, Özcan PE, Gulec L, Ince N, Tugrul S, Özsüt H, et al. Risk factors for mortality of nosocomial bacteremia in intensive care units. *Med Princ Pract* 2007;16:187-92.
5. Gould IM. The clinical significance of methicillin-resistant *Staphylococcus aureus*. *J Hosp Infect* 2005; 61: 277-82.
6. Alp E, Guven M, Yildiz O, Aygen B, Voss A, Doganay M. Incidence, risk factors and mortality of nosocomial pneumonia in Intensive Care Units: a prospective study. *Ann of Clin Microbiol and Antimicrob* 2004; 3: 17.
7. Aygen B, Yörük A, Yıldız O, Alp E, Kocagöz S, Sümerkan B, et al. Bloodstream infections caused by *Staphylococcus aureus* in a university hospital in Turkey: clinical and molecular epidemiology of methicillin resistant *Staphylococcus aureus*. *Clin Microb Infect* 2004;10:309-14.
8. Esel D, Doğanay M, Alp E, Sümerkan B. Prospective evaluation of blood cultures in Turkey university hospital: epidemiology, microbiology and patient outcome. *Clin Microbiol Infect* 2003; 9: 1038-44.
9. Lencastre H, Couto I, Santos J, Melo-Cristino J, Torres Pereira A. Methicillin-resistant *Staphylococcus aureus* disease in a Portuguese hospital: characterization of clonal types by a combination of DNA typing methods. *Eur J Clin Microbiol Infect Dis* 1994; 13: 64-73.
10. Tenover FC, Arbeit RD, Goering RV, Mickelsen PA, Murray BE, Persing DH, et al. Interpreting chromosomal DNA restriction patterns produced by pulse-field gel electrophoresis: criteria for bacterial strain typing. *J Clin Microbiol* 1995; 33: 2233-9.
11. Garner JS, Jarvis WR, Emori TG, Horan TC, Hughes JM. CDC definitions for nosocomial infections. *Am J Infect Control* 1988;16:128-40.
12. Schito GC. The importance of the development of antibiotic resistance in *Staphylococcus aureus*. *Clin Microbiol Infect* 2006; 12 (suppl): 3-8.
13. Kluytmans-VandenBergh MFQ, Kluytmans JAJW. Community-acquired methicillin-resistant *Staphylococcus aureus*: current perspectives. *Clin Microbiol Infect* 2006; 12 (suppl): 9-15.
14. Oztoprak N, Cevik MA, Akinci E, Korkmaz M, Erbay A, Eren SS, et al. Risk factors for ICU-acquired methicillin-resistant *Staphylococcus aureus* infections. *Am J Infect Control* 2006; 34: 1-5.
15. Marshall C, Harrington G, Wolfe R, Fairley CK, Wesselingh S, Spelman D. Acquisition of methicillin-resistant *Staphylococcus aureus* in a large intensive care unit. *Infect Control Hosp Epidemiol* 2003; 24: 322-6.
16. Garrouste-Orgeas M, Timsit JF, Kallel H, Ben Ali A, Dumay MF, Paoli B, et al. Colonization with methicillin-resistant *Staphylococcus aureus* in ICU patients: morbidity, mortality, and glycopeptide use. *Infect Control Hosp Epidemiol* 2001; 22: 687-92.
17. Korn GP, Martino MDV, Mimica IM, Mimica LJ, Chiavone PA, Musolino LRS. High frequency of colonization and absence of identifiable risk factors for methicillin-resistant *Staphylococcus aureus* (MRSA) in intensive care units in Brazil. *Br J Infect Dis* 2001; 5: 1-7.
18. Corea E, Silva T, Perera J. Methicillin-resistant *Staphylococcus aureus*: prevalence, incidence and risk factors associated with colonization in Sri Lanka. *J Hosp Infect* 2003; 55: 145-8.
19. Marshall C, Wesselingh S, McDonald M, Spelman D. Control of endemic MRSA-what is the evidence? A personal view. *J Hosp Infect* 2004; 56: 253-68.
20. McNeil JJ, Proudfoot AD, Tosolini FA, Morris P, Booth JM, Doyle AE, et al. Methicillin-resistant *Staphylococcus aureus* in an Australian teaching hospital. *J Hosp Infect* 1984; 5: 18-28.
21. Marshall C, Wolfe R, Kossmann T, Wesselingh S, Harrington G, Spelman D. Risk factors for acquisition of methicillin-resistant *Staphylococcus aureus* (MRSA) by trauma patients in the intensive care unit. *J Hosp Infect* 2004; 57: 245-52.
22. Davis KA, Stewart JJ, Crouch HK, Florez CE, Hospenthal DR. Methicillin-resistant *Staphylococcus aureus* (MRSA) nares colonization at hospital admission and its effect on subsequent MRSA infection. *Clin Infect Dis* 2004; 39: 776-82.
23. Aybar Türkoglu M, Topeli İskit A. Ventilator-associated pneumonia caused by high risk microorganisms: a matched case-control study. *Tüberküloz ve Toraks Dergisi* 2008;56:139-49.
24. Meric M, Willke A, Caglayan C, Toker K. Intensive care unit-acquired infections: incidence, risk factors and associated mortality in a Turkish University Hospital. *Jpn J Infect Dis* 2005;58:297-302.
25. Erbay RH, Yalcin AN, Zencir M, Serin S, Atalay H. Costs and risk factors for ventilator-associated pneumonia in a Turkish University Hospital's intensive care unit: a case-control study. *BMC Pulmonary Medicine* 2004;4:3.
26. Rello J, Sole-Violan J, Sa-Borges M, Garnacho-Montero J, Munoz E, Sirgo G, et al. Pneumonia caused by oxacillin-resistant *Staphylococcus aureus* treated with glycopeptides. *Crit Care Med* 2005;33:1983-7.
27. Topeli A, Ünal S, Akalin HE. Risk factors influencing clinical outcome in *Staphylococcus aureus* bacteremia in a Turkish University Hospital. *Int J Antimicrob Agents* 2000;14:57-63.
28. Merre J, Santoli F, Appere-De Vecchi C, Tran B, De Jonghe B, Outin H. "Colonization pressure" and risk of acquisition of methicillin-resistant *Staphylococcus aureus* in a medical intensive care unit. *Infect Control Hosp Epidemiol* 2000; 21: 718-23.
29. Bonten MJ, Slaughter S, Ambergen AW, Hayden MK, van Voorhis J, Nathan C, et al. The role of "colonization pressure" in the spread of vancomycin-enterococci: an important infection control variable. *Arch Intern Med* 1998; 158: 1127-32.

30. Huang SS, Yokoe DS, Hinrichsen VL, Spurchise LK, Datta R, Miroshnik I, et al. Impact of routine intensive care unit surveillance cultures and resultant barrier precautions on hospital-wide methicillin-resistant *Staphylococcus aureus* bacteremia. *Clin Infect Dis* 2006; 43: 971-8.
31. Cepeda JA, Whitehouse T, Cooper B, Hails J, Jones K, Kwaku F, et al. Isolation of patients in single rooms or cohorts to reduce spread of MRSA in intensive-care units: prospective two-centre study. *Lancet* 2005; 365: 295-304.
32. Karchmer TB, Durbin LJ, Simonton BM, Farr BM. Cost-effectiveness of active surveillance cultures and contact/droplet precautions for control of methicillin-resistant *Staphylococcus aureus*. *J Hosp Infect* 2002; 51: 126-32.
33. Cooper BS, Stone SP, Kibbler CC, Cookson BD, Roberts JA, Medley GF, et al. Isolation measures in the hospital management of methicillin resistant *Staphylococcus aureus* (MRSA): systematic review of the literature. *BMJ* 2004; 329: 533-8.
34. Harbarth S. Control of endemic methicillin-resistant *Staphylococcus aureus*-recent advances and future challenges. *Clin Microbiol Infect* 2006; 12: 1154-62.
35. Raza MW, Kazi BM, Mustafa M, Golud FK. Developing countries have their own characteristic problems with infection control. *J Hosp Infect* 2004; 57: 294-9.
36. Sacar S, Turgut H, Kaleli I, Cevahir N, Asan A, Sacar M, et al. Poor hospital infection control practice in hand hygiene, glove utilization, and usage of tourniquets. *Am J Infect Control* 2006;34:606-9.
37. Karabay O, Sencan I, Sahin I, Alpteker H, Ozcan A, Oksuz S. Compliance and efficacy of hand rubbing during in-hospital practice. *Med Princ Pract* 2005;14:313-7.
38. Akyol AD. Hand hygiene among nurses in Turkey:opinions and practices. *J Clin Nurs* 2007;16:431-7.
39. Kuzu N, Özer F, Aydemir S, Yalcin AN, Zencir M. Compliance with hand hygiene and glove use in a university-affiliated hospital. *Infect Control Hosp Epidemiol* 2005;26:312-5.
40. Karabay S, Ay P, Derbentli Ş, Nakıpoğlu Y, Esen F. Handwashing frequencies in an intensive care unit. *J Hosp Infect* 2002;50:36-41.
41. Makay Ö, İçöz G, Yılmaz A, Kolcu F. Yoğun bakım çalışanlarının el yıkama alışkanlıkları. *Ulus Travma Acil Cerrahi derg* 2008;14:149-53.
42. Hugonnet S, Chevreton JC, Pittet D. The effect of workload on infection risk in critically ill patients. *Crit Care Med* 2007; 35: 76-81.
43. Borg MA, Cookson BD, Rasslan O, Gür D, Ben Redjeb S, Benbachir M, et al. Correlation between methicillin-resistant *Staphylococcus aureus* prevalence and infection control initiatives within southern and eastern Mediterranean hospitals. *J Hosp Infect* 2008; (in press).