

1-1-2020

Effect of sesamin on protection of equine articular cartilage degradation in vitro

SIRIPORN PEANSUKMANEE

SIRIWAN TANGYUENYONG

SIRIWAN ONGCHAI

Follow this and additional works at: <https://journals.tubitak.gov.tr/veterinary>



Part of the [Animal Sciences Commons](#), and the [Veterinary Medicine Commons](#)

Recommended Citation

PEANSUKMANEE, SIRIPORN; TANGYUENYONG, SIRIWAN; and ONGCHAI, SIRIWAN (2020) "Effect of sesamin on protection of equine articular cartilage degradation in vitro," *Turkish Journal of Veterinary & Animal Sciences*: Vol. 44: No. 1, Article 20. <https://doi.org/10.3906/vet-1811-66>
Available at: <https://journals.tubitak.gov.tr/veterinary/vol44/iss1/20>

This Article is brought to you for free and open access by TÜBİTAK Academic Journals. It has been accepted for inclusion in Turkish Journal of Veterinary & Animal Sciences by an authorized editor of TÜBİTAK Academic Journals. For more information, please contact academic.publications@tubitak.gov.tr.

Effect of sesamin on protection of equine articular cartilage degradation in vitro

Siriporn PEANSUKMANEE^{1*}, Siriwan TANGYUENYONG², Siriwan ONGCHAI²

¹Department of Companion Animals and Wildlife Clinic, Faculty of Veterinary Medicine, Chiang Mai University, Chiang Mai, Thailand

²Thailand Excellence Center for Tissue Engineering and Stem Cells, Department of Biochemistry, Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand

Received: 25.11.2018 • Accepted/Published Online: 07.12.2019 • Final Version: 10.02.2020

Abstract: Black sesame seed is a potential herbal medicine for osteoarthritis treatment as it contains sesamin, which has antiinflammatory properties and has demonstrated chondroprotective effects in many species. Horses are particularly susceptible to osteoarthritis. Inflammatory mediators and matrix metalloproteinase are important factors causing cartilage degradation. This study aimed to investigate the protective potential of sesamin (0.5–2.0 μ M) against interleukin (IL)-1 β -induced equine cartilage explant degradation. The cartilage degradation was monitored by measuring the release of cartilage matrix molecules in culture media including sulfated-glycosaminoglycans and hyaluronan by calorimetric assay and ELISA assay, respectively. The remaining contents of collagen and uronic acid (UA) within the explant tissue were assessed by hydroxyproline assay and UA assay, respectively. Cartilage-degrading enzymes MMP-2 and MMP-3 were evaluated by gelatin zymography and ELISA assay, respectively. The results indicated that sesamin suppressed IL-1 β -induced equine cartilage degradation by reducing the levels of sulfated-glycosaminoglycans and hyaluronan, and preserving the contents of UA and collagen within the explant tissues. The activity of MMP-2 and the quantity of MMP-3 were also suppressed by sesamin. The chondroprotective efficacy of sesamin was comparable to that of diacerein, an antiarthritic drug. Lactate dehydrogenase assay showed that sesamin at concentrations of up to 2 mM did not cause cytotoxicity to chondrocytes in cartilage explants. These results may lead to new options for osteoarthritis treatment in horses.

Key words: Sesamin, equine articular cartilage, cartilage degradation, chondroprotection

Joint diseases cause chronic health issues in athletic animals, including horses. In Europe, more than 44.8% of sport horses were retired and/or consequently euthanized due to osteoarthritis [1]. Injury to joints and surrounding tissues induces inflammatory processes. Proinflammatory mediators, especially interleukin (IL)-1 β and proteolytic enzymes, are released into the synovial fluid, causing articular cartilage destruction [2]. Matrix metalloproteinases, including MMP-1 (collagenase 1), MMP-2 (gelatinase), MMP-3 (stromelysin 1), MMP-9 (gelatinase), and MMP-13 (collagenase 3), play major catalytic roles in the cartilage [3–9]. Osteoarthritis treatments usually combine medicine, surgery, and physical therapy. Alternative medicines such as acupuncture and herbal medicine are introduced in some cases in order to minimize the side effects of the main treatments and to increase the quality of life of the patient.

Black sesame is known as an herb that can be used as a health promotion supplement and as a medicine for clinical conditions such as dermatitis, migraine, and joint pain. Black sesame contains lignans such as sesamin

and sesamol [10]. Previous studies demonstrated the antiinflammatory effects of sesamin both in vivo [11–14] and in vitro [15]. It has been reported that sesamin extract reduced pathogenesis of arthritis by increasing articular cartilage thickness in inflammation-induced mice. Furthermore, the extract not only significantly reduced cartilage degradation but also enhanced both collagen and sulfated-glycosaminoglycan synthesis within the cartilage tissue [16].

It was therefore of interest to determine whether sesamin would have similar effects in horse cartilage. This study aimed to investigate the chondroprotective effect of sesamin on equine articular cartilage degradation induced by IL-1 β in a cartilage explant model.

Equine articular cartilage samples were obtained within 6 h post mortem from an equine patient after consent was obtained from the owner. The animal had no clinical signs of joint disease. The articular cartilage was collected from the stifle joints of both hindlimbs. Full thickness cartilage biopsies were harvested aseptically and trimmed into

* Correspondence: siriporn.pean@cmu.ac.th

cartilage explants of approximately 3×3 mm². The explants were then transported to the laboratory in Dulbecco's modified Eagle medium (DMEM; GIBCO) with 200 U/mL penicillin G sodium and 200 unit/mL streptomycin. The explants were rinsed at least three times with the same medium before being weighed and cultured in 12-well tissue culture plates containing 0.5 mL of serum-free DMEM with 200 U/mL penicillin G sodium and 200 U/mL streptomycin in each well. The cultures were maintained at 37 °C under 5% CO₂. The tissue culture media in the first 24 h were collected as day 0 samples. The media were replaced weekly, and the explants were cultured for 7 or 21 days with various treatments. Untreated control explants were cultured in media without any additional substances. The IL-1 β group was treated with 10 ng/mL recombinant human IL-1 β (Sigma-Aldrich). The sesamin groups

received a co-treatment of 10 ng/mL recombinant human IL-1 β in addition to 0.5, 1.0, or 2.0 μ M sesamin (Sigma-Aldrich). Diacerein (TRB Chemidica), an antiarthritic drug, at a concentration of 20 μ M was treated in parallel as the positive control. Collected tissues and culture media were kept at -20 °C until use.

Cartilage biomolecules that were released into the tissue culture media were evaluated. Sulfated-glycosaminoglycans were measured by a colorimetric dye binding assay [17]. Hyaluronan was measured by competitive inhibition-based ELISA [18]. IL-1 β successfully induced significant release of sulfated-glycosaminoglycans and hyaluronan compared to the control. Sesamin at a concentration of 2 μ M significantly suppressed the release of sulfated-glycosaminoglycans and hyaluronan induced by IL-1 β , comparable to the diacerein group (Figure 1). In addition

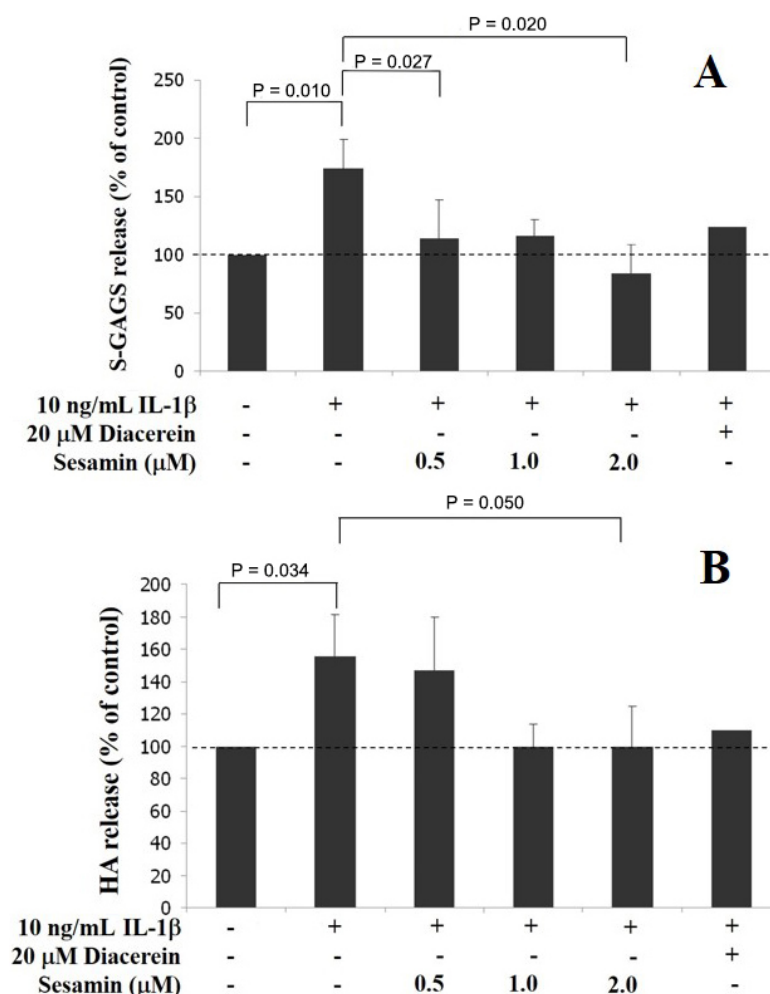


Figure 1. Sulfated-glycosaminoglycans (S-GAGS) (A) and hyaluronan (HA) (B) release percentages relative to the control groups. Comparison among the IL-1 β group (10 ng/mL IL-1 β), the diacerein group (20 μ M diacerein), and the sesamin groups (0.5, 1.0, and 2.0 μ M sesamin) on day 7 of the cartilage explant cultures. Data were analyzed using STATA version 9.2 with 95% confidence intervals ($P < 0.05$). The percentage concentration of sulfated-glycosaminoglycans and hyaluronan between the control and IL-1 β group were compared by unpaired t-test, while the IL-1 β group and the other treatment groups were compared by ANOVA.

to the detection of catabolic markers that were released into the tissue culture media, the remaining collagen and uronic acid (UA) contents within the explants were also analyzed on day 21 using a hydroxyproline assay [19] and a UA assay [18], respectively. The results showed an obvious but not statistically significant reduction of UA and collagen contents after IL-1 β induction. Although there was no statistically significance, sesamin and diacerein showed the increasing trend of UA and collagen contents in the explants compared to the cytokine-treated group (Figures 2). These findings suggest that sesamin protects against IL-1 β -induced release of cartilage matrix molecules similarly to diacerein, the standard antiarthritic drug.

Matrix metalloproteinase-2 (MMP-2) activity was measured by gelatin zymography [20], and the MMP-3 level was analyzed using a human MMP-3 test kit (Elabscience). The results from the gelatinolytic assay showed that tissue culture media of IL-1 β -induced explants had higher MMP-2 activity. This activity increase was reduced by co-treatment with diacerein or 2 μ M

sesamin (Figure 3A). The MMP-3 level was increased after treatment with the proinflammatory cytokine IL-1 β , but tended to be decreased with additional sesamin and diacerein (Figure 3B). However, these changes were not statistically significant. These results suggest that sesamin may attenuate the effects of IL-1 β on activation of the production of MMPs, leading to the slowing of cartilage matrix degradation in the equine model comparably to diacerein, an IL-1 β inhibitor [21–23], in our equine cartilage study.

Sesamin toxicity on cartilage explant cultures was tested by detecting lactate dehydrogenase (LDH), which indicates the death of cells [24], including chondrocytes. H₂O₂ at a concentration of 10 mM was used as the positive control. After culturing for 7 days, sesamin at concentrations of 0.5 to 2 μ M did not alter LDH levels compared to the control group, while the explants that were treated with hydrogen peroxide showed highly significant increases in LDH. However, sesamin at 4 μ M significantly increased LDH (Figure 4). Therefore, only 0.5 to 2 μ M sesamin was applied in the experiments.

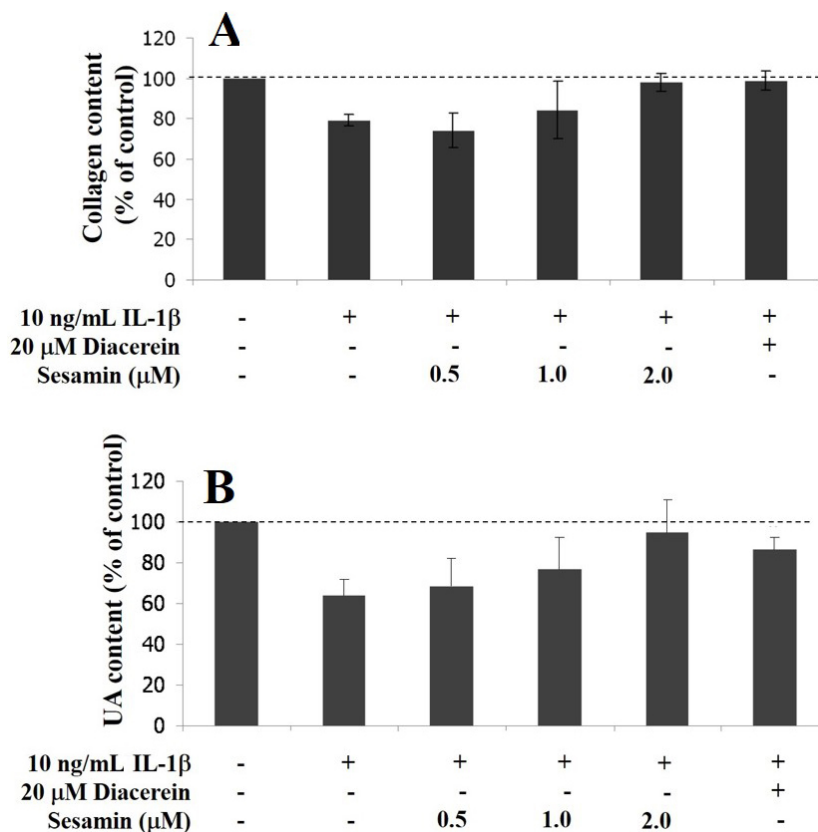


Figure 2. Percentage relative to the control group of UA (A) and collagen (B) content within the cartilage explants on day 21 of the cartilage explant cultures. Comparison among the IL-1 β group (10 ng/mL IL-1 β), the diacerein group (20 μ M diacerein), and the sesamin groups (0.5, 1.0, and 2.0 μ M sesamin). Data were analyzed using STATA version 9.2 with 95% confidence intervals ($P < 0.05$). The percentage concentrations of UA and collagen between the control and IL-1 β group were compared by unpaired t-test, while the IL-1 β group and the other treatment groups were compared by ANOVA.

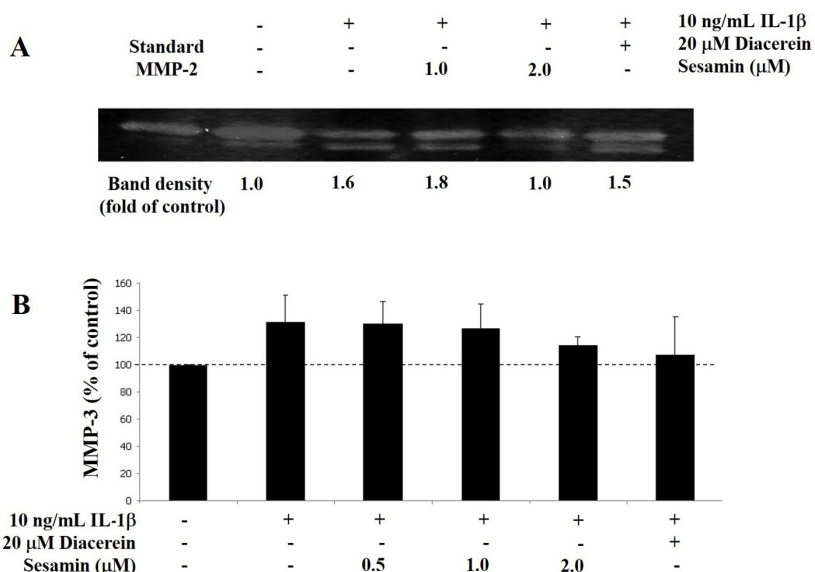


Figure 3. (A) Gelatin zymography of the activity of MMP-2 in the tissue culture media compared with the control group, the IL-1 β group (10 ng/mL IL-1 β), the diacerein group (20 μ M diacerein), and the sesamin groups (0.5, 1.0, and 2.0 μ M sesamin). Band densities were determined using TotalLab TL120 v2009 software and are represented as density fold compared to the control. (B) Percentage relative to the control of MMP-3 released into the tissue culture media. Comparison among the IL-1 β group (10 ng/mL IL-1 β), the diacerein group (20 μ M diacerein), and the sesamin groups (0.5, 1.0, and 2.0 μ M sesamin). The MMP-2 zymography result was not statistically tested. The percentage concentrations of MMP-3 between the control and IL-1 β group were compared by unpaired t-test, while the IL-1 β group and the other treatment groups were compared by ANOVA.

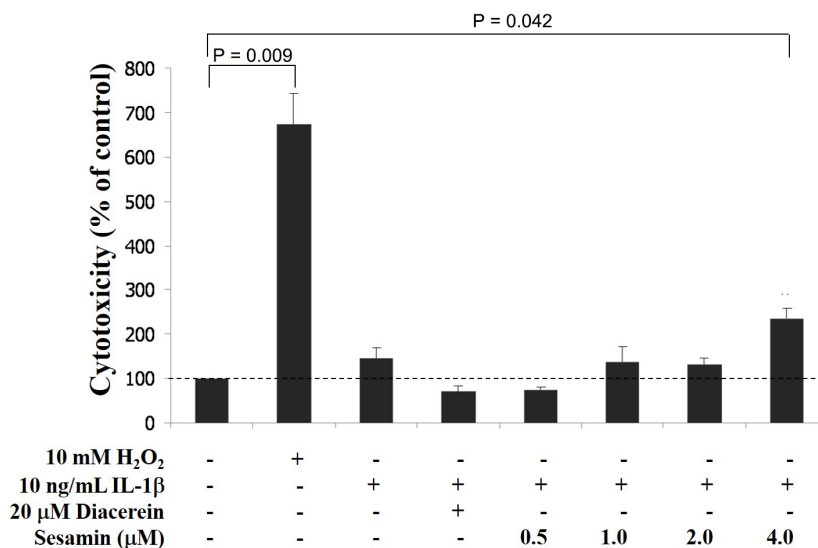


Figure 4. Lactate dehydrogenase (LDH) percentage relative to the control group. Comparison among treated group (10 mM H₂O₂), IL-1 β group (10 ng/mL IL-1 β), the diacerein group (20 μ M diacerein), and the sesamin groups (0.5, 1.0, 2.0, and 4.0 μ M sesamin). Data were analyzed using STATA version 9.2 with 95% confidence intervals ($P < 0.05$). The percentages of LDH between the control and IL-1 β group were compared by unpaired t-test, while the IL-1 β group and the other treatment groups were compared by ANOVA.

Taken all together, our results demonstrated the chondroprotective potential of sesamin in equine cartilage explant degradation induced by IL-1 β , one of the key proinflammatory cytokines that involves pathogenesis of osteoarthritis. Accordingly, sesamin may be applied as an alternative treatment for osteoarthritis in human and animals, including horses.

Acknowledgments and/or disclaimers, if any

This work was supported by a Chiang Mai University research grant (2013). The authors would like to thank the Thailand Excellence Center for Tissue Engineering and Stem Cells, Chiang Mai University, for general support. We also thank the Large Animal Teaching Hospital of Chiang Mai University for the cartilage supply.

References

- Wallin L, Strandberg E, Philipson J, Dalin G. Estimates of longevity and causes of culling and death in Swedish warmblood and coldblood horses. *Livestock Production Science* 2000; 63 (3): 275-289. doi: 10.1016/S0301-6226(99)00126-8
- McIlwraith CW. Traumatic joint injuries and disease: intraarticular fractures amenable to treatment and in which the horse can be returned to athletic activity. In: Stashak TS (editor). *Lameness in The Horse; An In-Depth Short Course for the Horseman*. 1st ed. Boulder, CO, USA: Equine Sciences of Colorado State University; 1997. pp. 39-42.
- Kubota E, Imamura H, Kubota T, Shibata T, Murakami KI. Interleukin 1 β and stromelysin (MMP3) activity of synovial fluid as possible markers of osteoarthritis in the temporomandibular joint. *Journal of Oral and Maxillofacial Surgery* 1997; 55 (1): 20-27. doi: 10.1016/S0278-2391(97)90438-9
- McIlwraith CW. General pathobiology of the joint and response to injury. In: McIlwraith CW, Trotter GW (editors). *Joint Diseases in the Horse*. 1st ed. Philadelphia, PA, USA: W.B. Saunders; 1996. pp. 51-54.
- Clegg PD, Coughlan AR, Riggs CM, Carter SD. Matrix metalloproteinase 2 and 9 in equine synovial fluids. *Equine Veterinary Journal* 1997; 29 (5): 343-348. doi: 10.1111/j.2042-3306.1997.tb03137.x
- Clegg PD, Mobasher A. Chondrocyte apoptosis, inflammatory mediators and equine osteoarthritis. *The Veterinary Journal* 2003; 166 (1): 3-4. doi: 10.1016/S1090-0233(02)00270-8
- Birkedal-Hansen H, Moore WG, Bodden MK, Windsor LJ, Birkedal-Hansen B et al. Matrix metalloproteinases; a review. *Critical Reviews in Oral Biology & Medicine* 1993; 4 (2): 197-250. doi: 10.1177/10454411930040020401
- Yoshihara Y, Nakamura H, Obata K, Yamada H, Hayakawa T et al. Matrix metalloproteinases and tissue inhibitors of metalloproteinases in synovial fluids from patients with rheumatoid arthritis or osteoarthritis. *Annals of the Rheumatic Diseases* 2000; 59 (6): 455-461. doi: 10.1136/ard.59.6.455
- Mehraban F, Kuo SY, Riera H, Chang C, Moskowitz RW. Prostromelysin and procollagenase genes are differentially UP-regulated in chondrocytes from the knees of rabbits with experimental osteoarthritis. *Arthritis & Rheumatology* 1994; 37 (8): 1189-1197. doi: 10.1002/art.1780370813
- Jeng KCG, Hou RCW. Sesamin and sesamol: nature's therapeutic lignans. *Current Enzyme Inhibition* 2005; 1 (1): 11-20. doi: 10.2174/1573408052952748
- Wu WH, Wang SH, Kuan II, Kao YS, Wu PJ et al. Sesamin attenuates intercellular cell adhesion molecule-1 expression in vitro in TNF-alpha-treated human aortic endothelial cells and in vivo in apolipoprotein-E-deficient mice. *Molecular Nutrition & Food Research* 2010; 54 (9): 1340-1350. doi: 10.1002/mnfr.200900271
- Utsunomiya T, Chavali SR, Zhong WW, Forse RA. Effects of sesamin-supplemented dietary fat emulsions on the ex vivo production of lipopolysaccharide-induced prostanoids and tumor necrosis factor alpha in rats. *American Journal of Clinical Nutrition* 2000; 72 (3): 804-808. doi: 10.1093/ajcn/72.3.804
- Utsunomiya T, Shimada M, Rikimaru T, Hasegawa H, Yamashita Y et al. Antioxidant and anti-inflammatory effects of a diet supplemented with sesamin on hepatic ischemia-reperfusion injury in rats. *Hepato-Gastroenterology* 2003; 50 (53): 1609-1613.
- Cui Y, Hou X, Chen J, Xie L, Yang L et al. Sesamin inhibits bacterial formylpeptide-induced inflammatory responses in a murine air-pouch model and in THP-1 human monocytes. *Journal of Nutrition* 2010; 140 (2): 377-381. doi: 10.3945/jn.109.117804
- Jeng KC, Hou RC, Wang JC, Ping LI. Sesamin inhibits lipopolysaccharide-induced cytokine production by suppression of p38 mitogen-activated protein kinase and nuclear factor-kappa B. *Immunology Letters* 2005; 97 (1): 101-106. doi: 10.1016/j.imlet.2004.10.004
- Phitak T. Molecular investigation of phytochemicals having effects on human chondrocyte metabolism. PhD, Chiang Mai University, Chiang Mai, Thailand, 2010.
- Farndale RW, Buttle DJ, Barrett AJ. Improved quantitation and discrimination of sulphated glycosaminoglycans by use of dimethylmethylene blue. *Biochimica et Biophysica Acta* 1986; 883 (2): 173-177. doi: 10.1016/0304-4165(86)90306-5
- Pothacharoen P, Choocheep K, Pitak T, Pompimon W, Premanode B et al. Effect of *Alpinia galanga* extract on cartilage degradation and on gene expression in human chondrocyte and synovial fibroblast metabolism. *Central European Journal of Biology* 2006; 1 (3): 430-450. doi: 10.2478/s11535-006-0030-6
- Hoemann CD, Sun J, Chrzanowski V, Buschmann MD. A multivalent assay to detect glycosaminoglycan, protein, collagen, RNA, and DNA content in milligram samples of cartilage or hydrogel-based repair cartilage. *Analytical Biochemistry* 2002; 300 (1): 1-10. doi: 10.1006/abio.2001.5436

20. Ito A, Nose T, Takahashi S, Mori Y. Cyclooxygenase inhibitors augment the production of pro-matrix metalloproteinase 9 (progelatinase B) in rabbit articular chondrocytes. *FEBS Letters* 1995; 360 (1): 75-79. doi: 10.1016/0014-5793(95)00085-N
21. Moldovan F, Pelletier JP, Jolicoeur FC, Cloutier JM, Martel-Pelletier J. Diacerhein and rheim reduce the ICE-induced IL-1 beta and IL-18 activation in human osteoarthritic cartilage. *Osteoarthritic and Cartilage* 2000; 8 (3): 186-196. doi: 10.1053/joca.1999.0289
22. Martel-Pelletier J, Mineau F, Jolicoeur FC, Cloutier JM, Pelletier JP. In vitro effects of diacerhein and rheim on interleukin 1 and tumor necrosis factor-alpha systems in human osteoarthritic synovium and chondrocytes. *Journal of Rheumatology* 1998; 25 (4): 753-762.
23. Yaron M, Shirazi I, Yaron I. Anti-interleukin-1 effects of diacerhein and rheim in human osteoarthritic synovial tissue and cartilage cultures. *Osteoarthritis and Cartilage* 1999; 7 (3): 272-280. doi: 10.1053/joca.1998.0201
24. Korzeniewski C, Callenwaert DM. An enzyme-release assay for natural cytotoxicity. *Journal of Immunological Methods* 1983; 64 (3): 313-320. doi: 10.1016/0022-1759(83)90438-6