

1-1-2000

The Effect of Kinetin, Gibberellic Acid and IndoleAcetic Acid on EMS-Induced Somatic Mutation andRecombination in Drosophila melanogaster

ELİF YEŞİLADA

Follow this and additional works at: <https://journals.tubitak.gov.tr/biology>



Part of the [Biology Commons](#)

Recommended Citation

YEŞİLADA, ELİF (2000) "The Effect of Kinetin, Gibberellic Acid and IndoleAcetic Acid on EMS-Induced Somatic Mutation andRecombination in Drosophila melanogaster," *Turkish Journal of Biology*. Vol. 24: No. 2, Article 10. Available at: <https://journals.tubitak.gov.tr/biology/vol24/iss2/10>

This Article is brought to you for free and open access by TÜBİTAK Academic Journals. It has been accepted for inclusion in Turkish Journal of Biology by an authorized editor of TÜBİTAK Academic Journals. For more information, please contact academic.publications@tubitak.gov.tr.

The Effect of Kinetin, Gibberellic Acid and Indole Acetic Acid on EMS-Induced Somatic Mutation and Recombination in *Drosophila melanogaster*

Elif YEŞİLADA

Department of Biology, Art and Science Faculty, İnönü University, 44069 Malatya-TURKEY

Received: 28.01.1999

Abstract: The effect of plant growth hormones (kinetin, gibberellic acid (GA₃) and indole acetic acid (IAA)) on EMS-induced mutant wing spots was studied with the somatic mutation and recombination test (SMART) in *Drosophila melanogaster*. GA₃ reduced all kinds of EMS-induced spot. While a 10⁻³ M concentration of kinetin reduced only the number of EMS-induced twin spots, a 10⁻⁴ M concentration was seen to increase the number of all types of spot. The same concentrations of IAA gave variable results. A 10⁻⁴ M concentration of IAA caused a decrease only in the number of large single spots. Our results confirmed that plant growth hormones, especially GA₃, can act as a bio-antimutagen.

Key Words: Ethyl Methanesulphonate, Gibberellic Acid, Indole Acetic Acid, Kinetin, Antimutagenicity, Somatic Mutation and Recombination Test, *Drosophila melanogaster*.

***Drosophila melanogaster*'de EMS ile İndüklenmiş Somatik Mutasyon ve Rekombinasyon Üzerine Kinetin, Gibberellik Asit ve İndol Asetik Asitin Etkisi**

Özet: Bu çalışmada, *Drosophila melanogaster*'in somatik mutasyon ve rekombinasyon testi kullanılarak EMS ile indüklenmiş mutant kanat benekleri üzerine bitki büyüme hormonlarının (kinetin, gibberellik asit (GA₃) ve indol asetik asit (IAA)) etkisi araştırıldı. GA₃, EMS ile indüklenen bütün kanat beneklerini azalttı. Kinetinin 10⁻³ M konsantrasyonu EMS ile indüklenen ikili beneklerin sayısını azaltırken, 10⁻⁴ M konsantrasyonu beneklerin bütün tiplerinin sayısında artışa neden oldu. Aynı konsantrasyonlardaki IAA ise değişken sonuçlar verdi. 10⁻⁴ M IAA yalnızca büyük tekli beneklerin sayısını indirdi. Sonuçlarımız bitki büyüme hormonlarının (özellikle GA₃'ün) bio-antimutagen olabileceğini düşündürmektedir.

Anahtar Sözcükler: Etil Metansulfonat, Gibberellik Asit, Indol Asetik Asit, Kinetin, Antimutajenite, Somatic Mutasyon ve Rekombinasyon Testi, *Drosophila melanogaster*.

Introduction

Various types of inhibitor and suppressor against mutagens and carcinogens are found in several organisms and environments. Two main chemical groups of inhibitors against various

mutagens have been identified, based on their modes of action: desmutagens, which inactivate the chemical mutagen; and bio-antimutagens, which activate the repair system until fixation of the mutation occurs (1-3). Many antimutagens were discovered by bacterial tests such as the Ames test. Cancer depression tests are also required, but those using mammals require too much time and expense. The wing spot test with *Drosophila melanogaster* has become useful as a second screening method for mutagens between bacterial and mammalian systems. *Drosophila* is a higher organism than bacteria, being more similar to mammals with its chromosomes and P450 enzymes (4).

There are some reports on the antimutagenic effect of plant growth hormones in plants. It was reported that plant growth hormones reduced the mutagenesis caused by gamma rays, X-rays, EMS (Ethyl Methanesulfonate) and sodium azide (NaN_3) (5-10). The mechanism of antimutagenic effect has yet to be elucidated. This study was carried out in order to determine whether plant growth hormones were effective in a similar fashion when applied to animals.

Thus, the suppressing effect of plant growth hormones (kinetin, gibberellic acid (GA_3) and indole acetic acid (IAA)) on EMS-induced mutant wing spots in the SMART assay of *Drosophila melanogaster* was tested. There have been several reports on the application of the wing spot test for bio-antimutagens (2, 11-13).

Materials and Methods

1. Chemicals

Drosophila instant medium was obtained from Ward's Biology. Ethyl methanesulfonate (EMS) was from Merck (Darmstadt, Germany). Indole acetic acid (IAA), gibberellic acid (GA_3) and kinetin were obtained from the Sigma Chemical Co. (St. Louis, Mo, USA).

2. *Drosophila* Stocks

We used two stocks of *D. melanogaster* for a cross of mwh males with $\text{flr}^3/\text{In}(3\text{LR})\text{TM}\bar{3},\text{r}^{\text{ip}}\text{sep bx}^{34\text{e}}\text{e}^{\text{s}}\text{Ser}$ females (for a description of the markers see Lindsley and Zimm 1992) (14). These stocks were originally obtained from the laboratory of Ulrich Graf, Swiss Federal Institute of Technology (ETH), University of Zurich. Both mwh and flr^3 recessive wing mutations are located on the left arm of chromosome 3. Homozygous mwh wing blade cells develop several trichomes per cell instead of the usual single one and homozygous flr^3 cells produce deformed trichomes. Flies were reared at $25\pm 1^\circ\text{C}$ and 60% relative humidity on cornmeal-agar medium (15).

3. Experimental Procedures

Eggs from crosses between the two strains mentioned above were collected during an 8 h period. The larvae were allowed to grow to the 3rd instar for 72 ± 4 h and were transferred to a culture bottle which contained cellulose powder and EMS (50 mM), to which they were exposed for 6 h. The EMS-treated larvae were counted and moved to an instant medium which contained a plant growth hormone. To test the mutagenicity, the mutagen-treated larvae were

transferred to an instant medium without a plant growth hormone. Emerged adult flies of trans-heterozygous (*mwh/flr³*) were stored in 70% ethanol. Their wings were mounted in Faure's solution and examined under 500x magnification for the presence of mosaic spots. The number of spots as well as their types and size were recorded. The observed wing hair spots were classified as small single spots (SSS), large single spots (LSS) or twin spots (TS) according to the method of Graf et al. (4). One or two *mwh* or *flr* mutant cells were scored as small single spots. Three or more *mwh* or *flr* mutant cells were scored as large single spots. Neighboring *mwh* and *flr* mutant cells were scored as twin spots. The χ^2 -test was performed on the number of spots per wing for each plant growth hormone-treated group and the positive control (mutagen only) (16).

Results and Discussion

EMS is a alkylating agent which is able to produce point mutations and small deletions as well as chromosome breakage; its recombinogenic activity is also known (17, 18). First, the mutagenicity of EMS was investigated. In this study all kinds of spot (SSS, LSS and TS) were seen to be increased with EMS at 50 mM (Table 1). This concentration of EMS was used throughout the experiments.

The antimutagenic effects of plant growth hormones (kinetin, GA₃ and IAA) tested on EMS-induced mutation, are shown in Table 1 and Fig. 1. Three types of spot are considered to be the result of gen mutations and chromosome aberrations in SMART (4, 19, 20). Small single spot and large single spot are due to gene mutations, chromosome deletion, non-disjunction, or mitotic recombination. The twin spot is due exclusively to mitotic recombination (4). In this study, 10⁻⁴ M kinetin had no ability to decrease the mutagenicity of EMS. However, a significant

Table 1. The effects of kinetin, GA₃ and IAA on the mutagenicity of EMS.

Treatment	No. of wings	No. of spots / wing			Total spots
		Small single spots	Large single spots	Twin spots	
Control (water)	200	0.04*	0.02*	0.01*	0.06*
Control (EMS)	253	0.23	0.45	0.17	0.85
10 ⁻⁴ M Kinetin	165	0.33*	0.82*	0.20	1.35*
10 ⁻³ M Kinetin	88	0.20	0.38	0.08*	0.65
10 ⁻⁴ M GA ₃	210	0.11*	0.40	0.01*	0.52*
10 ⁻³ M GA ₃	98	0.13*	0.27*	0.02*	0.42*
10 ⁻⁴ M IAA	89	0.13	0.29*	0.13	0.56*
10 ⁻³ M IAA	79	0.23	0.62*	0.13	0.97

* Significantly different from the control (EMS) at P < 0.05 (χ^2 -test).

The Effect of Kinetin, Gibberellic Acid and Indole Acetic Acid on EMS-Induced Somatic Mutation and Recombination in *Drosophila melanogaster*

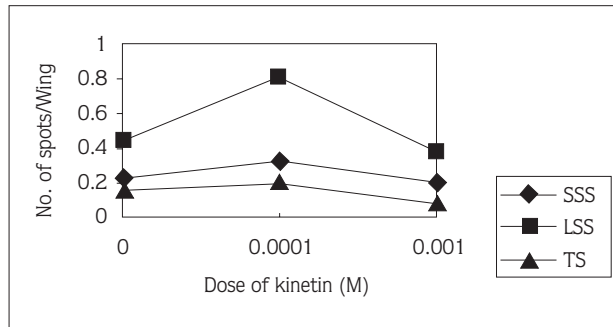
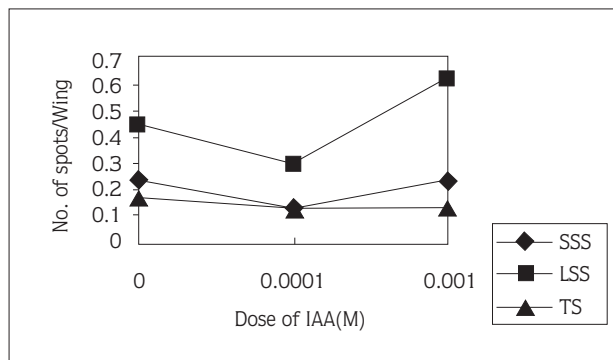
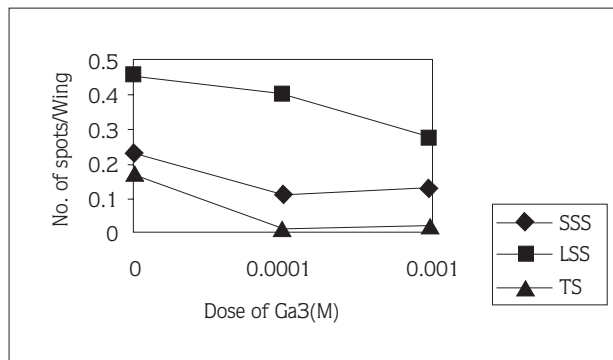


Figure 1. The effect of kinetin, GA₃ and IAA on mutations induced with EMS.



increase in the number of SSS and TS was observed at the same concentration ($P < 0.05$). A 10^{-3} M kinetin concentration decreased the number of TS ($P < 0.05$). Also 10^{-4} M and 10^{-3} M GA₃ reduced the number of all types of spot induced by EMS. Therefore GA₃ might suppress the EMS-induced gen mutations, chromosome deletion, non-disjunction and mitotic recombination.

When IAA was used, the number of LSS decreased significantly at 10^{-4} M concentration, but an increase was observed with 10^{-3} M IAA concentration. The observation of an increase in the number of single spots of the 10^{-4} M kinetin and 10^{-3} M IAA groups may indicate a mutagenic effect of plant growth hormones. In future studies, studying the mutagenic effect of the plant growth hormones studied here and others will be important for an explanation of this phenomenon.

In some previous studies, it has been reported that some chromosomal abnormalities, resulting from EMS and gamma rays treatment, were corrected or at least the effect of those mutagens was decreased with the application of GA_3 and IAA (6, 8-10). Here, it was confirmed that plant growth hormones, especially GA_3 , were also an effective bio-antimutagen in animals.

References

1. Kada, T., Inoue, T., Namiki, M. in: Klekowski, Jr. (Ed) Environmental mutagenesis, Carcinogenesis and Plant Biology, New York, Praeger Scientific, 1981, 132
2. Nakamura, Y. K., Kawai, K., Furukawa, H., Matsuo, T., Shimo, K., Tomita, I., Nakamura, Y., Suppressing effects of S-methyl methanethiosulfonate and diphenyl disulfide on mitomycin C-induced somatic mutation and recombination in *Drosophila melanogaster* and micronuclei in mice. Mutation Res., 385: 41-46. 1997.
3. Ramel, C., Alekperov, U., Ames, B., Kada, T., Wattenberg, L., Inhibitors of mutagenesis and their relevance with carcinogenesis. Mutation Res., 2: 47-65, 1986.
4. Graf, O., Würzler, F.E., Katz, A.J., Frei, H., Juon, H., Hall, C.B., Kale, P.G. Somatic mutation and recombination test in *Drosophila melanogaster*. Environ. Mutagen, 6: 153-188, 1984.
5. Sümer, Ş., Aşkın, T., Öztürk, N. EMS uygulanmış arpa tohumları üzerine gibberellik asitin etkisi hakkında bir ön çalışma. IX Ulusal Biyoloji Kongresi. Eylül 1988.
6. Devi, J., Deka, P.C. Hormonal repair of chromosomal damage caused through gamma rays and EMS in barley. J. Nuclear Agric. Biol. 12: 59-62. 1983.
7. Singh, C., Olegniczak, J., Hoppe, P., Patyne, H. The effect of growth regulators on sodium azide induced genetic damage in barley. Biologia Plantarum. (PRAHA) 22 (2): 91-96, 1988.
8. Mandal K.S., Basu, K.R. Cytological effects of single and combined treatments with X-rays, IAA and GA_3 in *Allium cepa*. Cytologia. 46: 133-139, 1981.
9. Alekperov, U.K., Mekhti-zadeh, E.R., Nagieva, G.N., Hormone regulation of mutation process. Sammar. Of 3th Symp. On Plant Growth Regulators. Varna, Bulgaria, P. 77, 1981
10. Sümer, Ş., Çelik T. Etil metan sülfonat uygulanmış arpa tohumları üzerine indol asetik asitin etkisi hakkında bir çalışma. Tr. J. of Biology. 19: 67-72, 1995.
11. Sato, T., Nishino, H., Nagase, H., Niikawa, M., Kito, H. Bio-antimutagen detection method with wing spot test by *Drosophila melanogaster*. Jpn. Toxicol. Environ. Health, 40(6): 498-505, 1994.

The Effect of Kinetin, Gibberellic Acid and Indole Acetic Acid on EMS-Induced Somatic Mutation and Recombination in *Drosophila melanogaster*

12. Sato, T., Nishino, H., Nagase, H., Niikawa, M., Kito, H. The effect of bio-antimutagens on chlorombucil and methotrexate using the wing spot test in *Drosophila melanogaster*. Jpn. Toxicol. Environ. Health, 41(6): 452-457, 1995.
13. Sato, T., Nishino, H., Nagase, H., Niikawa, M., Kito, H. The effect of several antipyretic analgesics on mitomycin C-induced mutagenesis using the wing spot test in *Drosophila melanogaster*. Jpn. J. Toxicol. Environ. Health, 42(2): 136-141, 1996.
14. Lindsley, D.L., Zimm, G.G. The Genom of *Drosophila melanogaster*. San Diego, Academic Press 1992
15. Bozcuk, A.N. The effects of some genotypes on the longevity of adult *Drosophila*. Exp. Geront., 13: 279-286, 1978.
16. Frei, H., Würgler F.E., Statistical methods to decide whether mutagenicity test data from *Drosophila* Assays indicate a positive, negative or inconclusive result. Mutation Res., 203: 297-308, 1988.
17. Segal, G.A., A Review of the genetic effects of ethyl methanesulfonate. Mutation Res., 134: 403-418, 1984.
18. Marec, F., Socha, R. Dose-response analysis of data obtained in the *Drosophila* wing spot test after ethyl methanesulfonate treatment. Acta Entomol. Bohemoslov., 84: 401-407, 1987.
19. Frei, H., Clements, J., Howe, D., Würgler, F.E. The Genotoxicity of the anti-cancer drug mitoxantrone in somatic and germ cells of *Drosophila melanogaster* Mutation Res., 279: 21-33, 1992.
20. Marec, F., Gelbic, I. High recombinagenic activities of three antiviral agents, adenin derivatives, in the *Drosophila* wing spot test. Mutation Res., 311: 305-317, 1994.