Abstract: Aims: Rates of certain histopathologic features were evaluated in hepatitis C and in hepatitis B materials and analyzed statistically in a Turkish population.

Method: The presence of lymphoid aggregate, steatosis, ductal lesions, sinusoidal inflammation and portal iron deposition were investigated in 80 hepatitis C and 104 hepatitis B specimens and evaluated using the chi-square test and Fisher’s exact test. Pan B and T cell markers were used for lymphoid aggregate analysis. P53 and PCNA were applied for the possible impact of steatosis on cell biology.

Results: All mentioned parameters excluding sinusoidal inflammation were found to be higher in hepatitis C samples. The co-presence of lymphoid aggregate and ductal lesions was found to be significant in hepatitis C. The distribution of T and B cells was similar in both groups. P53 and PCNA expressions were found to be non-significant in both groups.

Conclusion: Our results except sinusoidal inflammation were thought to be in correlation with the reports in the literature. Lower rates of iron deposition might be related to geographic differences that could be observed as a feature of the C virus.

Key Words: Hepatitis B virus, Hepatitis C virus, histopathology, P53, PCNA

Introduction

Chronic hepatitis is not a single disease, but rather a clinical and pathological syndrome, which has several causes (1). Liver biopsy is of major importance in the evaluation of patients with chronic hepatitis. The histological features of chronic hepatitis have been studied extensively and certain characteristic morphological features have been ascribed to chronic hepatitis of differing origins.

Histopathological changes in chronic hepatitis C have been described in previous reports (2,3). A histologic pattern of mild chronic hepatitis with portal lymphoid aggregate and/or follicles, varying degrees of bile duct damage, lymphocytic infiltration of sinusoids and steatosis was found to be more frequent than other chronic hepatitis. However, in some chronic hepatitis B cases, similar histologic findings are seen (4).

In the present study, our aim was to determine the rate and significance of these features in hepatitis C by comparing their presence in hepatitis B. The intraportal lymphoid cell population in both groups was analyzed.

The presence of viral markers of each type were investigated immunohistochemically in order to determine their significance and reliability in the diagnosis. The samples with steatosis were stained with anti-PCNA and anti-P53 for their possible relation with carcinogenesis (5).

Materials and Methods

Needle biopsy samples fixed in 10% neutral formalin belonging to 80 hepatitis C and 104 hepatitis B cases were included. The patients in the first group were all seropositive for antiHCV, whereas the second group’s patients had HBs antigen in their sera. In addition to the routine H&E stain, various other stains like periodic acid Schiff (PAS) with and without diastase digestion for bile duct injury, Masson’s trichrome and reticuline for fibroblastic stage and Prussian blue for iron deposition were performed.

Five histological features were defined and evaluated as follows:
1. Lymphoid aggregates/follicles in portal tracts: Follicles consisting of a germinal center with surrounding small lymphocytes or a densely packed collection of small lymphocytes within a portal tract, typically near an interlobular bile duct, were assessed (Figure 1).

2. Bile duct damage (Figure 1): Bile duct damage is characterized by a lymphocytic infiltrate surrounding the interlobular bile ducts. These inflammatory cells may be found between the epithelial cells or migrate through the basement membrane. Beyond this inflammation, loss of polarity, variations in nuclear chromaticity, cytoplasmic vacuolization and mitotic activity in epithelial cells are part of the damage.

3. Fatty change (Figure 2): Large and small droplet vacuoles were defined as absent, mild, moderate and marked.

4. Activation of sinusoidal inflammatory cells (Figure 3): The prominence of lymphocytes and Kupffer cells in sinusoids, in a “beads-on-a-string” pattern, was evaluated as present or absent.

5. Iron deposition (Figure 4): Only portal iron depositions in macrophages and especially in the venous endothelial cells were noted as present or absent.

Immunohistochemical studies.

The presence of viral antigens in the liver was shown by using antiHBs (Dako-Copenhagen, Denmark), antiHBc (Dako-Copenhagen, Denmark) for the B virus and antiHCV (Signet-Dedham, USA) for the C virus. The phenotype of immunocompetent cells present in and around intraportal lymphoid nodules was analyzed with pan T (CD43-DAKO-Copenhagen, Denmark) and pan B (CD20-DAKO-Copenhagen, Denmark) markers.

Thirty-one hepatitis B and 40 hepatitis C materials having steatosis were stained with antiP53 (DAKO, Copenhagen-Denmark) and antiPCNA (Biogenex-California, USA).

After deparaffinization and hydration steps, 4-micrometer thick sections were stained by the streptavidin-biotin peroxidase indirect technique. Diaminobenzidine (DAB) or AEC was used as the chromogen.

Statistical analysis

The results were evaluated with Chi-square and Fisher’s exact tests. Probability values less than 0.05 were considered significant.

Results

The frequencies of each histologic feature in biopsy specimens from HCV and HBV groups are shown in Table 1. Activation of sinusoidal cells, reflecting the lobular inflammatory component of chronic hepatitis, was observed in 18 (22.5%) of 80 HCV cases and 45 (45%) of 104 HBV cases. The higher incidence in B hepatitis was found to be significant (p<0.05).
Other histologic features, namely, steatosis, bile duct injury, lymphoid aggregate formation and portal iron deposition in HCV materials were observed at the following rates respectively: 61 (75%), 36 (45%), 46 (60%) and 12 (15%). We observed steatosis in 62 (60%), bile duct injury in 28 (27%), lymphoid aggregate formation in 27 (24%) and portal iron deposition in 7 (5%) of 104 hepatitis B samples.

On statistical analysis, steatosis, bile duct injury and lymphoid aggregate formation were found to be significantly higher in hepatitis C than hepatitis B (p<0.05). Although portal iron deposition was seen more frequently in C cases, the difference was not significant.

Immunohistochemically, lymphoid aggregates showed similar cellular populations in both hepatitis C and B samples. While portal inflammation and the peripheral zone of lymphoid aggregate were stained with T cell marker prominently, B cell dominancy was observed in the middle of the aggregate (Figures 5 and 6).

Immunohistochemically, the viral markers were shown in 96 (95%) of 104 hepatitis B samples. While all 96 cases were positive with antiHBs, only 44 of 96 were stained with antiHBc. Detection of antiHCV was as low as 22, only 27.5% of all hepatitis C cases (Table 1).

The possible impact of steatosis on molecular carcinogenesis was examined using antiP53 and antiPCNA stains. Forty hepatitis C and 31 hepatitis B samples all having certain amount of steatosis were chosen for this purpose. Only nuclear staining was noted for both antibodies. P53 expression was seen in 4 of (6%) the
hepatitis C and 3 of (4%) the hepatitis B cases. PCNA staining (Figure 7) was shown in 6 of (8%) the hepatitis B and 17 (24%) of the hepatitis C cases (Table 2). There was no significant difference between the two groups for P53 and PCNA staining.

Discussion

Some chronic hepatitis types have certain characteristic histologic features (1). Although no pathognomonic cellular marker of HCV infection has been shown to date, a number of histological findings have been cited for their potential diagnostic value, including bile duct damage, lymphoid follicles and/or aggregates within portal tracts, fatty change and sinusoidal reaction. In the present study, the frequency of these lesions in 80 hepatitis C and 104 hepatitis B liver biopsy specimens was examined to determine if any of these features distinguishes HCV from HBV infection.

Although the above-mentioned lesions were observed in both groups, the best set of histological lesions, three features statistically more likely to be associated with hepatitis C, were lymphoid aggregate formation (HCV 60%, HBV 24%), bile duct damage (HCV 45%, HBV 27%) and steatosis (HCV 75%, HBV 60%). These lesions have also been described as histologic markers of chronic hepatitis C in various studies (2-4).

The hepatitis C virus is related to flaviviruses and pestiviruses and cell damage has been attributed to its direct cytopathic effect (6). Fatty change appears to fall into this category, and although it may be present in hepatitis B cases, it seems to be more severe and extensive in hepatitis C materials.

In our previous study, we detected notable rates of P53 overexpression in the materials of steatohepatitis and some nonviral chronic hepatitis materials with steatosis (7). In contrast, in the present study, very few cases of both viral hepatitis groups were stained positively with antiP53 and there were no significant difference between them (Table 2). Steatosis in viral hepatitis may be a different cytological alteration, which does display itself as a mutagenic phenotype but just as an innocent viral effect (8-10). Proliferating cell nuclear antigen (PCNA) expression was higher in hepatitis C materials than in hepatitis B materials. However, the positive cell rate was hardly more than 5% of all hepatocytes and this finding was in the normal ranges of the positivity for PCNA in the liver. Nuclear pleomorphism, which is not very uncommon in liver biopsies, seems to affect the positive staining rates (7,11).

Intraportal lymphoid aggregates and/or follicles are located close to interlobular bile ducts or surround them in both hepatitis C and hepatitis B materials. In the pathogenesis of these lesions, immunologically mediated reactions are suggested (12,13).
Improvement of these lesions after interferon a
treatment has been reported and seems to reflect
immune pathogenesis (14). The aggregates showed
similar cellular population in both groups and while portal
and periportal cells were positive with CD43, B cell
dominancy was observed in the middle of the aggregates.
Our immunophenotyping findings were similar to those in
previous studies in which no differences have been
reported between chronic hepatitis C or B and
autoimmune hepatitis (12-14). In this context, they seem
to be true functional lymphoid follicles resembling lymph
nodes (14). In a recent study the authors focused on B
cell response and extensively analysed B cell activation,
proliferation and maturation in the livers of patients with
chronic hepatitis C (14). They found similar lymphoid
marker expressions with lymph nodes and concluded that
the liver is a potential secondary lymphoid organ. The
reason why HCV infection preferentially induces the
formation of functional B cell response in the liver is
currently unclear. However, our results, in accordance
with those in the literature, suggest that lymphoid
aggregates/follicles are not unique but are a characteristic
feature of chronic hepatitis C. Intraportal lymphoid
aggregate formations and bile duct injury can be seen in
some other nonviral liver diseases like autoimmune
hepatitis and primary biliary cirrhosis (15). None of the
patients in either group had serologic immune markers,
and no histological features of autoimmune hepatitis like
plasma cell dominancy, periportal liver cell rosettes or
severe necroinflammatory reaction were observed.

The duct damage was not severe and did not seem to
end with destruction in our materials. Since it is always
located close to lymphoid reaction and bile duct
epithelium expresses histocompatibility (HLA) antigens, the
presence of a lesion appears to be related with immune
reaction. Because of the higher frequency of bile duct
injury in hepatitis C than in hepatitis B, we assumed that
this lesion was another characteristic feature of viral
hepatitis C.

During the past few years, there has been much
interest in the role of iron in viral hepatitis (16). Iron has
long been known to be an essential element for the
replication of all organisms, including virulent
microorganisms. Stainable iron is found commonly in
liver biopsy specimens of patients with acute or chronic
viral hepatitis. Hemosiderin and lipofuchsins were defined
in Kupffer cells and this finding was related to repair
after injury (17). The implication was that the iron found
in the liver in viral hepatitis had been released from
hepatocytes damaged by the virus.

Farinatia et al. addressed the question of whether HCV
may have a direct cytopathic effect on hepatocytes
through the activation of iron dependent lipid
peroxidation (18). Their results were interpreted as
suggesting that mechanisms of hepatocyte damage are
different in hepatitis B and C and that the altered iron
metabolism and iron accumulation in hepatitis may be
related to an effect of the virus itself on hepatocytes or
possibly on the immune cells of the liver. In our study,
iron deposition was observed more frequently in hepatitis
C than hepatitis B but the difference was not statistically
significant. The localization of iron deposition was similar
but deposition in portal macrophages and venous
endothelial cells was noted more in hepatitis C materials.
Barton et al. were first to stress the importance of portal
distribution of a hepatic iron as a predictor of poor
response to IFNa (19). It was speculated that iron in
portal endothelial cells could interfere with their functions
during the inflammatory process, such as clearance of the
virus, or could participate in immune mediated reaction
(20). However, they did not detect any significant
differences between the cases of hepatitis B, C or
autoimmune hepatitis according to iron deposition
localization (19).

The number of samples showing iron deposition in
our study was not as high as reported in the literature.
But there have been some reports about similar
observations from Israel (16).

We speculate that this could be due to differences in
the type and geographic distribution of the virus, which is
a very well known feature of the C virus. Thus further
investigations about serum iron and iron binding protein
analyses in correlation with different viral genotypes,
which are beyond the scope of this study, may be needed.

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References


