

1-1-2020

## Physiological and pharmacological roles of melatonin in the pathophysiological components of cellular injury after ischemic stroke

ERTUĞRUL KILIÇ

BERRAK ÇAĞLAYAN

MUSTAFA CAGLAR BEKER

Follow this and additional works at: <https://journals.tubitak.gov.tr/medical>



Part of the [Medical Sciences Commons](#)

---

### Recommended Citation

KILIÇ, ERTUĞRUL; ÇAĞLAYAN, BERRAK; and BEKER, MUSTAFA CAGLAR (2020) "Physiological and pharmacological roles of melatonin in the pathophysiological components of cellular injury after ischemic stroke," *Turkish Journal of Medical Sciences*: Vol. 50: No. 10, Article 9. <https://doi.org/10.3906/sag-2008-32>

Available at: <https://journals.tubitak.gov.tr/medical/vol50/iss10/9>

This Article is brought to you for free and open access by TÜBİTAK Academic Journals. It has been accepted for inclusion in Turkish Journal of Medical Sciences by an authorized editor of TÜBİTAK Academic Journals. For more information, please contact [academic.publications@tubitak.gov.tr](mailto:academic.publications@tubitak.gov.tr).

## Physiological and pharmacological roles of melatonin in the pathophysiological components of cellular injury after ischemic stroke

Ertuğrul KILIÇ<sup>1,2,\*</sup>, Berrak ÇAĞLAYAN<sup>2,3</sup>, Mustafa Çağlar BEKER<sup>1,2</sup>

<sup>1</sup>Department of Physiology, School of Medicine, İstanbul Medipol University, İstanbul, Turkey

<sup>2</sup>Regenerative and Restorative Medicine Research Center (REMER), Research Institute for Health Sciences and Technologies (SABITA), İstanbul Medipol University, İstanbul, Turkey

<sup>3</sup>Department of Medical Biology, International School of Medicine, İstanbul Medipol University, İstanbul, Turkey

Received: 04.08.2020 • Accepted/Published Online: 21.09.2020 • Final Version: 03.11.2020

**Abstract:** Apart from its metabolic or physiological functions, melatonin has a potent cytoprotective activity in the physiological and pathological conditions. It is synthesized by the pineal gland and released into the blood circulation but particularly cerebrospinal fluid in a circadian manner. It can also easily diffuse through cellular membranes due its small size and lipophilic structure. Its cytoprotective activity has been linked to its potent free radical scavenger activity with the desirable characteristics of a clinically- reliable antioxidant. Melatonin detoxifies oxygen and nitrogen-based free radicals and oxidizing agents, including the highly toxic hydroxyl- and peroxynitrite radicals, initiating cellular damage. However, the cytoprotective activity of melatonin is complex and cannot be solely limited to its free radical scavenger activity. It regulates cellular signaling pathways through receptor- dependent and independent mechanisms. Most of these downstream molecules, such as PI3K/AKT pathway components, also contribute to the cytoprotective effects of melatonin. In this term, melatonin is a promising molecule for the treatment of neurodegenerative disorders, such as ischemic stroke, which melatonin reduces ischemic brain injury in animal models of ischemic stroke. It regulates also circadian rhythm proteins after ischemic stroke, playing roles in cellular survival. In this context, present article summarizes the possible role of melatonin in the pathophysiological events after ischemic stroke.

**Key words:** Ischemic stroke, melatonin, free radicals, apoptosis, signaling, circadian rhythm proteins

### 1. Introduction

Stroke is one of the leading causes of death and long-term disability worldwide [1] with increasing incidence rates in low and middle-income countries [2]. Disability due to a stroke incidence has significant emotional, social and economic burden on the patients and their society. It was suggested that deaths due to stroke cases will reach to 12 million and number of patients surviving from a stroke incidence will reach to about 70 million by the year 2030 [3].

Stroke cases are categorized into two groups: hemorrhagic and thrombotic (or ischemic) stroke. In hemorrhagic stroke, intracerebral or subarachnoid hemorrhage occurs in the brain due to the rupture of a cerebral blood vessel, but this type of stroke is less common (about 15% of all stroke cases in USA and Europe) worldwide [4,5]. On the other hand, ischemic stroke contributes to the majority of stroke cases and occurs when a cerebral artery is occluded by thrombus or

thromboembolism, meaning that a blood clot may occur and cause a block in a small cerebral artery or a blot clot formed in a large vessel lodges in a small vessel in the brain. Due to the occlusion of the cerebral artery, blood supply to a certain brain region is restricted. Therefore, this event is called ischemia or ischemic stroke.

Despite being a global problem with major social and economic burden on the society, treatment strategies are limited, and prevention measures are not enough [1]. Currently, patients survived from an ischemic stroke are treated with tissue plasminogen activator (tPA) that helps to relieve the obstruction in the blood vessels [6]. However, tPA has a very short therapeutic window which makes it impossible to use in the majority of the patients because of unfavorable actions of thrombolytics. Therefore, researchers have been in the search for a treatment option that can be used in patients who were diagnosed after the end of the therapeutic window of tPA or add-on treatment with thrombolytics. In this sense, it was shown that

\* Correspondence: kilic44@yahoo.com

melatonin also reverses unfavorable actions of tPA [7] and, we will focus on the novel findings for the use of melatonin in ischemic brain injury in the present article.

The pathophysiologic mechanisms underlying ischemic stroke or ischemia/reperfusion injury in humans include loss of the prooxidant/antioxidant balance, excitotoxicity and related increase in intracellular  $Ca^{++}$  levels, dysfunction in mitochondrial processes, increased neuroinflammation and eventually apoptotic neuronal cell death [8].

## 2. Justification of use of melatonin in stroke treatment

The studies reporting the antioxidant effects of melatonin justified its use for treatment of ischemic injury in animal models [9]. Thereafter, further research provided evidence that melatonin is able to alleviate several of the detrimental effects of stroke-related pathophysiologic processes [10,11]. In addition, melatonin was shown to protect the integrity of blood-brain barrier [12] and vascular function in the brain [13], bind and neutralize heavy metals [14] promote neuronal survival [15,16], and functional recovery [17] with minimal, if any, side effects even in high doses [18]. Moreover, melatonin exerts neurological protection by reducing the cerebral inflammatory response, cerebral edema and brain-blood barrier permeability after ischemic stroke [19].

Although melatonin can be synthesized in a number of tissues, it is believed that the main production center may be the pineal gland due to the fact that melatonin's protective effects are severely diminished by pinealectomy as reported in experimental model of stroke [20]. Melatonin synthesis in the pineal gland occurs in a circadian manner, in which the melatonin levels peak in the night-time, whereas in the day-time melatonin production is inhibited [21]. Secreted melatonin is believed to modulate the circadian clock mechanisms through the ubiquitin-proteasome signaling pathway. Several gene transcripts [22] and proteins [23] have been identified to be expressed in a circadian fashion. Of these genes, the six core clock genes are known as *Per1* and *Per2*, *Cry1* and *Cry2*, *Clock* and *Bmal1* [24,25]. We have reported that melatonin regulates *Bmal1* expression under normal conditions in vitro after hypoxia and in vivo after ischemic stroke [26]. Interestingly, in parallel with the highest blood concentrations of melatonin, the ischemic stroke incidence is lowest at the midnight hours in human [27]. In this term, we have indicated that tolerance to ischemic injury changes according to the time of day in which the injury occurs and the underlying mechanism of this tolerance includes circadian clock genes, specifically *Bmal1*, which is also regulated by the phosphorylation of AKT signaling pathway [24]. Consistently, *Bmal1* expression is enhanced following melatonin treatment following ischemic brain injury in mice and this increase

is blocked when the survival kinase AKT inhibitors are present [26]. In addition to the transcriptional control of *Bmal1* gene, melatonin may also regulate clock genes by stabilizing the protein through the inhibition of the ubiquitin-proteasome system [28]. It has been speculated that melatonin can act as a proteasome inhibitor and we showed that melatonin inhibits the proteasome machinery by downregulating *Nedd4-1* E3 ligase expression [29]. Collectively, these data suggest that melatonin may still have unelucidated roles in the body that promotes endogenous recovery systems.

## 3. Melatonin's antioxidant effect

The experimental models in rodents focusing on the antioxidant effect of melatonin in stroke pathophysiology mainly involved in the ischemia/reperfusion. In this model, generally transiently middle cerebral artery occlusion (MCAo) is performed and melatonin is usually administered either at the onset of ischemia or reperfusion [8]. In the ischemia/reperfusion injury, a blood vessel is obstructed and blood supply to a certain brain region is restricted, causing immediate apoptotic and necrotic cell death in the ischemic core. The surrounding tissue, called the penumbra, has relatively higher levels of blood supply compared to the ischemic core, however, apoptotic cell death through the complex interplay of several mechanisms can be observed in the penumbra even several days after ischemic injury [8]. In order to resupply the ischemic brain tissue with oxygen, hyperbaric (HBO) and normobaric oxygen (NBO) treatments gained considerable interest due to the possibility of oxygen to diffuse through the blood-brain barrier to reach the injured brain tissue. However, conflicting literature exists for HBO treatment in transient or permanent MCAO models possibly because of increased free radical production upon effect of oxygen [30]. Even detrimental effects of HBO treatment were reported [31], but it should be kept in mind that the underlying reason for conflicting results may be partly because of the different timing of HBO treatment. Conversely, NBO therapy is an inexpensive and easy-to-access strategy that can be administered by simple facemasks. The favorable effects of NBO on the infarct volume and cell death were demonstrated in different models of brain injury [32–35]. However, the NBO treatment in these studies is usually started during the ischemia period or immediately at the onset of reperfusion. Although NBO can be beneficial in these models, this therapy can be only translated to patients who were admitted to hospitals with stroke symptoms that last for less than 12 h. In the light of these results, our group investigated the use of NBO treatment during reperfusion [12]. We also evaluated the combination of melatonin with NBO treatment. Our results indicated that melatonin potentiated the protective effect of NBO therapy in terms

of infarct volume, brain swelling, neurological deficit score and DNA fragmentation [12].

Even though the most important step in rescuing the cells' from apoptotic death in the penumbra is to remove the obstruction and resupply the blood flow (i.e. reperfusion), this results in oxidative damage due to the excess production of free radicals. The cells of the central nervous system (CNS) are already exposed to high amounts of free radicals in the physiological state. The adult human brain uses about 20% of the total oxygen intake even though it weighs only about 2% of the total body weight [36]. Not only brain produces the highest number of free radicals compared to any other organ in the body, but also it has high levels of polyunsaturated fatty acids which make it more prone to oxidative stress. Under normal conditions, the production of free radicals and antioxidant enzymes are kept in a delicate balance. If an imbalance occurs in favor of oxidants as a result of brain injury, the ability of the CNS cells to neutralize oxidants using endogenous antioxidant systems is overwhelmed and eventually inflammation and apoptotic cell death are observed.

It is well-known that the free radicals including reactive oxygen species (ROS) and reactive nitrogen species (RNS), including superoxide anions ( $O_2^{\bullet-}$ ), hydrogen peroxide ( $H_2O_2$ ), hydroxyl radicals ( $HO^{\bullet}$ ), nitric oxide ( $\bullet NO$ ) and peroxynitrite ( $ONOO^-$ ) are mainly produced during the reperfusion phase of an ischemic stroke. The sources of these free radicals are diverse. For instance, nitric oxide is a signaling molecule that is involved in vasodilatation, neurotransmission and blood pressure maintenance and is synthesized by NO synthase (NOS) enzymes mainly in a calcium-dependent manner [37]. Three types of NOS enzymes were characterized as neuronal NOS (nNOS), inducible NOS (iNOS) and endothelial NOS (eNOS) in the central nervous system [38]. During ischemic brain injury, excess nitric oxide is generated, which in turn results in lipid peroxidation, energy depletion, and formation of highly neurotoxic oxidizing agent  $ONOO^-$  [39]. Although the nitric oxide synthesized by the eNOS at the early phases of ischemic stroke is believed to provide protective effects, NO synthesis by iNOS and nNOS exacerbates the injury by activating the inflammatory mechanisms [40].

It has become clear that excessive free radical generation is a critical pathophysiological step in ischemic injury and regulates several other steps. This prompted researchers to investigate the possible use of chemical or biological molecules to reduce the generation or accumulation of free radicals. We and others have tested several antioxidant molecules, such as glutathione [41], vitamin E [42], vitamin C [43] or melatonin [20]. To the best of our knowledge, melatonin is the only antioxidant whose metabolites also have antioxidant capacities. In

fact, the direct and indirect capacity of melatonin and its metabolites (c3OHM, AFMK and AMK) to scavenge free radicals is called the "antioxidant cascade" [44]. As the members of the antioxidant cascade of melatonin are all free radical scavengers, the detoxification capacity of melatonin was predicted to be up to 10 times more than any other antioxidant molecule [45]. Therefore, antioxidant capacity of melatonin has been studied in many ischemia/reperfusion injury models, including brain, kidney or heart. In fact, melatonin can interact with and detoxify the free radicals by donating an electron or a hydrogen atom in the injury models. In addition to its free radical scavenger activity, melatonin was also shown to upregulate antioxidant enzymes, such as superoxide dismutase (SOD), catalase or glutathione reductase (GR) [46]. Our group evaluated the effect of melatonin on the production of nitric oxide and our results demonstrate that melatonin treatment significantly downregulates nNOS and iNOS after ischemic brain injury [7,47].

#### 4. Melatonin in mitochondria

Considering the higher melatonin concentrations compared to blood levels were measured in rat liver mitochondria [48], it is not surprising to assume that high melatonin levels are required to protect the mitochondrial DNA from the continuous production of reactive oxygen species by the oxidative phosphorylation [49]. Because of its lipophilic structure, melatonin can easily pass through the biological membranes and accumulate in organelles, such as mitochondria and nuclei. Since the discovery of melatonin in bacteria and chloroplasts, it has been speculated that melatonin synthesis can also occur in the mitochondria [50]. Furthermore, enzymes (although not all) required for the synthesis of melatonin were detected in oocyte mitochondria [51]. These results strongly suggest that melatonin synthesized in the mitochondria is used as direct free radical scavenger and indirect antioxidant enzyme regulator in this organelle. Moreover, in a recent study, mitochondrial melatonin was shown to be secreted into the cytosol where it can bind to the receptors on the surface of the mitochondria [52]. It is proposed that this feature gives another advantage to melatonin in its role in mitochondrial homeostasis. This involves the restoration of the activity and expression of complexes I and IV which are decreased during ischemic injury. This in turn, reduces the electron leakage and prevents further damage to the organelle.

In addition, increased free  $Ca^{++}$  levels following ischemic brain injury are also responsible for mitochondrial dysfunction and formation of further free radicals. If these toxic effects are not neutralized, ATP production would be severely affected. Intriguingly, Xu et al. implicated that melatonin can control free  $Ca^{++}$  movement in the

cytoplasm and protects the homeostasis of mitochondria [53].

### 5. Melatonin's antiapoptotic effect

Loss of blood supply during ischemic brain injury depletes the cellular energy and results in the release of glutamate neurotransmitter into the extracellular space [8]. As a result, the transmembrane glutamate receptors (such as NMDA, AMPA or kainic acid receptors) and several channel and transporter proteins (such as TRPM2, TRPM7, NCX, ASICs, CaV1.2) are overactivated. Activation of these receptors is followed by an excessive Ca<sup>++</sup> influx, either due to the release from mitochondria and endoplasmic reticulum or due to activation of plasma membrane proteins and lead to apoptotic cell death [54]. Of these receptors, overactivation of postsynaptic NMDA receptors by glutamate results in increased Ca<sup>++</sup> load in the cytosol and mitochondria [55]. In this term, we used a combination treatment with an NMDA receptor inhibitor, memantine, and melatonin in a MCAo model and our results indicate that administration of melatonin/memantine combination significantly reduces the infarct volume, while improving the vascular leakage [56]. Furthermore, electrophysiological studies revealed that melatonin depresses NMDA receptor activity in the brain, possibly due to reduced nNOS activity [56,57].

In an attempt to rescue from Ca<sup>++</sup> overload, the cells in the injured brain tissue try to reduce the high cytosolic Ca<sup>++</sup> levels by storing Ca<sup>++</sup> ions into mitochondria or endoplasmic reticulum. However, when Ca<sup>++</sup> levels are increased in the mitochondrial matrix, free radical production is enhanced. Increased free radical accumulation disrupts mitochondrial membrane, results in permeabilization and depolarization followed by the release of proapoptotic molecules, such as cytochrome c and apoptosis inducing factor (AIF) into the cytosol [58,59]. Andrabi et al. suggested that melatonin inhibits the cytochrome c release and decreases DNA damage in transient MCAO in rats [60]. Our results proved that melatonin improves neuronal survival by caspase-3 inhibition [16]. In addition, we showed that melatonin inhibits antiapoptotic Bcl-xL, while promoting the expression of proapoptotic Bax [12]. Downregulation of survival kinases in the injured brain tissue also contributes to apoptotic cell death. It is well-known that ischemia/reperfusion inhibits PI3K/AKT pathway. Our planar immunoassay analyses revealed that melatonin treatment results in the increased AKT phosphorylation especially at the Thr308 site of the activation loop via PDK1 and PTEN as well as decreased GSK-3 $\alpha/\beta$ , and p53 phosphorylations [19], suggesting that neuroprotective activity of melatonin directly involves the activation of survival signaling pathways. Additionally, we have observed that melatonin

treatment phosphorylates AMPK $\alpha$ , which is particularly activated by the reduced intracellular energy. AMPK $\alpha$  drives the cell to a catabolic state which this molecule mobilizes alternative energy sources, such as fatty acid oxidation, in order to supply ATP in the condition of ischemic stroke, suggesting also that melatonin may activate alternative collateral survival pathways [19].

### 6. Melatonin's antiinflammatory effect

It has been shown that one of the early mechanisms of ischemic injury is the release of inflammatory cytokines, such as IL1 $\beta$ , IL6 and TNF $\alpha$  [61]. Following an ischemic attack in the brain, the free radicals are generated in high amounts. This excessive free radical production especially during the reperfusion phase also contributes to the disruption of the integrity of the blood-brain barrier and stimulates the infiltration of lymphocytes, neutrophils, monocytes, T cells, and macrophages to the injury site [62]. Simultaneously, resident microglial cells are activated, change their morphology (deramification) and start to release proinflammatory cytokines. Microglial activation following ischemia also results in increased proliferation and accumulation in the penumbra region. Microglial activation seems to have a dual role in the pathophysiology of stroke. During the course of ischemia/reperfusion, microglial cells are believed to switch from an antiinflammatory state to proinflammatory phenotype [63]. Melatonin treatment was shown to inhibit the proinflammatory shift of microglial cells through the regulation of SIRT1 and STAT3 [64,65]. Moreover, TLR4 activates NF- $\kappa$ B after stroke, resulting in the secretion of inflammatory molecules (IL1 $\beta$ , IL6 and TNF $\alpha$ ). Melatonin decreases the secretion of inflammatory mediators by downregulating NF- $\kappa$ B, while promoting Nrf2 upregulation [66].

### 7. Roles of melatonin receptors in stroke treatment

Studies indicated increased melatonin levels can be observed as early as in the first 10 minutes of intraperitoneal or subcutaneous application, indicating that melatonin can easily pass through the blood-brain barrier [67]. Melatonin is also able to diffuse through cellular and organelle membranes due its small size and lipophilic structure [68]. On the other hand, melatonin has two G protein-coupled transmembrane receptors; MT1 and MT2 (earlier names Mel1a and Mel1b) which are ubiquitously found in almost all cells in the body [69]. It has been proposed that melatonin exerts its neuroprotective effects on the brain by both receptor-independent or receptor dependent mechanisms [70]. Models of acute and chronic ischemia have been used to investigate the role of MT1/MT2 receptors in ischemic brain injury by using melatonin receptor agonists, such as ramelteon [71,72], agomelatine

[73,74] or Neu-P11 [75]. Activation of MT1/MT2 receptors by the aforementioned agonists reduced infarct volume through the regulation of different signaling molecules. In 2010, it was reported that melatonin administration promotes the expression of MT2 in the ischemic tissue and due to this increase, the authors suggested that MT2 may partially mediate the effects of melatonin [76]. Chern et al. reported that chemical antagonists of MT2 could reverse the protective effect of melatonin in a transient brain ischemia model [77]. However, in a previous report where we used a transient focal cerebral ischemia model in MT1/MT2 knockout mice, we showed that melatonin treatment significantly decreased brain damage, suggesting that it does not require the interplay of these receptors for exerting neuroprotective effects [47]. In the light of these results, we hypothesized that although the activation of MT1/MT2 receptors protects the brain from ischemic injury, melatonin does not require MT1/MT2 receptor activation to exert neuroprotection. However, it should be noted that melatonin has another membrane-associated receptor, MT3 (also known as quinone reductase 2, QR2), found in the cells of liver, kidney, heart, lung, intestine, adipose tissues or in brain cells [78,79] and whether it is involved in melatonin's antioxidant, antiapoptotic or antiinflammatory activity should be further investigated. Interestingly, MT3 knockout mice were less susceptible to menadione toxicity [80], suggesting that the inhibition of this receptor may have protective effects. Moreover, knockdown of MT3 by RNA interference *in vitro* resulted in enhanced expression of antioxidant enzymes [81], while overexpression of MT3 resulted in excessive production of reactive oxygen species [82]. It was shown that melatonin is able to inhibit this receptor at nanomolar levels in which antioxidant effects were documented [83]. Therefore, it is tempting to speculate that MT3 inhibition by melatonin is involved in melatonin's protective effects on the pathophysiological outcomes of brain injury not only by increasing the expression of antioxidant enzymes, but also by providing resistance to oxidative stress.

Moreover, the nuclear receptors RZR/ROR from the retinoic acid receptor superfamily were proposed as nuclear binding sites for melatonin [84]. RZR/ROR expression was demonstrated in the brain, and in the pineal gland [69]. ROR receptors were shown to induce the expression of several clock genes, including *Clock*, *Cry* or *Bmal1* [85–87] by binding to retinoic acid-related orphan receptor response elements (ROREs) in the promoter region. In parallel with these data, cyclic expression of ROR mRNAs were noted in different tissues, suggesting a circadian function possibly under the control of melatonin. However, whether these receptors play a part in the protective mechanisms induced by melatonin should be further investigated.

## 8. New roles for melatonin: regulation of circular RNA in ischemic injury

Recently, noncoding RNAs including microRNAs, long noncoding RNAs, and circular RNAs have gained considerable attention as regulatory molecules. Recent studies indicated that circular RNAs (circRNAs) are found abundantly in the brain and are involved in the embryonic development [88]. CircRNAs are enriched in specific brain regions such as cerebellum, cortex, striatum, olfactory bulbs, and hippocampus. Since they are circular, these types of noncoding RNAs are more resistant to digestion. CircRNAs can be made from introns, coding or noncoding exons, or from both exons and introns by a process called “back-splicing” [89]. Zhang et al. suggested that because the circRNAs are formed by the circularization of skipped exons, formation of circRNA can cause the downregulation of its parental gene by using up the pre-mRNA molecules [90]. In addition to their regulatory roles in the brain, circRNAs are also associated with neurological disorders, including, but not limited to, ischemia/reperfusion injury, traumatic brain injury, and Alzheimer's disorder [91]. In a study performed in acute ischemic stroke patients to profile the changes in circRNA, 3 circRNAs have been proposed as diagnostic and predictive biomarkers for stroke [92]. Significantly altered circRNAs in a transient MCAO model in C57BL/6J mice were characterized, and bioinformatics data suggested that all these circRNAs possess binding sites for microRNAs [93]. However, the exact pathophysiological mechanisms that they have a role in are not fully elucidated. It has been predicted that melatonin may be involved in the regulation of circRNAs as the circRNA *proliferate* in the pineal gland has been altered in a mouse model of Alzheimer's disease [94]. In fact, a recent study reported that melatonin exerts protective effects through the regulation of CircRIC3/miR-204-5p/DPP4 signaling in calcific aortic valve disease [95]. Therefore, we hypothesized that melatonin also regulates circRNAs in ischemia/reperfusion injury, however, these circRNAs should be further investigated in future studies.

## 9. Prophylactic use in high risk individuals

Moreover, we reported that lower endogenous melatonin concentrations were associated with increased injury after transient ischemic stroke in pinealectomized rats and when those deprived animals were given exogenous melatonin, injury size could be reduced [20]. It has been shown that melatonin levels are reduced in elderly compared to young adults; therefore, aged population is more prone to serious ischemic injury [96]. In addition, accumulation of other risk factors, such as cardiovascular disorders, diabetes, obesity or hypertension with ageing increases the risk of ischemic injury incidence [97,98]. In animal models, we

demonstrated that prophylactic uses, and delayed uses of melatonin successfully protects the brain from the ischemic injury [13,17]. In conclusion, prophylactic doses could be considered in the elder population in order to compensate for the reduced melatonin levels due to the calcification of pineal gland and to promote the endogenous repair

mechanisms against stroke or other neurodegenerative diseases.

### Conflict of interest

The authors declare no conflict of interest related to this paper.

### References

1. Flynn RW, MacWalter RS, Doney AS. The cost of cerebral ischaemia. *Neuropharmacology* 2008; 55 (3): 250-256. doi: 10.1016/j.neuropharm.2008.05.031
2. Feigin VL, Lawes CM, Bennett DA, Barker-Collo SL, Parag V. Worldwide stroke incidence and early case fatality reported in 56 population-based studies: a systematic review. *The Lancet Neurology* 2009; 8 (4): 355-369. doi: 10.1016/S1474-4422(09)70025-0
3. Feigin VL, Forouzanfar MH, Krishnamurthi R, Mensah GA, Connor M et al. Global and regional burden of stroke during 1990-2010: findings from the Global Burden of Disease Study 2010. *Lancet* 2014; 383 (9913): 245-254. doi: 10.1016/s0140-6736(13)61953-4
4. Rosamond W, Flegal K, Friday G, Furie K, Go A et al. Heart disease and stroke statistics--2007 update: a report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. *Circulation* 2008; 115 (5): e69-e171. doi: 10.1161/CIRCULATIONAHA.106.179918
5. Wolfe CD, Giroud M, Kolominsky-Rabas P, Dundas R, Lemesle M et al. Variations in stroke incidence and survival in 3 areas of Europe. *European Registries of Stroke (EROS) Collaboration. Stroke* 2000; 31 (9): 2074-2079. doi: 10.1161/01.str.31.9.2074
6. Zivin JA, Fisher M, DeGirolami U, Hemenway CC, Stashak JA. Tissue plasminogen activator reduces neurological damage after cerebral embolism. *Science* 1985; 230 (4731): 1289-1292. doi: 10.1126/science.3934754
7. Kilic E, Kilic U, Reiter RJ, Bassetti CL, Hermann DM. Tissue-plasminogen activator-induced ischemic brain injury is reversed by melatonin: role of iNOS and Akt. *Journal of Pineal Research* 2005; 39 (2): 151-155. doi: 10.1111/j.1600-079X.2005.00228.x
8. Dirnagl U, Iadecola C, Moskowitz MA. Pathobiology of ischaemic stroke: an integrated view. *Trends in Neurosciences* 1999; 22 (9): 391-397. doi: 10.1016/s0166-2236(99)01401-0
9. Hardeland R, Reiter RJ, Poeggeler B, Tan DX. The significance of the metabolism of the neurohormone melatonin: antioxidative protection and formation of bioactive substances. *Neuroscience and Biobehavioral Reviews* 1993; 17 (3): 347-357. doi: 10.1016/s0149-7634(05)80016-8
10. Reiter RJ, Tan DX, Gitto E, Sainz RM, Mayo JC et al. Pharmacological utility of melatonin in reducing oxidative cellular and molecular damage. *Polish Journal of Pharmacology* 2004; 56 (2): 159-170.
11. Reiter RJ, Tan DX, Leon J, Kilic U, Kilic E. When melatonin gets on your nerves: its beneficial actions in experimental models of stroke. *Experimental Biology and Medicine* 2005; 230 (2): 104-117. doi: 10.1177/153537020523000205
12. Beker MC, Caglayan AB, Kelestemur T, Caglayan B, Yalcin E et al. Effects of normobaric oxygen and melatonin on reperfusion injury: role of cerebral microcirculation. *Oncotarget* 2015; 6 (31): 30604-30614. doi: 10.18632/oncotarget.5773
13. Kilic E, Kilic U, Reiter RJ, Bassetti CL, Hermann DM. Prophylactic use of melatonin protects against focal cerebral ischemia in mice: role of endothelin converting enzyme-1. *Journal of Pineal Research* 2004; 37 (4): 247-251. doi: 10.1111/j.1600-079X.2004.00162.x
14. Romero A, Ramos E, De Los Rios C, Egea J, Del Pino J et al. A review of metal-catalyzed molecular damage: protection by melatonin. *Journal of Pineal Research* 2014; 56 (4): 343-370. doi: 10.1111/jpi.12132
15. Kilic E, Hermann DM, Isenmann S, Bahr M. Effects of pinealectomy and melatonin on the retrograde degeneration of retinal ganglion cells in a novel model of intraorbital optic nerve transection in mice. *Journal of Pineal Research* 2002; 32 (2): 106-111. doi: 10.1034/j.1600-079x.2002.1823.x
16. Kilic E, Kilic U, Yulug B, Hermann DM, Reiter RJ. Melatonin reduces disseminate neuronal death after mild focal ischemia in mice via inhibition of caspase-3 and is suitable as an add-on treatment to tissue-plasminogen activator. *Journal of Pineal Research* 2004; 36 (3): 171-176. doi: 10.1046/j.1600-079x.2003.00115.x
17. Kilic E, Kilic U, Bacigaluppi M, Guo Z, Abdallah NB et al. Delayed melatonin administration promotes neuronal survival, neurogenesis and motor recovery, and attenuates hyperactivity and anxiety after mild focal cerebral ischemia in mice. *Journal of Pineal Research* 2008; 45 (2): 142-148. doi: 10.1111/j.1600-079X.2008.00568.x
18. Jahnke G, Marr M, Myers C, Wilson R, Travlos G et al. Maternal and developmental toxicity evaluation of melatonin administered orally to pregnant Sprague-Dawley rats. *Toxicological Sciences* 1999; 50 (2): 271-279. doi: 10.1093/toxsci/50.2.271
19. Kilic U, Caglayan AB, Beker MC, Gunal MY, Caglayan B et al. Particular phosphorylation of PI3K/Akt on Thr308 via PDK-1 and PTEN mediates melatonin's neuroprotective activity after focal cerebral ischemia in mice. *Redox Biology* 2017; 12: 657-665. doi: 10.1016/j.redox.2017.04.006

20. Kilic E, Gürsoy Özdemir Y, Bolay H, Keleştimur H, Dalkara T. Pinealectomy aggravates and melatonin administration attenuates brain damage in focal ischemia. *Journal of Cerebral Blood Flow and Metabolism* 1999; 19 (5): 511-516. doi: 10.1097/00004647-199905000-00005
21. Stehle JH, Saade A, Rawashdeh O, Ackermann K, Jilg A et al. A survey of molecular details in the human pineal gland in the light of phylogeny, structure, function and chronobiological diseases. *Journal of Pineal Research* 2011; 51 (1): 17-43. doi: 10.1111/j.1600-079X.2011.00856.x
22. Panda S, Antoch MP, Miller BH, Su AI, Schook AB et al. Coordinated transcription of key pathways in the mouse by the circadian clock. *Cell* 2002; 109 (3): 307-320. doi: 10.1016/s0092-8674(02)00722-5
23. Tian R, Alvarez-Saavedra M, Cheng HY, Figeys D. Uncovering the proteome response of the master circadian clock to light using an AutoProteome system. *Molecular & Cellular Proteomics* 2011; 10 (11): M110.007252. doi: 10.1074/mcp.M110.007252
24. Beker MC, Caglayan B, Yalcin E, Caglayan AB, Turkseven S et al. Time-of-day dependent neuronal injury after ischemic stroke: implication of circadian clock transcriptional factor Bmal1 and survival kinase AKT. *Molecular Neurobiology* 2018; 55 (3): 2565-2576. doi: 10.1007/s12035-017-0524-4
25. Buhr ED, Takahashi JS. Molecular components of the Mammalian circadian clock. In: Kramer A, Meroz M (editors). *Circadian Clocks. Handbook of Experimental Pharmacology*, Vol. 217. Berlin, Germany: Springer; 2013. pp. 3-27. doi: 10.1007/978-3-642-25950-0\_1
26. Beker MC, Caglayan B, Caglayan AB, Keleştemur T, Yalcin E et al. Interaction of melatonin and Bmal1 in the regulation of PI3K/AKT pathway components and cellular survival. *Scientific Reports* 2019; 9 (1): 19082. doi: 10.1038/s41598-019-55663-0
27. Pardiwalla FK, Yeolekar ME, Bakshi SK. Circadian rhythm in acute stroke. *The Journal of the Association of Physicians of India* 1993; 41 (4): 203-204.
28. Vriend J, Reiter RJ. Melatonin feedback on clock genes: a theory involving the proteasome. *Journal of Pineal Research* 2015; 58 (1): 1-11. doi: 10.1111/jpi.12189
29. Yalcin E, Beker MC, Turkseven S, Caglayan B, Gurel B et al. Evidence that melatonin downregulates Nedd4-1 E3 ligase and its role in cellular survival. *Toxicology and Applied Pharmacology* 2019; 379: 114686. doi: 10.1016/j.taap.2019.114686
30. Badr AE, Yin W, Mychaskiw G, Zhang JH. Dual effect of HBO on cerebral infarction in MCAO rats. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 2001; 280 (3): R766-R770. doi: 10.1152/ajpregu.2001.280.3.R766
31. Rusyniak DE, Kirk MA, May JD, Kao LW, Brizendine EJ et al. Hyperbaric oxygen therapy in acute ischemic stroke: results of the hyperbaric oxygen in acute ischemic stroke trial pilot study. *Stroke* 2003; 34 (2): 571-574. doi: 10.1161/01.str.0000050644.48393.d0
32. Kim HY, Singhal AB, Lo EH. Normobaric hyperoxia extends the reperfusion window in focal cerebral ischemia. *Annals of Neurology* 2005; 57 (4): 571-575. doi: 10.1002/ana.20430
33. Shin HK, Dunn AK, Jones PB, Boas DA, Lo EH et al. Normobaric hyperoxia improves cerebral blood flow and oxygenation, and inhibits peri-infarct depolarizations in experimental focal ischemia. *Brain* 2007; 130 (6): 1631-1642. doi: 10.1093/brain/awm071
34. Singhal AB, Wang X, Sumii T, Mori T, Lo EH. Effects of normobaric hyperoxia in a rat model of focal cerebral ischemia-reperfusion. *Journal of Cerebral Blood Flow and Metabolism* 2002; 22 (7): 861-868. doi: 10.1097/00004647-200207000-00011
35. Keleştemur T, Beker MC, Caglayan AB, Caglayan B, Altunay S et al. Normobaric oxygen treatment improves neuronal survival functional recovery and axonal plasticity after newborn hypoxia-ischemia. *Behavioural Brain Research* 2020; 379: 112338. doi: 10.1016/j.bbr.2019.112338
36. Paterniti I, Cordaro M, Esposito E, Cuzzocrea S. The antioxidative property of melatonin against brain ischemia. *Expert Review of Neurotherapeutics* 2016; 16 (7): 841-848. doi: 10.1080/14737175.2016.1182020
37. Garthwaite J, Charles SL, Chess-Williams R. Endothelium-derived relaxing factor release on activation of NMDA receptors suggests role as intercellular messenger in the brain. *Nature* 1988; 336 (6197): 385-388. doi: 10.1038/336385a0
38. Knowles RG, Moncada S. Nitric oxide synthases in mammals. *The Biochemical Journal* 1994; 298 (2): 249-258. doi: 10.1042/bj2980249
39. Beckman JS, Beckman TW, Chen J, Marshall PA, Freeman BA. Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proceedings of the National Academy of Sciences of the United States of America* 1990; 87 (4): 1620-1624. doi: 10.1073/pnas.87.4.1620
40. Samdani AF, Dawson TM, Dawson VL. Nitric oxide synthase in models of focal ischemia. *Stroke* 1997; 28 (6): 1283-1288. doi: 10.1161/01.str.28.6.1283
41. Cheung PY, Wang W, Schulz R. Glutathione protects against myocardial ischemia-reperfusion injury by detoxifying peroxynitrite. *Journal of Molecular and Cellular Cardiology* 2000; 32 (9): 1669-1678. doi: 10.1006/jmcc.2000.1203
42. Fujimoto S, Mizoi K, Yoshimoto T, Suzuki J. The protective effect of vitamin E on cerebral ischemia. *Surgical Neurology* 1984; 22 (5): 449-454. doi: 10.1016/0090-3019(84)90301-x
43. Chang CY, Chen JY, Wu MH, Hu ML. Therapeutic treatment with vitamin C reduces focal cerebral ischemia-induced brain infarction in rats by attenuating disruptions of blood brain barrier and cerebral neuronal apoptosis. *Free Radical Biology & Medicine* 2020; 155: 29-36. doi: 10.1016/j.freeradbiomed.2020.05.015
44. Tan DX, Reiter RJ, Manchester LC, Yan MT, El-Sawi M et al. Chemical and physical properties and potential mechanisms: melatonin as a broad spectrum antioxidant and free radical scavenger. *Current Topics in Medicinal Chemistry* 2002; 2 (2): 181-197. doi: 10.2174/1568026023394443



45. Galano A, Tan DX, Reiter RJ. On the free radical scavenging activities of melatonin's metabolites, AFMK and AMK. *Journal of Pineal Research* 2013; 54 (3): 245-257. doi: 10.1111/jpi.12010
46. El-Abhar HS, Shaalan M, Barakat M, El-Denshary ES. Effect of melatonin and nifedipine on some antioxidant enzymes and different energy fuels in the blood and brain of global ischemic rats. *Journal of Pineal Research* 2002; 33 (2): 87-94. doi: 10.1034/j.1600-079x.2002.02900.x
47. Kilic U, Yilmaz B, Ugur M, Yüksel A, Reiter RJ et al. Evidence that membrane-bound G protein-coupled melatonin receptors MT1 and MT2 are not involved in the neuroprotective effects of melatonin in focal cerebral ischemia. *Journal of Pineal Research* 2012; 52 (2): 228-235. doi: 10.1111/j.1600-079X.2011.00932.x
48. Martin M, Macias M, Escames G, Leon J, Acuna-Castroviejo D. Melatonin but not vitamins C and E maintains glutathione homeostasis in t-butyl hydroperoxide-induced mitochondrial oxidative stress. *The FASEB Journal* 2000; 14 (12): 1677-1679. doi: 10.1096/fj.99-0865fe
49. Acuna Castroviejo D, Lopez LC, Escames G, Lopez A, Garcia JA et al. Melatonin-mitochondria interplay in health and disease. *Current Topics in Medicinal Chemistry* 2011; 11 (2): 221-240. doi: 10.2174/156802611794863517
50. Tan DX, Manchester LC, Liu X, Rosales-Corral SA, Acuna-Castroviejo D et al. Mitochondria and chloroplasts as the original sites of melatonin synthesis: a hypothesis related to melatonin's primary function and evolution in eukaryotes. *Journal of Pineal Research* 2013; 54 (2): 127-138. doi: 10.1111/jpi.12026
51. He C, Wang J, Zhang Z, Yang M, Li Y et al. Mitochondria synthesize melatonin to ameliorate its function and improve mice oocyte's quality under in vitro conditions. *International Journal of Molecular Sciences* 2016; 17 (6). doi: 10.3390/ijms17060939
52. Suofu Y, Li W, Jean-Alphonse FG, Jia J, Khattar NK et al. Dual role of mitochondria in producing melatonin and driving GPCR signaling to block cytochrome c release. *Proceedings of the National Academy of Sciences of the United States of America* 2017; 114 (38): E7997-E8006. doi: 10.1073/pnas.1705768114
53. Xu S, Pi H, Zhang L, Zhang N, Li Y et al. Melatonin prevents abnormal mitochondrial dynamics resulting from the neurotoxicity of cadmium by blocking calcium-dependent translocation of Drp1 to the mitochondria. *Journal of Pineal Research* 2016; 60 (3): 291-302. doi: 10.1111/jpi.12310
54. Orrenius S, Zhivotovskiy B, Nicotera P. Regulation of cell death: the calcium-apoptosis link. *Nature Reviews Molecular Cell Biology* 2003; 4 (7): 552-565. doi: 10.1038/nrm1150
55. Lipton SA. Failures and successes of NMDA receptor antagonists: molecular basis for the use of open-channel blockers like memantine in the treatment of acute and chronic neurologic insults. *Neurotherapeutics* 2004; 1 (1): 101-110. doi: 10.1602/neurorx.1.1.101
56. Kilic U, Yilmaz B, Reiter RJ, Yüksel A, Kilic E. Effects of memantine and melatonin on signal transduction pathways vascular leakage and brain injury after focal cerebral ischemia in mice. *Neuroscience* 2013; 237: 268-276. doi: 10.1016/j.neuroscience.2013.01.059
57. Leon J, Vives F, Crespo E, Camacho E, Espinosa A et al. Modification of nitric oxide synthase activity and neuronal response in rat striatum by melatonin and kynurenine derivatives. *Journal of Neuroendocrinology* 1998; 10 (4): 297-302. doi: 10.1046/j.1365-2826.1998.00203.x
58. Abe K, Aoki M, Kawagoe J, Yoshida T, Hattori A et al. Ischemic delayed neuronal death: a mitochondrial hypothesis. *Stroke* 1995; 26 (8): 1478-1489. doi: 10.1161/01.str.26.8.1478
59. Tajeddine N. How do reactive oxygen species and calcium trigger mitochondrial membrane permeabilisation? *Biochimica et Biophysica Acta* 2016; 1860 (6): 1079-1088. doi: 10.1016/j.bbagen.2016.02.013
60. Andrabi SA, Sayeed I, Siemen D, Wolf G, Horn TF. Direct inhibition of the mitochondrial permeability transition pore: a possible mechanism responsible for anti-apoptotic effects of melatonin. *The FASEB Journal* 2004; 18 (7): 869-871. doi: 10.1096/fj.03-1031fe
61. Ferrarese C, Mascarucci P, Zoia C, Cavarretta R, Frigo M et al. Increased cytokine release from peripheral blood cells after acute stroke. *Journal of Cerebral Blood Flow and Metabolism* 1999; 19 (9): 1004-1009. doi: 10.1097/00004647-199909000-00008
62. Jin R, Yang G, Li G. Inflammatory mechanisms in ischemic stroke: role of inflammatory cells. *Journal of Leukocyte Biology* 2010; 87 (5): 779-789. doi: 10.1189/jlb.1109766
63. Hu X, Li P, Guo Y, Wang H, Leak RK et al. Microglia/macrophage polarization dynamics reveal novel mechanism of injury expansion after focal cerebral ischemia. *Stroke* 2012; 43 (11): 3063-3070. doi: 10.1161/STROKEAHA.112.659656
64. Liu ZJ, Ran YY, Qie SY, Gong WJ, Gao FH et al. Melatonin protects against ischemic stroke by modulating microglia/macrophage polarization toward anti-inflammatory phenotype through STAT3 pathway. *CNS Neuroscience & Therapeutics* 2019; 25 (12): 1353-1362. doi: 10.1111/cns.13261
65. Merlo S, Luaces JP, Spampinato SF, Toro-Urrego N, Caruso GI et al. SIRT1 mediates melatonin's effects on microglial activation in hypoxia: in vitro and in vivo evidence. *Biomolecules* 2020; 10 (3). doi: 10.3390/biom10030364
66. Ahmadi Z, Ashrafizadeh M. Melatonin as a potential modulator of Nrf2. *Fundamental & Clinical Pharmacology* 2020; 34 (1): 11-19. doi: 10.1111/fcp.12498
67. Miller E, Morel A, Saso L, Saluk J. Melatonin redox activity. Its potential clinical applications in neurodegenerative disorders. *Current Topics in Medicinal Chemistry* 2015; 15 (2): 163-169.
68. Leon J, Acuna-Castroviejo D, Escames G, Tan DX, Reiter RJ. Melatonin mitigates mitochondrial malfunction. *Journal of Pineal Research* 2005; 38 (1): 1-9. doi: 10.1111/j.1600-079X.2004.00181.x

69. Pandi-Perumal SR, Trakht I, Srinivasan V, Spence DW, Maestroni GJ et al. Physiological effects of melatonin: role of melatonin receptors and signal transduction pathways. *Progress in Neurobiology* 2008; 85 (3): 335-353. doi: 10.1016/j.pneurobio.2008.04.001
70. Reiter RJ, Tan DX, Sainz RM, Mayo JC, Lopez-Burillo S. Melatonin: reducing the toxicity and increasing the efficacy of drugs. *The Journal of Pharmacy and Pharmacology* 2002; 54 (10): 1299-1321. doi: 10.1211/002235702760345374
71. Stroethoff M, Christoph I, Behmenburg F, Raupach A, Bunte S et al. Melatonin receptor agonist ramelteon reduces ischemia-reperfusion injury through activation of mitochondrial potassium channels. *Journal of Cardiovascular Pharmacology* 2018; 72 (2): 106-111. doi: 10.1097/FJC.0000000000000600
72. Wu XL, Lu SS, Liu MR, Tang WD, Chen JZ et al. Melatonin receptor agonist ramelteon attenuates mouse acute and chronic ischemic brain injury. *Acta Pharmacologica Sinica* 2020; 41: 1016-1024. doi: 10.1038/s41401-020-0361-2.
73. Chumboatong W, Khamchai S, Tocharus C, Govitrapong P, Tocharus J. Agomelatine protects against permanent cerebral ischaemia via the Nrf2-HO-1 pathway. *European Journal of Pharmacology* 2020; 874: 173028. doi: 10.1016/j.ejphar.2020.173028
74. Chumboatong W, Thummayot S, Govitrapong P, Tocharus C, Jittiwat J et al. Neuroprotection of agomelatine against cerebral ischemia/reperfusion injury through an antiapoptotic pathway in rat. *Neurochemistry International* 2017; 102: 114-122. doi: 10.1016/j.neuint.2016.12.011
75. Buendia I, Gomez-Rangel V, Gonzalez-Lafuente L, Parada E, Leon R et al. Neuroprotective mechanism of the novel melatonin derivative Neu-P11 in brain ischemia related models. *Neuropharmacology* 2015; 99: 187-195. doi: 10.1016/j.neuropharm.2015.07.014
76. Lee CH, Yoo KY, Choi JH, Park OK, Hwang IK et al. Melatonin's protective action against ischemic neuronal damage is associated with up-regulation of the MT2 melatonin receptor. *Journal of Neuroscience Research* 2010; 88 (12): 2630-2640. doi: 10.1002/jnr.22430
77. Chern CM, Liao JF, Wang YH, Shen YC. Melatonin ameliorates neural function by promoting endogenous neurogenesis through the MT2 melatonin receptor in ischemic-stroke mice. *Free Radical Biology & Medicine* 2012; 52 (9): 1634-1647. doi: 10.1016/j.freeradbiomed.2012.01.030
78. Boutin JA, Ferry G. Is there sufficient evidence that the melatonin binding site MT3 is quinone reductase 2? *The Journal of Pharmacology and Experimental Therapeutics* 2019; 368 (1): 59-65. doi: 10.1124/jpet.118.253260
79. Dubocovich ML. Pharmacology and function of melatonin receptors. *The FASEB Journal* 1988; 2 (12): 2765-2773. doi: 10.1096/fasebj.2.12.2842214
80. Long II DJ, Iskander K, Gaikwad A, Arin M, Roop DR et al. Disruption of dihydronicotinamide riboside: quinone oxidoreductase 2 (NQO2) leads to myeloid hyperplasia of bone marrow and decreased sensitivity to menadione toxicity. *The Journal of Biological Chemistry* 2002; 277 (48): 46131-46139. doi: 10.1074/jbc.M208675200
81. Buryanovskyy L, Fu Y, Boyd M, Ma Y, Hsieh TC et al. Crystal structure of quinone reductase 2 in complex with resveratrol. *Biochemistry* 2004; 43 (36): 11417-11426. doi: 10.1021/bi049162o
82. Cassagnes LE, Chhour M, Perio P, Sudor J, Gayon R et al. Oxidative stress and neurodegeneration: the possible contribution of quinone reductase 2. *Free Radical Biology & Medicine* 2018; 120: 56-61. doi: 10.1016/j.freeradbiomed.2018.03.002
83. Delagrangre P, Boutin JA. Therapeutic potential of melatonin ligands. *Chronobiology International* 2006; 23 (1-2): 413-418. doi: 10.1080/07420520500464387
84. Becker-Andre M, Wiesenberg I, Schaeren-Wiemers N, Andre E, Missbach M et al. Pineal gland hormone melatonin binds and activates an orphan of the nuclear receptor superfamily. *The Journal of Biological Chemistry* 1994; 269 (46): 28531-28534.
85. Guillaumond F, Dardente H, Giguere V, Cermakian N. Differential control of Bmal1 circadian transcription by REV-ERB and ROR nuclear receptors. *Journal of Biological Rhythms* 2005; 20 (5): 391-403. doi: 10.1177/0748730405277232
86. Nakajima Y, Ikeda M, Kimura T, Honma S, Ohmiya Y et al. Bidirectional role of orphan nuclear receptor RORalpha in clock gene transcriptions demonstrated by a novel reporter assay system. *FEBS Letters* 2004; 565 (1-3): 122-126. doi: 10.1016/j.febslet.2004.03.083
87. Takeda Y, Jothi R, Birault V, Jetten AM. RORgamma directly regulates the circadian expression of clock genes and downstream targets in vivo. *Nucleic Acids Research* 2012; 40 (17): 8519-8535. doi: 10.1093/nar/gks630
88. Van Rossum D, Verheijen BM, Pasterkamp RJ. Circular RNAs: novel regulators of neuronal development. *Frontiers in Molecular Neuroscience* 2016; 9: 74. doi: 10.3389/fnmol.2016.00074
89. Guo JU, Agarwal V, Guo H, Bartel DP. Expanded identification and characterization of mammalian circular RNAs. *Genome Biology* 2014; 15 (7): 409. doi: 10.1186/s13059-014-0409-z
90. Zhang XO, Wang HB, Zhang Y, Lu X, Chen LL et al. Complementary sequence-mediated exon circularization. *Cell* 2014; 159 (1): 134-147. doi: 10.1016/j.cell.2014.09.001
91. Akhter R. Circular RNA and Alzheimer's disease. *Advances in Experimental Medicine and Biology* 2018; 1087: 239-243. doi: 10.1007/978-981-13-1426-1\_19
92. Zuo L, Zhang L, Zu J, Wang Z, Han B et al. Circulating circular RNAs as biomarkers for the diagnosis and prediction of outcomes in acute ischemic stroke. *Stroke* 2020; 51 (1): 319-323. doi: 10.1161/STROKEAHA.119.027348
93. Mehta SL, Pandi G, Vemuganti R. Circular RNA expression profiles alter significantly in mouse brain after transient focal ischemia. *Stroke* 2017; 48 (9): 2541-2548. doi: 10.1161/STROKEAHA.117.017469
94. Nam KI, Yoon G, Kim YK, Song J. Transcriptome analysis of pineal glands in the mouse model of Alzheimer's disease. *Frontiers in Molecular Neuroscience* 2019; 12: 318. doi: 10.3389/fnmol.2019.00318

95. Wang Y, Han D, Zhou T, Zhang J, Liu C et al. Melatonin ameliorates aortic valve calcification via the regulation of circular RNA CircRIC3/miR-204-5p/DPP4 signaling in valvular interstitial cells. *Journal of Pineal Research* 2020; e12666. doi: 10.1111/jpi.12666
96. Manev H, Uz T. The role of the light-dark cycle and melatonin in stroke outcome. *Journal of Stroke and Cerebrovascular Diseases* 1998; 7 (3): 165-167. doi: 10.1016/s1052-3057(98)80002-5
97. MacMahon S, Rodgers A. Blood pressure, antihypertensive treatment and stroke risk. *Journal of Hypertension Supplement* 1994; 12 (10): S5-S14.
98. Wolf PA, Singer DE. Preventing stroke in atrial fibrillation. *American Family Physician* 1997; 56 (9): 2242-2250. doi: