

Can the vascular endothelial growth factor (VEGF) and total estradiol 17 β be used as a marker of equine oocyte maturation?

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Abstract: The aim of this study was to quantitate vascular endothelial growth factor (VEGF) and estradiol-17 β in follicular fluid (FF) and serum and to correlate the levels of these substances with oocyte maturation. Transvaginal ultrasound-guided follicular aspirations were performed with the purpose of oocyte collection when the follicle was >35 mm in diameter. The mares were divided into 2 groups according to age, and the changes in VEGF and estradiol-17 β in follicular fluid and serum were examined. Levels of estradiol-17 β in the follicular fluid and serum of young mares (4-10 years old) did not differ from those in old mares (20-24 years old) ($P > 0.05$). The follicular fluid and serum concentrations of VEGF in old mares were lower ($P < 0.001$) than those in young mares. The VEGF levels in young mares did not differ in cases when either mature or intermediate oocytes were collected, whereas its levels in old mares were higher ($P < 0.01$) when mature oocytes were collected than when intermediate and immature oocytes were collected. Our data suggest that VEGF may play an important role in follicular growth and development, that VEGF levels appear to be age-dependent, and that VEGF levels might be valuable biochemical markers of oocyte maturation.

Key words: VEGF, oocyte maturation, follicular fluid, mare

Vasküler endotelial büyüme faktörü (VEBF) ve total östradiol 17 β kısraak oosit maturasyonunda bir belirleyici olarak kullanılabilir mi?

Özet: Bu çalışmanın amacı folliküler sıvı (FS) ve serumda vasküler endotelial büyüme faktörü (VEBF) ve total östradiol 17 β 'yi belirlemek ve bu maddelerin seviyelerini oosit maturasyonu ile ilişkilendirmektir. Follikül çapı 35 mm'den büyük olduğunda; oosit toplama amacıyla transvajinal ultrason rehberliğinde follikül aspirasyonu gerçekleştirildi. Kısraaklar yaşa göre iki gruba ayrıldı ve VEBF ve östradiol 17 β değişimleri folliküler sıvı ve serumda incelendi. Folliküler sıvı ve serum östradiol 17 β seviyeleri genç (4-10 yaş) ve yaşlı (20-24 yaş) kısraakların arasında farklı bulunmadı ($P > 0,05$). Yaşlı kısraakların folliküler sıvı ve serum VEBF konsantrasyonları genç kısraaklardakinden daha düşük bulundu ($P < 0,001$). Genç kısraaklardan elde edilen VEBF seviyesi mature ve intermediate oositler arasında fark göstermezken; yaşlı kısraaklardan elde edilen VEBF seviyesi mature oositlerde intermediate ve immature oositlere göre daha yüksek bulundu ($P < 0,01$). Yapılan çalışma sonucunda; VEBF'nün folliküler büyüme ve gelişimde önemli bir rol oynayabileceği, VEBF seviyelerinin yaşa bağımlı gözüktüğü ve bu nedenle VEBF seviyeleri oosit maturasyonun değerli biyokimyasal belirleyicisi olabileceği kanısına varıldı.

Anahtar sözcükler: VEBF, oosit maturasyonu, folliküler sıvı, kısraak

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Introduction

Although the process of ovarian follicular development and nuclear maturation of oocytes is governed by conventional endocrine principles such as pituitary gonadotropins and ovarian steroids, recent observations suggest that gonadotropins are only part of a complex system of autocrine and paracrine agents, including growth factors (1).

Growth factors play a regulatory role in ovarian function. Vascular endothelial growth factor (VEGF), a protein thought to play a critical role in the regulation of vascular permeability and angiogenesis during embryogenesis and in pathological tissue growth, is also expressed in the ovaries of numerous species (2-4). VEGF production increases in luteinizing granulosa cells of the ovulatory follicle as well as in the developing corpus luteum in primates, suggesting that this factor is important for vascular changes associated with the transition of the follicle to the corpus luteum (5).

In the ovary, VEGF is produced both by granulosa and thecal cells (6,7). A recent study of macaque granulosa cells suggested that VEGF expression increases in response to gonadotrophins such as LH, FSH, and human chorionic gonadotrophin (HCG) (5).

In women with normal non-stimulated cycles and those undergoing IVF, the local VEGF production in follicular fluid is correlated with the degree of follicular luteinization (8,9). Progesterone also appears to play a role in determining VEGF concentration in follicular fluid (10). A positive correlation has been observed between follicular fluid VEGF concentrations and patient age, especially in patients ≥ 38 years undergoing IVF (11,12). Recently, it has been reported that elevated VEGF concentrations in follicular fluid might predict poor conception rates after IVF (13).

The aim of this study was to quantitate the VEGF and total estradiol 17 β in follicular fluid (FF) and serum, and to correlate the levels of these substances with oocyte maturation and mare age.

Materials and methods

The study was conducted during the breeding season on younger (4-10 years old) and older (20-24 years old) non-lactating Arabian mares. Transvaginal ultrasound-guided follicular aspirations were performed with the purpose of oocyte collection when the follicle was >35 mm in diameter.

Oocytes were immediately identified, rinsed twice in culture medium, and placed in a petri dish containing 2.5 mL of culture medium (Tissue Culture Medium 199 with additions of 10% fetal calf serum, 0.2 mM pyruvate, and 50 $\mu\text{g}/\text{mL}$ gentamicin). Oocytes were incubated in air containing 6% CO_2 at 38.5 $^\circ\text{C}$ for approximately 16 h.

Oocytes were graded based on morphology (scores from 1, excellent, to 4, poor). Cumulus expansion was graded as immature, intermediate, or mature. Grades and maturity were assigned after culture. Blood and follicular fluid were not used from cycles in which oocytes were not recovered. Critical evaluations of oocytes were difficult, as cumulus cells frequently obstructed direct imaging of the oocyte under a stereomicroscope.

Jugular blood (10 mL) was collected at the time of follicular aspiration. Blood samples and follicular fluid were centrifuged at $3500 \times g$ for 15 min and stored at -20 $^\circ\text{C}$ until being analyzed for VEGF and estradiol-17 β . Concentrations of estradiol in follicular fluid and serum were estimated by radioimmunoassay (RIA) using an active estradiol RIA kit (active Estradiol RIA DSL-43100, Webster, TX, USA). The amount of [125I]-labeled estradiol bound to antibody is inversely proportional to the concentration of unlabeled estradiol present. The separation of free and bound antigen is achieved by decanting or aspirating the antibody-coated tubes. The sensitivity, defined as the lowest detectable level of estradiol that can be distinguished from the 0 pg/mL Estradiol Standard, is 11 pg/mL at the 95% confidence limit. The intra-assay precision was determined from the mean of 12 replicates each and the coefficient of variation was 2.01% when the mean was 241.73 pg/mL. The inter-assay precision was determined from the mean of average duplicates for 24 separate runs and the coefficient of variation was 7.37% when the mean was 247.48 pg/mL. The assay has low cross-reactivity with equilenin (6.1%),

estrone (3.4%), 17 β -estradiol-3-glucuronide (1.8%), estrone- β -d-glucuronide (0.3%), 16-ketoestradiol (0.29%), 17 α -estradiol (0.26%), estradiol-3-sulfate (0.21%), estriol (0.75%), and equilin (0.84%). The results were measured by gamma counter (MGM Instruments, ISOCOMP I, USA).

Concentrations of VEGF in follicular fluid and serum were assayed with the ELISA test kit for Equine Vascular Endothelial Cell Growth Factor (USCN Life Science Inc. Wuhan, China). The minimum detectable dose of equine VEGF is typically less than 0.04 ng/mL. According to the manufacturer this assay has high sensitivity and excellent specificity for detection of equine VEGF. No significant cross-reactivity or interference was observed.

The least squares procedures were used to analyze for VEGF and total estradiol-17 β . The model used in the analysis included fixed effects of age (young or old), sample (follicular fluid or serum) and maturation (mature, intermediate, or immature) and 2-way interactions of these effects.

Results

The mares were divided into 2 groups based on age, and the changes in VEGF in follicular fluid and serum were examined (Table). The follicular fluid and serum concentrations of VEGF in old mares were lower ($P < 0.001$) than those in young mares. Levels of estradiol-17 β in the follicular fluid and serum of young mares did not differ from those in

old mares ($P > 0.05$). Estradiol-17 β level in follicular fluid was higher ($P < 0.001$) than that in the serum of the young and old mares. VEGF level in follicular fluid was lower ($P < 0.05$) than that in the serum of the young and old mares.

Oocyte grades varied ($P < 0.05$) with age, with higher grades representing worse morphology scores (young 4-10 years, 1.9 ± 0.1 , $n = 14$ and old 20-24 years, 2.6 ± 0.2 , $n = 20$).

VEGF level for young mares did not differ between when mature and intermediate oocytes were collected whereas its level for old mares when mature oocytes were collected was higher ($P < 0.01$) than when intermediate and immature oocytes were collected (Figure).

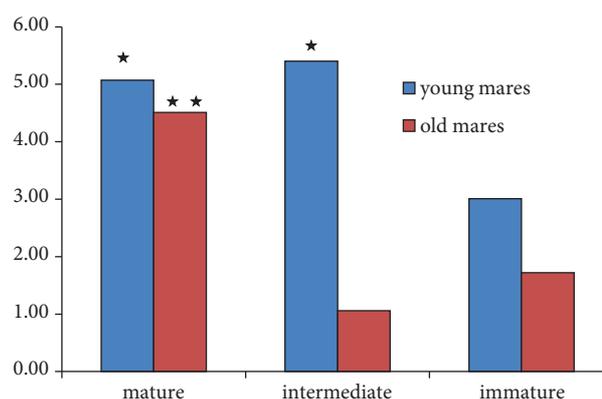


Figure. The mean concentrations of VEGF for young and old mares. Oocytes were classified as immature, intermediate, or mature.

Table. The comparison of VEGF and estradiol-17 β concentrations in follicular fluid (FF) and serum in young and old mares.

	VEGF (ng/mL)		Estradiol-17 β	
	FF	Serum	FF (pg/mL)	Serum (ng/mL)
Young (n = 14)	3.46 \pm 0.47	5.06 \pm 0.47	250.7 \pm 70.4	68.5 \pm 7.2
Old (n = 20)	1.91 \pm 0.5	2.9 \pm 0.5	303.5 \pm 67.3	53.3 \pm 4.7
Significance				
Age		$P < 0.001$		$P > 0.05$
Sample		$P < 0.05$		$P < 0.001$
Age & Sample		$P > 0.05$		$P > 0.05$

Discussion

Estradiol-17 β and progesterone have been suggested as markers for oocyte maturity and fertilization in humans (14,15). On the other hand, it has been reported that there is no correlation between oocyte maturity and the concentrations of estradiol-17 β and progesterone in FF (16-19). Consequently, no relationship between ovarian steroid hormones and oocyte maturation was observed.

Therefore, we hypothesized that local factors, such as growth factors produced in follicles, might act upon follicular development and oocyte maturation rather than ovarian steroid hormones in mares. In our study the levels of estradiol-17 β in the follicular fluid and serum of young mares did not differ from those in old mares. We previously (20) reported that IGF-I and progesterone concentrations were significantly reduced and correlated with oocyte maturation. IGF-I levels in follicular fluid and serum were affected by mare age, and the IGF-I concentrations in follicular fluid from young mares were higher than those from old mares. It was suggested that follicular fluid levels of IGF-I and progesterone could be available biochemical markers of oocyte maturation.

In the present study, we demonstrated that VEGF is present in the follicular fluid. The follicular fluid and serum concentrations of VEGF in old mares

were lower than those in young mares. Moreover, VEGF levels in young mares were not different between when mature and intermediate oocytes were collected, whereas its levels in old mares when mature oocytes were collected were higher than when intermediate and immature oocytes were collected. Kawano et al. (21) reported that VEGF was increased in older women. Friedman et al. (13) reported that VEGF in the FF of women of advanced reproductive age was elevated compared with that of younger women. They suggested that the increasing VEGF level was caused by a hypoxic environment within the follicles of older women. It has been reported that VEGF did not reveal significant differences in the mature group versus immature oocytes, but VEGF with fertilized oocytes was significantly higher than non-fertilized oocytes in mature oocytes (22). Our data suggest that VEGF may play an important role in follicular growth and development, that VEGF levels appear to be age-dependent, and that VEGF levels might be valuable biochemical markers of oocyte maturation.

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References

1. Tonetta, S.A., Dizerega, G.S.: Intragonadal regulation of follicular maturation. *Endocr. Rev.*, 1989; 10: 205-229.
2. Redmer, D.A., Reynolds, L.P.: Angiogenesis in the ovary. *Rev. Reprod.*, 1996; 1: 182-192.
3. Ferrara, N., Davis-Smyth, T.: The biology of vascular endothelial growth factor. *Endocr. Rev.*, 1997; 18: 4-25.
4. Hazzard, T.M., Christenson, L.K., Stouffer, R.L.: Changes in expression of vascular endothelial growth factor (VEGF), angiopoietin (Ang)-1 and Ang-2 in the macaque corpus luteum during the menstrual cycle. *Mol. Hum. Reprod.*, 2000; 6: 993-998.
5. Christenson, L.K., Stouffer, R.L.: Follicle-stimulating hormone and luteinizing hormone/chorionic gonadotropin stimulation of vascular endothelial growth factor production by macaque granulosa cells from pre- and periovulatory follicles. *J. Clin. Endocrinol. Metab.*, 1997; 82: 2135-2142.
6. Ravindranath, L., Little-Ihring, L.L., Phillips, H.S., Ferrara, N., Zeleznik A.S.: Vascular endothelial growth factor messenger ribonucleic acid expression in primate ovary. *Endocrinology*, 1992; 131: 254-260.
7. Kamat, B.R., Brown, L.F., Manseau, E.J., Senger, D.R., Dvorak, H.F.: Expression of VEGF by human granulosa and theca luteal cells: role in corpus luteum development. *Am. J. Pathol.*, 1995; 146: 157-165.
8. Lee, A., Christenson, L.K., Stouffer, R.L., Burry, K.A., Patton, P.E.: Vascular endothelial growth factor levels in serum and follicular fluid of patients undergoing in vitro fertilization. *Fertil. Steril.*, 1997; 68: 305-311.
9. Anasti, J.N., Kalantaridou, S.N., Kimzey, L.M., George, M., Nelson N.M.: Human follicle fluid vascular endothelial growth factor concentrations are correlated with luteinization in spontaneously developing follicles. *Hum. Reprod.*, 1998; 13: 1144-1147.

10. Moncayo, H.E., Penz-Koza, A., Marth, C., Gasti, G., Herold, M., Moncayo, R.: Vascular endothelial growth factor in serum and in the follicular fluid of patients undergoing hormonal stimulation for in-vitro fertilization. *Hum. Reprod.*, 1998; 13: 3310-3314.
11. Friedman, C.I., Arbogast, L., Danforth, D.R.: Follicular fluid vascular endothelial growth factor concentrations are elevated in women of advanced reproductive age undergoing ovulation induction. *Fertil. Steril.*, 1997; 68: 607-612.
12. Manau, D., Balasch, J., Jimenez, W., Fabregues, F., Civico, S., Casamitjana, R., Creus, M., Vanrell, JA.: Follicular fluid concentrations of adrenomedullin, vascular endothelial growth factor and nitric oxide in IVF cycles: relationship to ovarian response. *Hum. Reprod.*, 2000; 15: 1295-1299.
13. Friedman, C.I., Seifer, D.B., Kennard, E.A., Arbogast, L., Alak, B., Danforth, D.R.: Elevated level of follicular fluid vascular endothelial growth factor is a marker of diminished pregnancy potential. *Fertil. Steril.*, 1998; 70: 836-839.
14. Fishel, S.B., Edwards, R.G., Walters, D.E.: Follicular steroids as prognosticator of successful fertilization of human oocytes in vitro. *J. Endocrinol.*, 1983; 99: 335-344.
15. Kreiner, D., Liu, H.C., Itskovitz, J., Veeck, L., Rosenwaks, Z.: Follicular fluid oestradiol and progesterone are markers of preovulatory oocyte quality. *Fertil. Steril.*, 1987; 48: 991-997.
16. Enien, W.M., El Sahwy, S., Harris, C.P., Seif, M.W., Elstein, M.: Human chorionic gonadotrophin and steroid concentrations in follicular fluid: the relationship to oocyte maturity and fertilization rates in stimulated and natural in vitro fertilization cycles. *Hum. Reprod.*, 1995; 10: 2840-2844.
17. Messinis, I.E., Templeton, A.A.: Relationship between intrafollicular levels of prolactin and sex steroids and in vitro fertilization of human oocytes. *Hum. Reprod.*, 1987; 2: 607-609.
18. Stone, B.A., Vargyas, J.P.M., Mars, R.P., Quinn, P.J., Batzofin, J.H., Tan, T., Kerin, J.F.P., Serafini, P.C.: Levels of steroids and protein hormones in antral fluids of women treated with different combinations of gonadotrophins, clomiphene citrate and a gonadotrophin-releasing hormone analog. *Fertil. Steril.*, 1988; 49: 249-257.
19. Tavmergen, E., Tavmergen, E.N., Capanoglu, R.: Do analogues of gonadotrophin releasing hormone influence follicular fluid steroids, oocyte quality and fertilization rates? *Hum. Reprod.*, 1992; 7: 479-482.
20. Gündüz, M.C., Turna, O., Apaydın, S., Uçmak, M., Kaşıkçı G., Ates, A., Esen, F., Evkuran, G., Töre D.: Potential relevance of insulin-like growth factor (IGF-I), total estradiol and progesterone to the in vitro maturation of equine oocytes. *Anim. Reprod. Sci.*, 2010; 121S: 245-246.
21. Kawano, Y., Hasan, K.Z., Fukuda, J., Mine, S., Miyakawa, I.: Production of vascular endothelial growth factor and angiogenic factor in human follicular fluid. *Molecular and Cellular Endocrinology*, 2003; 202: 19-23.
22. Malamitsi-Puchner, A., Sarandakou, A., Baka, S.G., Tziotis, J., Rizos, D., Hassiakos, D., Creatsas, G.: Concentrations of angiogenic factors in follicular fluid and oocyte-cumulus complex culture medium from women undergoing in vitro fertilization: association with oocyte maturity and fertilization. *Fertil. Steril.*, 2000; 76: 98-1011.