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Accumulation of Poly- β -Hydroxybutyrate in *Streptomyces* Species During Growth with Different Nitrogen Sources

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Abstract: The accumulation of poly- β -hydroxybutyric acid (PHB) in 27 *Streptomyces* isolates with different taxonomical properties was investigated. The amount of synthesized PHB was determined as crotonic acid by spectrophotometry. Eighty percent of the tested isolates accumulated PHB between 0.3 and 7.6% of dry mycelial weight. PHB was produced by a selectant of *Streptomyces* isolates in media containing different nitrogen sources (KNO_3 , glycine, peptone, proteose peptone, L-asparagine, tryptone and malt extract), each combined with 2 g/l glucose as the carbon source. The most effective PHB production was observed on glycine/glucose medium with a 7.6% dry mycelial weight. Nitrogen limiting conditions were inhibitory to *Streptomyces* growth, but stimulated PHB accumulation. Based on the morphological and biochemical test results, selected isolates were identified as *Streptomyces halstedii* (MU112), *S. anulatus* (MU117) and *S. rochei* (MU119).

Key Words: Poly- β -hydroxybutyric acid (PHB), *Streptomyces*, nitrogen sources

Farklı Azot Kaynakları ile Büyüme Sırasında *Streptomyces* Türlerinde Poly- β -Hydroxybutyratin Birikimi

Özet: Farklı taksonomik özelliklere sahip 27 *Streptomyces* izolatında Poly- β -hydroxybutyric asit (PHB) birikimi araştırıldı. Sentezlenen PHB miktarı krotonik asit olarak spektrofotometre ile belirlendi. Denenen izolatların %80'i PHB yi kuru miselyum ağırlığının %0.3-7.6'sı arasında akümüle etti. PHB, seçilmiş *Streptomyces* izolatları tarafından, farklı azot kaynakları (KNO_3 , glisin, pepton, proteoz pepton, L-asparagin, tripton ve malt ekstrakt) ve karbon kaynağı olarak da herbiri 2 g/l glukoz içeren besiyerlerinde üretildi. En etkili PHB üretimi kuru miselyum ağırlığının %7.6'sı ile glisin/glukoz ortamında gözlemlendi. Azotca sınırlandırılmış şartlar *Streptomyces* gelişimini inhibe ederken, PHB akümülesyonunu teşvik etmiştir. Morfolojik ve biyokimyasal test sonuçlarına göre seçilen izolatlar, *Streptomyces halstedii* (MU112), *S. anulatus* (MU117) ve *S. rochei* (MU119) olarak tanımlandı.

Anahtar Sözcükler: Poly- β -hydroxybutyric asit (PHB), *Streptomyces*, azot kaynakları

Introduction

At least 75 different genera of bacteria have been known to accumulate poly- β -hydroxybutyric acid (PHB) as intracellular granules. This polymer is synthesized under limited culture conditions and is used as a carbon and energy reserve. PHB production has most commonly been studied on micro-organisms belonging to the genera *Alcaligenes*, *Azotobacter*, *Bacillus* and *Pseudomonas* (1).

Aerobic, Gram-positive bacteria belonging to the genera *Rhodococcus*, *Nocardia* and *Streptomyces* are widely distributed in nature. Members of these bacteria are also able to synthesize and accumulate PHB when cultivated on different carbon sources under nitrogen

limiting conditions (2). However, the occurrence and formation of PHB have not been investigated to a significant extent.

In the present study, we investigated the ability of some *Streptomyces* isolates to accumulate PHB under different nitrogen-limiting conditions.

Materials and Methods

Bacterial isolates, media and growth conditions: Cultures of soil *Streptomyces* used in this study were obtained from the culture collection of Muğla University (MU). For screening PHB production ability and other nitrogen-limited conditions, isolates were grown in 50 ml

of nitrogen basal medium (3) in 250 ml Erlenmayer flasks. The medium was inoculated with 1 ml of activated homogeneous mycelial suspension and incubated at 30 °C in a rotary shaker (105 t/min) for 72 h. Growth was determined by measuring the dry weight of mycelial mass.

Experiments with nitrogen sources: Twenty-seven *Streptomyces* isolates were screened for PHB producing ability. For screening 2 g/l (NH₄)₂SO₄ was used as the nitrogen source. After this experiment, selected isolates were grown on different nitrogen sources to determine their production potential for PHB. Nitrogen basal medium was used as the main medium, but (NH₄)₂SO₄ was replaced by other nitrogen sources. These compounds were KNO₃, glycine, peptone, proteose peptone, L-asparagine, tryptone and malt extract at 1 g/l concentration. All media were adjusted to pH 6.8. Cells were grown in a rotary shaker (105 t/min) at 30 °C and harvested after 72 h of incubation by centrifugation at 10,000 g for 10 min. Then they were washed twice with distilled water before lysis in Na-hypochloride.

Analysis of PHB: For extraction of PHB the dry mycelial mass was lysed in Na-hypochloride at 37 °C for 12 h. Then PHB was extracted by chloroform. The chloroform extract was evaporated to dryness, and converted to crotonic acid by treatment with concentrated sulphuric acid. Absorbance spectra were determined by scanning the samples between 220 and 300 nm with a Shimadzu UV/VIS 1601 spectrophotometer (4).

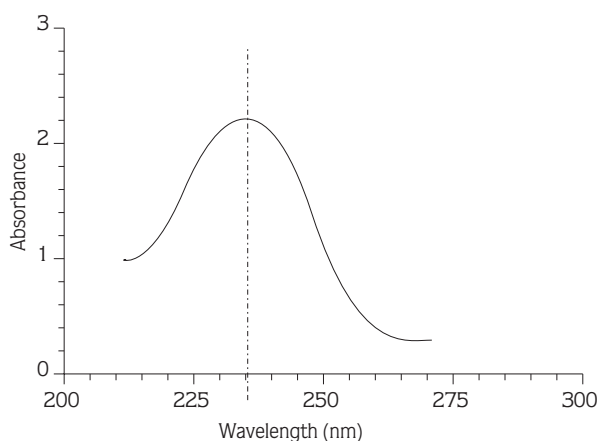


Figure 1. UV spectra of PHB isolated from strain MU112. (lane indicates λ_{max} = 235 nm).

Characterization of the *Streptomyces*: All isolates were cultivated on glucose/yeast extract/malt extract agar (ISP 2). Some diagnostic characters of highly productive *Streptomyces* isolates were determined by following the directions given in the probabilistic identification matrix (3) and Bergey's Manual of Systematic Bacteriology (5). A Willcox probability (P) matrix was used to assign and identify the isolates where P scores of 0.8 and above indicated a positive identification (6).

Results and Discussion

A large proportion (80%) of the tested soil *Streptomyces* isolates was able to produce PHB as energy reserve material (Fig. 1). However, the actual amounts of PHB accumulation (percent of cell DW) were generally lower for the industrial applications than for the other bacterial groups, which are known to produce PHB.

Based on the screening data, the isolates MU112, MU117 and MU119, which represent different taxonomical groups, were selected for experiments with various nitrogen sources. The selected isolates were characterized by some morphological and physiological properties presented in the Table. According to the test results isolates MU112, MU117 and MU119 were identified as *Streptomyces halstedii*, *S. anulatus* and *S. rochei* respectively.

The accumulation of PHB varied with taxonomical properties of *Streptomyces* isolates and nitrogen type in the medium. The main characteristics of the PHB

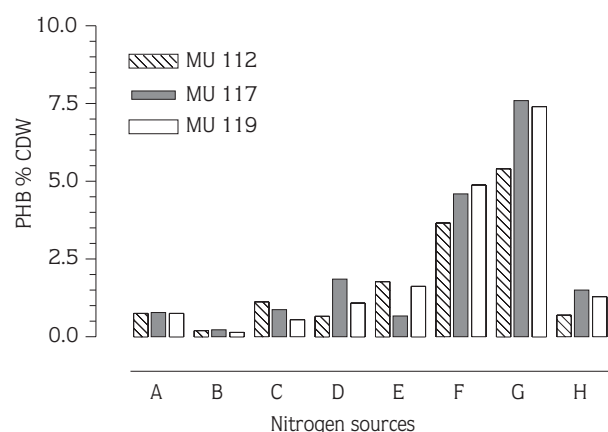


Figure 2. PHB accumulation by some *Streptomyces* grown on different nitrogen sources. A: (NH₄)₂SO₄, B: Asparagine, C: Malt extract, D: Peptone, E: Proteose peptone, F: Tryptone, G: Glycine, H: KNO₃.

Character	MU112	MU117	MU119
Spore chain Rectiflexibiles	+	+	+
Spore chains Spirales	-	-	-
Spore mass grey	+	+	+
Diffusible pigment produced	+	-	-
Melanin on pepton/yeast/iron agar	-	-	-
Antibiosis against			
<i>Bacillus subtilis</i> ATCC 6633	-	-	-
<i>Staphylococcus aureus</i> ATCC 6538	-	-	-
<i>Candida albicans</i> ATCC 10239	-	+	-
<i>Escherichia coli</i> ATCC 11230	-	-	-
Growth at 45 °C	-	-	+
Growth with (% w/v)			
NaCl (7.0)	+	+	+
Sodium azide (0.01)	+	+	+
Phenol (0.1)	-	+	+
DNase	+	+	+
Nitrate reduction	-	-	+
Utilization of:			
Citrate	-	-	-
Malonate	-	-	-
Tartrate	-	-	-
Oxalate	-	-	-
Willcox Probability (P)	0.96	0.40	0.50

Table. Some taxonomical properties of selected PHB producing *Streptomyces*.

producing *Streptomyces* were the grey colour of mature sporulated aerial mycelium, rectiflexibiles type of sporophore and negative melanoid pigment production.

Maximum PHB content was found in MU117 when glycine (7.6%) or tryptone (4.6%) was used as the nitrogen source. The PHB content was much lower when asparagine and $(\text{NH}_4)_2\text{SO}_4$ were used as the nitrogen source. In a previous study, the PHB content of the less active strain of *Rhizobium meliloti* was reported to be high when grown on KNO_3 and glycine as nitrogen sources (7). PHB accumulations by selected *Streptomyces* isolates after cultivation on different nitrogen sources are shown in Figure 2. Nitrogen-limiting conditions were inhibitory to mycelial growth, but stimulated PHB production (Fig. 3).

The PHB producing efficiency of these isolates under different nitrogen stress conditions ranged from 0.3 to

7.6% of their dry mycelial weight. In earlier experiments, similar to our observations, researchers reported very low PHB formation by *Streptomyces* (8).

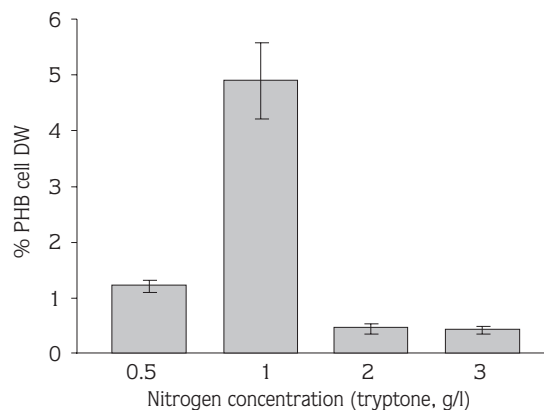


Figure 3. Effect of nitrogen concentration on PHB production.

References

- Anderson, A.J., Dawes, E.A. Occurrence, metabolism, metabolic role and industrial uses of bacterial polyhydroxyalkanoates. *Microbiol Rev.*, 54: 450-472, 1990.
- Alvarez, H.M., Kalscheuer, R., Steinbüchel, A. Accumulation of storage lipids in species of *Rhodococcus* and *Nocardia* and effect of inhibitors and polyethylene glycol. *Fett-Lipid*, 99: 7: 239-246, 1997.

3. Williams, S.T., Goodfellow, M., Wellington, E.M.H., Vickers, J.C., Alderson, G., Sneath, P.H.A., Sackin, M.J., Mortimer, A.M. A probability matrix for identification of some Streptomyces. J. Gen. Microbiol., 129: 1815-1830, 1983.
4. Gerhardt, P., Murray, R.G.E., Wood, W.A., Krieg, N.R. Methods for General and Molecular Bacteriology, Washington, DC., 1994 American Society for Microbiology, 628.
5. Williams, S.T., Goodfellow, M., Alderson, G. Genus *Streptomyces* (Waksman & Hanrici 1943) 339^{AL}. In *Bergey's Manual of Systematic Bacteriology*, Edited by Williams, S.T., Sharpe, M.E., & Holt, J.G. vol. 4, Baltimore, 1989, Williams & Wilkins, pp. 2452-2492.
6. Willcox, W.R., Lapage, S.P., Bascomb, S., Curtis, M.A. Identification of bacteria by computer: theory and programming. J. Gen. Microbiol., 77: 317-330, 1973.
7. Bonartseva, G.A., Myshkina, V.L., Zagreba, E.D. Poly- β -hydroxybutyrate content in cells of various *Rhizobium* species during growth with different carbon and nitrogen sources. Microbiol., 63: 1, 45-48, 1994.
8. Kannan, L.V., Rehacak, Z. Formation of Poly- β -hydroxybutyrate by Actinomycetes. Ind. J. Biochem., 7: 126-129, 1970.