

Highly stereoselective and efficient synthesis of the dopa analogue in pepticcinnamin E via enantioselective hydrogenation of dehydroamino acids

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An efficient and new method was developed to prepare the dopa analogue **11** in natural pepticcinnamin via catalytic hydrogenation of dehydroamino acids (DDAA) with a good yield and *ee*. Product **11** is a key intermediate towards the total synthesis of pepticcinnamin E and its analogues.

Key Words: Synthesis; dopa analogue; enantioselective hydrogenation; dehydroamino acid.

Introduction

Pepticcinnamin E¹ (Figure) is a major product of the pepticcinnamins, which are isolated from the culture of *Streptomyces* sp. OH-4652. It was identified as a depsipeptide having an *O* – *Z*-pentenylcinnamin acid and a novel dopa analogue, whose configuration has been determined as *S* by the Waldmann group using the Schöllkopf method.² Pepticcinnamin E shows rather potent inhibitory activity against farnesyl protein transferase (FPTase) with an IC₅₀ of 0.3 μM and is the first competitive inhibitor derived from a natural product. Our interest in the exploitation of a new methodology to synthesise naturally bioactive peptides containing non-ribosomal amino acids led us to initiate the synthesis of pepticcinnamin E (**1**). Emphasis was placed on preparing both dopa analogue **2** and *O* – *Z*-pentenylcinnamin acid.³ This study reports the enantioselective synthesis of the precursor of the dopa analogue **2**, which would be suitable for the total synthesis of pepticcinnamin E and its analogues.

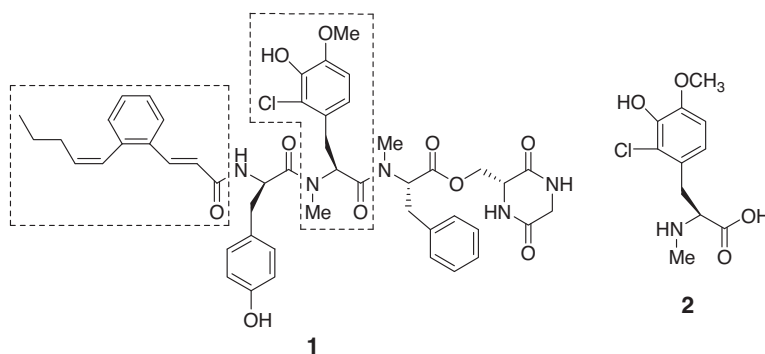


Figure. Structures of pepticinnamin E (**1**) and the dopa analogue **2**.

Experimental section

General. Melting points were determined with an Electrothermol digital melting point apparatus, and were uncorrected. Optical rotation was recorded on a Perkin-Elmer Model 341 polarimeter, at the sodium D line. Elemental analysis was undertaken on a Carlo-1106 model automatic instrument. Infrared spectra (IR) were run on a Nicolet MX-1 and Nicolet-560 MAGNA. ¹H-NMR and spectra were run either on a Bruker-200 and Bruker-300 or on a Varian-400. ¹³C-NMR was given by a Bruker-200. MS-EI mass spectra were obtained on a VG 7070E.

2-Chloro-3-hydroxy-4-methoxybenzaldehyde (6): A solution of compound **5** (9.35 g, 61.0 mmol) in freshly distilled CH₂Cl₂ (260 mL) was bubbled into gas Cl₂ until the yellow colour disappeared. After stirring at RT for 1 h, the white solid was collected by filtration and washed with warm CH₂Cl₂ to give compound **6**, 8.0 g, yield 71%, mp 207~208 °C. FT-IR (KBr): 3209, 3199, 1662, 1590, 1490, 1286, 1248, 1213, 1030, 810, 670 cm⁻¹. ¹H-NMR (200 MHz, DMSO-d₆)δ = 10.3 (s, 1H, CHO), 7.57 (d, *J* = 8.0 Hz, 1H, Ar-H), 6.90 (d, *J* = 8 Hz, 1H, Ar-H), 3.98 (s, 3H, OCH₃); MS-EI (*m/z*): 187 (M⁺), peak of isotope for Cl: 185/187 = 3:1, 187/189 = 3:1. Anal. Calcul. for C₈H₇ClO₃: C 51.49, H 3.78, Found: C 51.41, H 3.83.

2-Chloro-3-acetyloxy-4-methoxybenzaldehyde (7): To a solution of compound **6** (1.94 g, 10.4 mmol) in dry pyridine (25 mL) was added dropwise the acetyl chloride (0.75 mL, 10.5 mmol) at 0 °C. After stirring at RT for another 2 h, the reaction solution was diluted with ethyl acetate, and then washed with cold 1N H₂SO₄ and water (until pH 7). The organic layer was dried over anhydrous MgSO₄, and concentrated to give a crude product, which was purified by flash column chromatography on silica gel (with petr./ethyl acetate 6:1, 4:1 as eluent) to obtain white solid **7**, 2.12 g, yield 89.1%, mp 64~65 °C. FT-IR (KBr): 3082, 3012, 2883, 1778, 1685, 1594, 1496, 1435, 1370, 1298, 1256, 1190, 1167, 1037, 891, 818 cm⁻¹. ¹H-NMR (200 MHz, CDCl₃)δ = 10.32 (s, 1H, CHO), 7.88 (d, *J* = 8.8 Hz, 1H, Ar-H), 7.0 (d, *J* = 8.8 Hz, 1H, Ar-H), 3.93 (s, 3H, OCH₃), 2.41 (s, 3H, COCH₃). Anal. Calcul. for C₁₀H₉ClO₄: C 52.53, H 3.97, Found: C 52.47, H 4.02

(Z)-Methyl3-(3-acetyloxy-2-chloro-4-methoxyphenyl)-2-(tert-butyloxycarbonyl) acrylate (8): To a solution of methyl *N*-protected-2-(diethoxycarbonyl)phosphinyl-glycinate (2.2 mmol) in dry CH₂Cl₂ (6 mL) was added DBU (0.3 mL, 2.2 mmol) at 0 °C; after 1 min, the solution of compound **7** (2.0 mmol) in dry CH₂Cl₂ (6 mL) was added. After stirring at 0 °C for another 2 h, the reaction solution was diluted with 40

mL of ethyl acetate and then washed with cold 1 N H₂SO₄ (5 mL). The organic layer was dried over anhydrous Na₂SO₄, and concentrated to give a crude product, which was purified by column chromatography on silica gel (with petr./ethyl acetate as eluent) to obtain white solid **8**, 0.78 g, yield 98%, mp 126-127 °C. FT-IR (KBr): 3526, 3349, 2980, 2949, 1782, 1720, 1642, 1601, 1494, 1440, 1371, 1337, 1302, 1247, 1174, 1034, 880, 772 cm⁻¹; ¹H-NMR (200 MHz, CDCl₃)δ = 7.56 (d, *J* = 8.91 Hz, Ar-H), 7.35 (s, 1H, = CH), 6.88 (d, *J* = 8.91 Hz, Ar-H), 6.17 (bs, 1H, NH), 3.87 (s, 3H, ArOCH₃), 3.86 (s, 3H, COOMe), 2.38 (s, 3H, COMe), 1.41 (s, 9H, Boc). MS-EI (*m/z*): 399 (M⁺); Anal. Calcul. for C₁₈H₂₂ClNO₇: C 54.52, H 5.57, N 3.46. Found: C 54.34, H 5.51, N 3.51.

(*S*)-*N*-tert-butyloxycarbonyl-(3-acetyloxy-2-chloro-4-methoxy)-phenylalanine methyl ester (9**):** To a solution of DIPAMP (2.7 mg, 0.0059 mmol) in 1.5 mL of absolute acetone (deoxygenation before use) was added [Rh(COD)BF₄] (2.4 mg, 0.0059 mmol) under Ar₂. After stirring at RT for 1 h, this catalyst with a concentration of 0.0004 mmol/0.1 mL was prepared and used in subsequent procedure right away. To a solution of compound **8** (0.70 g, 1.75 mmol) in absolute acetone (26 mL) (deoxygenation before use) was added the catalyst prepared in the above procedure. The reaction solution was hydrogenated under 1 atm for 42 h at RT. Active carbon was then added by stirring. After 30 min, the solid was filtered off through a Celite pad and the filtrate was concentrated to give a crude product, 0.70 g, with a 90.6% *ee* value and *S* configuration (determined by chiralpak OD, Hexane:iPrOH/90:10, rate: 1 mL/min). The crude product was twice recrystallised from ethyl acetate and hexane to give a white needle solid **9**, 0.62 g with >99% *ee* value and 89.1% yield, mp 108~109 °C. [α]_D²⁸ +18 (*c* 0.35, CH₂Cl₂). FT-IR (KBr): 3368, 2980, 2942, 1774, 1753, 1686, 1606, 1514, 1441, 1370, 1286, 1207, 1171, 1042 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃)δ = 7.06 (d, *J* = 8.4 Hz, 1H, Ar-H), 6.82 (d, *J* = 8.4 Hz, 1H, Ar-H), 5.06 (bs, 1H, NH), 4.56 (m, 1H, CH), 3.82 (s, 3H, OCH₃), 3.68 (s, 3H, COOCH₃), 3.25-3.11 (m, 2H, β-CH₂), 2.37 (s, 3H, COCH₃), 1.37 (s, 9H, Boc). MS-EI (*m/z*): 401 (M⁺); Anal. Calcul. for C₁₈H₂₄ClNO₇: C 53.80, H 6.03, N 3.49. Found: C 53.88, H 5.96, N 3.45.

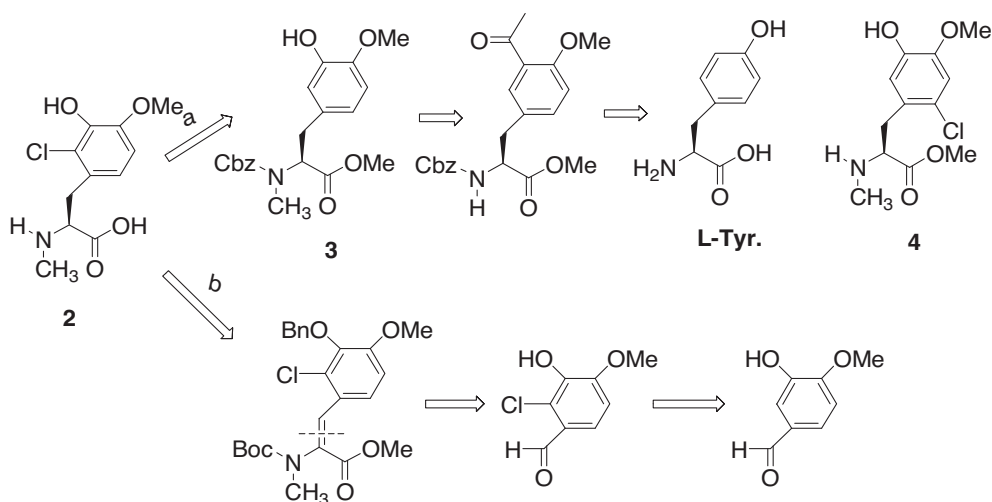
(*S*)-*N*-tert-butyloxycarbonyl-(3-benzyloxy-2-chloro-4-methoxy)-phenylalanine methyl ester (10**):** To a solution of compound **9** (0.10 g, 0.25 mmol) in CH₃OH (6 mL) and distilled water (2 mL) was added dropwise saturated NaHCO₃ (2 mL). The reaction solution was stirred at RT for 3 h and acidified to pH 3~4 at 0 °C, and then extracted with ethyl acetate 4 times. The combined ethyl acetate layer was washed with brine and dried over anhydrous MgSO₄, and then concentrated to give a slight yellow slurry, which was dissolved in freshly distilled DMF (5 mL) at once. The powdered anhydrous K₂CO₃ (86 mg, 0.63 mmol) was added, followed by addition of BnBr (36.0 μL, 0.30 mmol) at 0 °C. After stirring at RT for 3 h under N₂, cold water was added and extracted with ethyl acetate 4 times. The combined organic layer was then washed in cold 1 N KHSO₄ and brine and dried over anhydrous MgSO₄, and concentrated to give a slight slurry, which was purified by column chromatography on silica gel (with hexane/ethyl acetate as an eluent) to obtain compound **10**^{2a} as a slight yellow slurry, 63 mg, (23 mg of compound **9** was recovered), yield 73.7% for 2 steps (based on transformed starting material). [α]_D²⁸ +17.2 (*c* 0.25, CH₂Cl₂). ¹H-NMR (400 MHz, CDCl₃)δ (main rotamer) = 7.45 (m, 2H, Ar-H), 7.29 (m, 3H, Ar-H), 6.84 (d, *J* = 8.0 Hz, 1H, Ar-H), 6.71 (d, *J* = 8.0 Hz, 1H, Ar-H), 4.95 (s, 2H, OCH₂Ph), 4.52 (m, 1H, α-CH), 3.77 (s, 3H, OCH₃), 3.63 (s, 3H, COOCH₃), 3.16 (dd, ²*J* = 13.6 Hz, ³*J* = 6 Hz, 1H, β-CH₂), 3.01 (dd, ²*J* = 13.6 Hz, ³*J* = 6 Hz, 1H, β-CH₂), 1.33 (s, 9H, Boc). MS-EI (*m/z*): 449 (M⁺). Anal. Calcul. for C₂₃H₂₈ClNO₆: C 61.40, H 6.27, N 3.11. Found: C 61.37, H 3.18, N 3.20.

(*S*)-*N*-tert-butyloxycarbonyl-*N*-methyl-(3-benzyloxy-2-chloro-4-methoxy)-phenylalanine methyl ester (11**):** To a solution of compound **10** (0.10 g, 0.22 mmol) in THF (2.5 mL) was added a solution

of LiOH.H₂O (25.0 mg) in water (0.5 mL) at 0 °C. After stirring at RT for 3 h, cold water was added and acidified to pH 3~4 with 0.1 N HCl, and then extracted with ethyl acetate 4 times. The combined organic layer was washed with brine and dried over anhydrous Na₂SO₄, and concentrated to give a slight slurry, which was purified by column chromatography (petr./ethyl acetate: 3:1, 1:1, 1:2 as an eluent) to obtain the acid 81 mg (0.186 mmol), which was used for next *N*-methylation directly. The acid was dissolved in 4.5 mL of dry THF. To this solution was added NaH (22.4 mg, 0.56 mmol), followed by MeI (46 μL, 0.75 mmol) at 0 °C. The reaction suspension was then stirred at 27 °C for 16 h. Cold water was added and acidified to pH 3~4 with 0.1 N HCl, and then extracted with ethyl acetate 4 times. The combined organic layer was washed with brine and dried over anhydrous MgSO₄, and concentrated to give a crude product, which was purified by column chromatography on a silica gel (with petr./ethyl acetate: 3:1, 1:1, 1:2 as an eluent) to obtain a colourless oil **11**^{2a} 50 mg, yield 50%. $[\alpha]_D^{20}$ -85.9 (*c* 0.11, CH₂Cl₂). FT-IR (Neat): 3010, 2984, 2936, 2851, 1739, 1705, 1487, 1444, 1374, 1245, 1947, 758 cm⁻¹. ¹H-NMR (200 MHz, CDCl₃)δ (main rotamer) = 7.54 (m, 2H, Ar-H), 7.34 (m, 3H, Ar-H), 6.88 (d, 1H, *J* = 8.6 Hz, Ar-H), 6.77 (d, *J* = 8.6 Hz, 1H, Ar-H), 5.05 (s, 2H, OCH₂Ph), 4.63 (m, 2H, α-CH, COOH), 3.86 (s, 3H, OCH₃), 3.51-3.07 (m, 2H, β-CH₂), 2.70 (s, 3H, NCH₃), 1.43 (s, 9H, Boc). MS-EI (*m/z*): 449 (M⁺). Anal. Calcul. for C₂₃H₂₈ClNO₆: C 61.40, H 6.27, N 3.11. Found: C 61.31, H 6.35, N 3.17.

Results and discussion

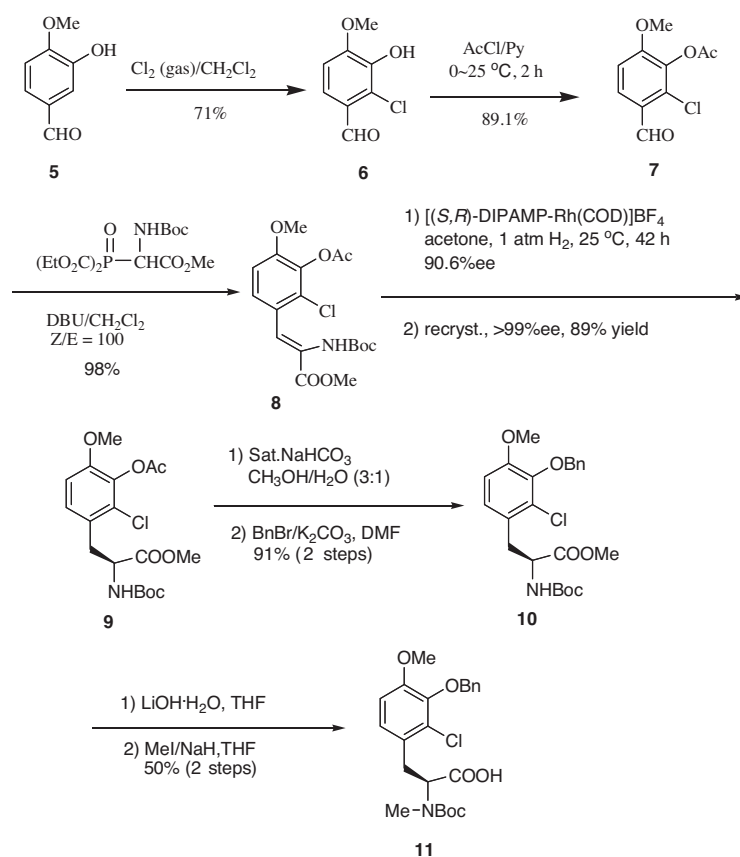
Many methods have been published for the preparation of enantiomerically pure dopa analogues, such as the improved synthesis of selectively protected *L*-dopa derivatives from *L*-tyrosine,⁴ the traditional Schöllkopf method for chiral amino acids, and enantioselective catalysis with chiral auxiliaries.⁵ We also wanted to obtain the dopa analogue **2** from *L*-tyrosine. As shown in the retro-synthetic route a (Scheme 1), compound **3** was obtained conveniently from *L*-tyrosine through acetylation, hydroxylation, and methylation under the Coggins condition.⁶ Unfortunately, the final chlorination of compound **3** was unsuccessful, maybe because of the steric hindrance effect. Even when Cbz was firstly removed before chlorination, and the undesired compound **4** was obtained.



Scheme 1. Retro-synthesis of the dopa analogue.

Another method (according to retro-synthetic route b; Scheme 1) was then developed with the aim of obtaining the target compound via enantioselective catalytic hydrogenation of the chlorinated dehydroamino acid by employing a chiral catalyst such as DIPAMP.⁷

Chlorination of isovanillin gave compound **6**, and protection of the OH group by acetylation afforded aldehyde **7**, which was coupled with *N*-protected-2-(diethoxycarbonyl)phosphinylglycine ester^{8,9} by using DBU as a base under the Horner–Emmons condensation condition⁸ to afford DDAA **8**. Subsequently, **8** has been hydrogenated using the chiral catalyst DIPAMP to give **9** with 100% conversion and 90.6% *ee*. In particular, this step can be conducted on a large scale with good *ee*. After recrystallisation, compound **9** was obtained in enantiopure form (>99% *ee*, Scheme 2).



Scheme 2. Asymmetric synthesis of the dopa analogue **11**.

Since the acetyl group was unstable under strong basic conditions (such as LiOH and NaH), it was selectively removed by treatment of **9** with a saturated solution of NaHCO₃ in MeOH/H₂O (Scheme 2) without removing the Me group;¹⁰ then the OH group was protected by benzylation to give the closely related known compound **10**.^{2a} Subsequently, under the Bnoiton condition,⁶ **10** has been transformed into the known compound **11**.^{2a}

Conclusion

A new synthetic route was developed to prepare the dopa analogue by employing the methodology of asymmetric catalytic hydrogenation of dehydroamino acids. Such a method is of practical use for the total synthesis of peptidocinnamin E and its analogues.

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