

1-1-2002

Muscle Fiber Types of the Tunica Muscularis Externa at the Upper Part of the Sparrow (*Passer domesticus*) Esophagus

FATİME GEYİKOĞLU

AYSEL TEMELLİ

ABDULLAH ÖZKARAL

Follow this and additional works at: <https://journals.tubitak.gov.tr/zoology>



Part of the [Zoology Commons](#)

Recommended Citation

GEYİKOĞLU, FATİME; TEMELLİ, AYSEL; and ÖZKARAL, ABDULLAH (2002) "Muscle Fiber Types of the Tunica Muscularis Externa at the Upper Part of the Sparrow (*Passer domesticus*) Esophagus," *Turkish Journal of Zoology*. Vol. 26: No. 2, Article 11. Available at: <https://journals.tubitak.gov.tr/zoology/vol26/iss2/11>

This Article is brought to you for free and open access by TÜBİTAK Academic Journals. It has been accepted for inclusion in Turkish Journal of Zoology by an authorized editor of TÜBİTAK Academic Journals. For more information, please contact academic.publications@tubitak.gov.tr.

Muscle Fiber Types of the Tunica Muscularis Externa at the Upper Part of the Sparrow (*Passer domesticus*) Esophagus

Fatime GEYİKOĞLU

Department of Biology, Faculty of Arts and Sciences, Atatürk University, 25240, Erzurum - TURKEY

Aysel TEMELLİ

Department of Biology, Education Faculty of Kazım Karabekir, Atatürk University, 25240, Erzurum - TURKEY

Abdullah ÖZKARAL

Division of Histology and Embryology, Medical Faculty, Atatürk University, 25240, Erzurum - TURKEY

Received: 13.12.2000

Abstract: Muscle fiber types of the tunica muscularis externa of the upper part of the sparrow esophagus were characterised by the method of myofibrillar adenosine triphosphatase (mATPase). Muscle fibers were described as type II and type III. Muscle fiber types were histochemically different from those of the esophagus of broiler ross. It is thought that this difference should be evaluated according to the functional difference between the esophagus of domestic and wild bird species.

Key Words: *Passer domesticus*, Aves, Esophagus, Histochemistry, Muscle

Serçe (*Passer domesticus*) Özofagusunun Yukarı Bölümünde Tunika Muskularis Eksterna'da Kas Lifi Tipleri

Özet: Serçe özofagusunun yukarı bölümünde, tunika muskularis eksterna'nın kas lifi tipleri miyofibriller adenzin trifosfataz (mATPase) metodu ile karakterize edildi. Kas lifleri bu metoda göre, tip II ve tip III olarak tanımlandı. Kas lifi tipleri, histokimyasal olarak etlik besi pilici özofagusundakinden farklıydı. Bu farklılığın, evcil ve yabani kuş türleri arasında özofagusun fonksiyonuna bağlı olarak değerlendirilmesi gerektiği kanaatine varıldı.

Anahtar Sözcükler: Serçe, Kuş, Özofagus, Histokimya, Kas

Introduction

The esophagus of birds was studied experimentally (1) and comparative physiological investigations of mammalian and bird digestive systems were performed (2). It has been reported that the striated muscle fibers of birds are less developed in various nutritional conditions (3) whereas smooth muscle fibers show different development (4). Myogenic variations depending on oxygen and nutrition have been discussed previously (5, 6).

There are more studies on the neuronal control of the esophagus than on the myogenic control system (MCS) (7). Irregularity of the esophageal movements is thought to be related to the upper esophageal striated muscles instead of the lower esophageal smooth muscles (8).

There are many investigations on the mammalian esophagus (9-14) while less studies have been done on the bird esophagus (15-18). The mammalian upper esophageal sphincter muscles have been studied by the mATPase method (19). Cryostat sections used for mammalian muscle fibers (20) have also been used to study bird breast and leg muscles (21,22). It has been reported that the morphological and physiological study of the muscle structures of different animal groups is very important for evaluating the pharmacological properties of the esophagus (23). Birds have been chosen as a model to study the pharmacological properties of the skeletal muscle (24). When a comparison was made between domestic and wild bird species, it was observed that there were differences in the mATPase activity of the muscle fibers (25). However, until now, there has been

no study on the histochemical structure of the esophagus muscles of the sparrow. Therefore, in this study, cryostat sections from the upper esophagus of the sparrow were analyzed by the mATPase method and the types of muscle fibers were evaluated histochemically.

Materials and Methods

Ten sparrow esophagus samples were removed. Frozen sections of the wet upper esophagus were cut at 12-15 μm thickness in a cryostat. These sections were subjected to the mATPase method at 25°C at pH 4.25 for five minutes and at pH 10.4 for 15 minutes after acid and alkaline preincubations at 40°C in order to determine the fiber types (26-30). Evaluation of the fiber types was similar to that carried out by other authors (27,31-35).

Results and Discussion

As in the striated muscle of the birds (3,36,37), and as in our previous studies (38,39) three fiber types were identified in the esophagus of the broiler ross (broiler chicken) (type I, type IIA and type IIB). In this study, two fiber types (II and III) were determined in the sparrow esophagus. With the application of mATPase following acid preincubation, it was observed that type II fibers and type III fibers with small diameters give weak and mid-level mATPase reactions respectively (Fig. 1). It has been reported that small fiber diameters have a high degree of aerobic capacity (40). After alkaline

preincubation, following differentiation with acid preincubation, fibers were stained dark and showed high mATPase reaction (Fig. 2a, b).

Different mATPase reactions have been reported for type III fibers in the bird skeletal muscle (27,28,30,36,41). But, as we have reported for the sparrow triceps (30), the term type III, generally used for the sparrow esophagus in this study, is preferred also for this fiber type due to its resistance to both alkaline and acidic applications (31,41). When we compared it with the other fiber types, we thought that different mATPase reactions in type III fibers are specialized (42,43) for slow and supportive contractions in the sparrow esophagus, as in the bird skeletal muscle (27,28,30,36,44).

The fibers we have differentiated in the sparrow esophagus that are irrisistant in acid and resistant in alkaline, are called twitch (28,33), fast-twitch (27,36,45) or type II (31,32,41) by various authors. The fibers that we have described as type II using the mATPase method are also called fast-twitch glycolytic (IIB) (46-50) or white (51,52) after nitroblue tetrazolium nicotinamide adenin dinucleotide phosphate (NBT) or succinic dehydrogenase (SDH) staining methods. It is reported that they have low mATPase activity (53) and low aerobic capacity (3,52), like the mammalian type IIB fibers, and consume glycogen (48). As in the bird skeletal muscle (49-55), in the chicken esophagus (38) and sparrow esophagus, these fibers might be related to the ability to increase the rate in the swallowing motion as well (56).

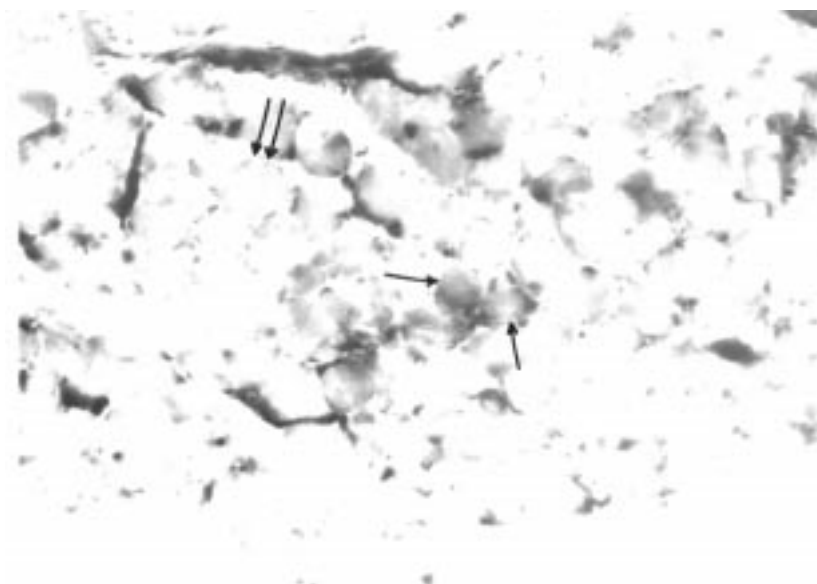


Figure 1. Muscle fiber types of the tunica muscularis externa of the upper part of the sparrow esophagus. Arrow: type III, Double arrow: type II. mATPase after acid preincubation (x256).



Figure 2. Tunica muscularis externa at the upper part of the sparrow esophagus (serial transversal sections). Arrow: type III. Double arrow: type II. Asterisk: The part evaluated in section. a) mATPase after acid preincubation. b) mATPase after alkaline preincubation (the fiber types are stained dark) (x40).

Different types of muscle fibers (32) in every species and in every particular muscle (21,33,57-61) may have adapted to meet functional needs (60,61). Furthermore, morphological adaptations of bird muscles depending on changes in nutrition type and nutrition model are recorded (62). Similar adaptations have also been reported histochemically (50). The mATPase activity of the muscle fibers was compared with that in the chicken, and it was found to be higher in wild bird species (25).

Thus, in birds such as the sparrow (eating hard cereals, fruits, insects, etc.) (63,64), the muscle fibers at the upper portion of the esophagus can be differentiated into specific types according to their functional roles (types II and III). We think that the muscle fibers of the upper esophagus of the bird should be analyzed morphologically, histochemically and functionally, with more intensive and different approaches.

References

1. Delikaris, P. G., Hatzipantelis, K. P., Filintatzi, C., Kotakidou, R. E., Kitis, G. and Raptopoulos, D., The Use of Dura Mater Patch to Cover Oesophageal Defects of Different Sizes: An Experimental Study in Chickens. *Eur. J. Surg.*, 165 (2): 152-157, 1999.
2. Vylitova, M., Miksik, I. and Pacha, J., Metabolism of Corticosterone in Mammalian and Avian Intestine. *Gen. Comp. Endocrin.*, 109 (3): 315-324, 1998.
3. Ono, Y., Goto, T., Nasu, R., Iwamoto, H., Takayama, K., Nakanishi, Y. and Manda, M., Comparative Studies of Muscle Histochemical Characteristics of Ducks Between Free Ranged and Housed, and Among Three Types of Breeds. *Jap. Poultry Sci.*, 35 (6): 367-375, 1998.
4. Shamoto, K., Yamauchi, K. and Kamisoyama, H., Morphological Alterations of the Duodenal Villi in Chicks Fed Rice Bran or Grower Mash After Fasting. *Jap. Poultry Sci.*, 36(1): 38-46, 1999.
5. Chiou, P. W. S., Chen, C. L., Chen, K. L. and Wu, C. P., Effect of High Dietary Copper on the Morphology of Gastro-Intestinal Tract in Broiler Chickens. *J. Animal Sci.*, 12(4): 548-553, 1999.
6. Murray, B. and Wilson, D. J., Muscle Patterning, Differentiation and Vascularisation in the Chick Wing Bud. *J. Anat.*, 190 (2): 261-273, 1997.
7. Perikisaitis, H. G. and Diamant, N. E., Myogenic Mechanism for Peristalsis in the Cat Esophagus. *Am. J. Physiol.*, 277 (2 Part 1): 306-313, 1999.
8. Haylett, K. R., Vales, P. and McCloy, R. F., An Investigation Into the Spatial Relationship Between Complexity and Motility Within the Esophagus. *Physiol. Measurement*, 19 (4): 463-479, 1998.
9. Stein, H. J., Meffert, D. L., Demeester, T. R. and Seiwert, R. S., Three-Dimensional Pressure Image and Muscular Structure of the Human Lower Esophageal Sphincter. *Surg.*, 117 (6): 692-698, 1995.
10. Uddman, R., Grunditz, T., Luts, A., Desai, H., Fernstrom, G. and Sundler, F., Distribution and Origin of the Peripheral Innervation of Rat Cervical Esophagus. *Dysph.*, 10(3): 203-212, 1995.
11. Luiking, Y. C., Weusten, B. L.A., Portincasa, P., Meer, V. D., Smout, J. P. M. and Akkermans, L. M. A., Effects of Long-Term Oral L-Arginine on Esophageal Motility and Gallbladder Dynamics in Healthy Humans. *Am. J. Physiol.*, 274 (6 Part 1): 984-991, 1998.

12. Anggiansah, A., Taylor, G., Marshall, R. E. K., Bright, N. F., Owen, W. A. and Owen, W.J., Oesophageal Motor Responses to Gastro-Oesophageal Reflux in Healthy Controls and Reflux Patients. *Gut*, 14 (5): 600-605, 1997.
13. Frizen, N. A. and Frizen, B. N., New Data on the Pharyngoesophageal Junction. *Fiziol. Cheloveka*, 23 (4): 106-109, 1997.
14. Percy, W. H., Miller, A. J. and Brunz, J. T., Pharmacologic Characteristics of Rabbit Esophageal Muscularis Mucosae in Vitro. *Digest. Disease Sci.*, 42 (12): 2537-2546, 1997.
15. Salvi, E., Vaccaro, R. and Renda, T. G., An Immunohistochemical Study of the Neuroendocrine System in the Chicken Oesophagus. *Anat. Embryol.*, 197 (4): 283-291, 1998.
16. Bryk, S. G., Gheri, G. and Sgambati, E., Computerized Morphometric Analysis of the Chick Embryo Esophagus. *Ital. J. Anat. Embryol.*, 98 (1): 1-12, 1993.
17. Postorino, A., Vetri, T., Bonvisutto, F., Fileccia, R. and Urso, S.A., Evidence for Involvement of Nitric Oxide (NO) or a Related Nitroso-Compound in NANC Inhibitory Neurotransmission in the Pigeon Oesophageal Smooth Muscle. *J. Aut. Pharmacol.*, 19 (2): 85-90, 1999.
18. Gille, U., Salamon, F.V. and Roennert, J., Growth of the Digestive Organs in Duck with Considerations on their Growth in Birds in General. *Poultry Sci.*, 40 (2): 194-202, 1999.
19. Mu, L. and Sanders, I., Neuromuscular Organization of the Human Upper Esophageal Sphincter. *Ann. Otol. Rhinol. Laryngol.*, 107 (5 Part 1): 370-377, 1998.
20. Manta, P., Kalfakis, N., Kararizou, E., Vassilopoulos, D. and Papageorgiou, C., Size and Proportion of Fiber Types in Human Muscle Fascicles. *Clin. Neuropathol.*, 15 (2) : 116-118, 1996.
21. Pyornila, A. E. I., Putaala, A. P. and Hissa, R.K., Fibre Types in Breast and Leg Muscles of Hand-Reared and Wily Grey Partridge (*Perdix perdix*). *Can. J. Zool.* 76 (2): 236-242, 1998.
22. Gille, U., Saloman, F.V. and Kattein, S., Posthatching Myofiber Development in the M. Pectoralis of White Pekin Ducks. *Anat. Rec.*, 250 (2): 154-158, 1998.
23. Calore, E. E., Cavaliere, M.J., Haraguchi, M., Gorniak, S. L. Dagli, M.L.Z., Raspantini, P.C. and Calore, N.M.Z., Experimental Mitochondrial Myopathy Induced by Chronic Intoxication by *Senna occidentalis* Seeds. *J. Neurol. Sci.*, 146 (1): 1-6, 1997.
24. Carmona, M., Finol, H. J., Marquez, A. and Noya, O., Skeletal Muscle Ultrastructural Pathology in *Serinus canarius* Infected with *Plasmodium cathemerium*. *J. Submic. Cytol. Pathol.*, 28 (1): 87-91, 1996.
25. Dalla, L. and Carpena, E., Myosin Heavy and Light Chains Kinase in Skeletal and Smooth Muscle of Some Wild Avian Species. *Compar. Biochem. Physiol. B*, 116 (1): 45-50, 1997.
26. Fouces, V., Torella, J. R., Palameque, J. and Viscor, G., A Histochemical ATPase Method for the Demonstration of the Muscle Capillary Network. *J. Histochem. Cytochem.*, 41 (2): 283-289, 1993.
27. Rosser, B. W. C. and George, J. C., The Avian Pectoralis: Histochemical Characterization and Distribution of Muscle Fiber Types. *Can. J. Zool.*, 64: 1174-1185, 1985.
28. Rosser, B. W. C. and George, J. C., Some Histochemical Properties of the Fiber Types in the Pectoralis Muscle of an Emu (*Dromaius novaehollandiae*). *Anat. Rec.*, 209: 301-305, 1984.
29. Bancroft, S. D. and Stevens, A., Theory and Practice of Histological Techniques. Churchill Livingstone, Edinburgh (2nd ed.): p. 662, 1982.
30. Geyikoğlu, F. and Özkara, A., A Histochemical Study of the Origin Regions of the Triceps Muscle of the Sparrow (*Passer domesticus*). *Tr. J. Zool.*, 24 (1): 107-111, 2000.
31. Sohail, G. S. and Sickles, D. W., Embryonic Differentiation of Fiber Types in Normal, Paralysed and Aneurial Avian Superior Oblique Muscle. *J. Embryol. Exp. Morphol.*, 96: 79-97, 1986.
32. Ogata, T., Morphological and Cytochemical Features of Fiber Types in Vertebrate Skeletal Muscle. *Anat. Cell Biol.*, 1 (2): 229-275, 1988.
33. Torella, J. R., Fouces, V., Palameque, J. and Viscor, G., Innervation Distribution Pattern, Nerve Ending Structure and Fiber Types in Pigeon Skeletal Muscle. *Anat. Rec.*, 237: 178-186, 1993.
34. Hooper, A. C. B., Length, Diameter and Number of Aging Skeletal Muscle Fibres. *Geront.*, 27: 121-126, 1981.
35. Armstrong, R. B. and Phelps, R. O., Muscle Fiber Type of Composition of the Rat Hindlimb. *Am. J. Anat.*, 171: 259-272, 1984.
36. Hikida, R. S., Quantitative Ultrastructure of Histochemically Identified Avian Skeletal Muscle Fiber Types. *Anat. Rec.*, 218: 128-135, 1987.
37. Spurway, N. C., Objective Characterization of Cells in Terms of Microscopical Parameters: An Example from Muscle Histochemistry. *J. Histochem.*, 13(2): 269-317, 1981.
38. Geyikoğlu, F. and Temelli A., The Postnatal Histochemical Changes in the Tunica Muscularis Externa of the Upper Part of Esophagus of Broiler Ross. *J. Fac. Sci. Ege Univ.*, (In Press).
39. Geyikoğlu, F., Temelli, A. and Özkara, A., The Changes in Mitochondrial Content and Development of Tunica Muscularis Externa in the Upper Esophagus of the Broiler Ross. *Univ. İstanbul, Fac. Sci.*, (In Press).
40. Mathieu-Costello, O., Brill, R. W. and Hochachka, P. W., Structural Basis for Oxygen Delivery: Muscle Capillaries and Manifolds in Tuna Red Muscle. *Comp. Biochem. Physiol. A*, 113 (1): 25-31, 1996.
41. Bakou, S., Cherel, Y., Gabinaud, B., Guigand, L. and Wyers, M., Type-Specific Changes in Fibre Size and Satellite Cell Activation Following Muscle Denervation in Two Strains of Turkey (*Meleagris gallopavo*). *J. Anat.* 188(3) 677-691, 1996.
42. Suzuki, A., Tsuchiya, T. and Tamate, H., Histochemical Properties of Myofiber Types in Thigh Muscles of the Chicken. *Acta Histochem. Cytochem.*, 15: 362-371, 1982.

43. Putnam, R. W., Gleeson, T.T. and Bennet, A. F., Histochemical Determination of the Fiber Composition of Locomotory Muscles in a Lizard, *Dipsosaurus dorsalis*. J. Exp. Zool., 214: 303-309, 1980.
44. Meyers, R. A., Anatomy and Histochemistry of Spread-Wing Posture in Birds. I. Wing Drying Posture in the Double-Crested Cormorant, *Phalacrocorax auritus*. J. Morphol., 233 (1): 67-76, 1997.
45. Light, J. K., Rapoll, E. and Wheeler, T.M., The Striated Urethral Sphincter: Muscle Fibre Types and Distribution in the Prostatic Capsule. British J. Urol., 79 (4): 539-542, 1997.
46. Whitmore, I., Oesophageal Striated Muscle Arrangement and Histochemical Fibre Types in Guinea pig, Marmoset, Macaque and Man., J. Anat., 134 (4): 685-695, 1982.
47. Wada, N., Miyata, H., Tomita, R., Ozawa, S. and Tokuriki, M., Histochemical Analysis of Fiber Composition of Skeletal Muscles in Pigeons and Chickens. Arch. Italiennes Biol., 137 (1): 75-82, 1999.
48. Sokoloff, A. J. and Goslow Jr, G. E., Neuromuscular Organization of Avian Flight Muscle: Architecture of Single Muscle Fibres in Muscle Units of the Pectoralis (Pars Thoracicus) of Pigeon (*Columba livia*). Philosoph. Transact. Royal Soc. London B Sci., 354 (1385): 917-925, 1999.
49. Sokoloff, A. J., Ryan, J. M., Valerie, E., Wilson, D.S. and Goslow Jr, G. E., Neuromuscular Organization of Avian Flight Muscle: Morphology and Contractile Properties of Motor Units in the Pectoralis (Pars Thoracicus) of Pigeon (*Columba livia*). J. Morphol., 236 (3): 179-208, 1998.
50. Benjamin, W. C. R., Norris, B. J. and Nementh, P.M., Metabolic Capacity of Individual Muscle Fibers from Different Anatomic Locations. J. Histochem. Cytochem., 40 (6): 819-825, 1992.
51. Baeza, S., Marche, G. and Wacrenier, N., Effect of Sex on Muscular Development of Muscovy Ducks. Reproduct. Nutrit. Develop., 39 (5-6): 675-682, 1999.
52. Prasad, R. V., Chandrasekhara, T.S. and Vijayaragavan, C., Histochemical Characterization of Fibre Types in M. Supracoracoideus and M.Latissimus Dorsi (Anterior and Posterior) of Japanese Quail (*Coturnix coturnix japonica*). Ind. Vet. J., 72 (7): 716-717, 1995.
53. Lana, D. P., Leferovich, J. M., Kelly, A. M. and Hughes, S.H., Selective Expression of A Ski Transgene Affects IIB Fast Muscles and Skeletal Structure. Develop. Dynam. 205 (1): 13-23, 1996.
54. Spurway, N. C., Murray, M. G., Gilmour, W.H. and Montgomery, I., Quantitative Skeletal Muscle Histochemistry of Four East African Ruminants. J. Anat., 188 (2): 455-472, 1996.
55. Dial, K. P., Kaplan, S. R. and Goslow Jr., G. E. A Functional Analysis of the Primary Upstroke and Downstroke Muscles in the Domestic Pigeon (*Columbia livia*) During Flight. J. Exp. Biol., 134: 1-16, 1988.
56. Eriksson, P. O., Eriksson, A., Ringavist, M. and Thornel, L.E., Histochemical Fibre Composition of the Human Digastric Muscle. Arch. Oral Biol., 27 (3): 207-215, 1982.
57. Torella, J. R., Fouces, V. Palomaque, J. and Viscor, G., Capillarity and Fibre Types in Locomotory Muscles of Wild Yellow-Legged Gulls (*Larus cachinnans*). Physiol. Zool., 71 (4): 425-434, 1998.
58. Zhang, M., Korshi, K. and McLennan, I.S., Skeletal Muscle Fibre Types: Detection Methods and Embryonic Determinants. Histol. Histopathol., 13(1): 201-207, 1998.
59. Torella, J. R., Fouces, V., Palomeque, J. and Viscor, G., Capillarity and Fibre Types in Locomotory Muscles of Wild Mallard Ducks (*Anas platyrhynchos*). J. Comp. Physiol. B Biochem. Syst. and Environ. Physiol., 166 (3): 164-177, 1996.
60. Torella, J. R., Fouches, V., Palomeque, J. and Viscor, G., Capillary and Fiber Types in Locomotor, Muscles of Wild Common Coots, *Fulica atra*. J. Morphol., 237 (2): 147-164, 1998.
61. Burkholder, T. J., Fingado, B., Baron, S. and Lieber, R. L., Relationship Between Muscle Fiber Types and Sizes and Muscle Architectural Properties in the Mouse Hindlimb. J. Morphol., 221 (2): 177-190, 1994.
62. Bhattacharyya, B. N., The Role of M. Pterygoideus in Closure of the Beaks in Certain Columbidae Birds: A Functional Morphological Analysis Reflecting Diversity in Feeding. Proceed. Zool. Soc., 50 (2): 171-180, 1997.
63. Demirsoy, A., Yaşamın Temel Kuralları (Omurgalılar / Amniyota). Meteksan A. Ş., Ankara, s.942, 1992.
64. Kuru, M., Omurgalı Hayvanlar, Gazi Üniv. İletişim Fak. Matbaası (2.Baskı), Ankara, s. 841, 1996.